

**International Transporter Consortium (ITC)**  
**4<sup>th</sup> Workshop (virtual)**  
April 19-21, 2021



<https://www.itc-transporter.org/itcw4.html>  
provides links to agenda and registration at **ASCPT**

Pre-Workshop Webinar, March 31<sup>st</sup>, 2021, 5-6:30 PM EDT (**free registration**)  
Prof Yuichi Sugiyama

**Use of Extended Clearance Concept and PBPK Modeling in New Drug Discovery  
and Development: Predicting Target Tissue Exposure from In Vitro to In Vivo**

Fourth Workshop on  
Membrane Transporters in  
Drug Development

# Contents

- 1) Introduction;**  
**Rate-determining process (focusing on the liver)**  
**(Uptake, efflux, elimination, metabolism) DDI**
- 2) PGx of OATP1B1: PBPK model based analysis**
- 3-1) PBPK model based analysis of OATPs mediated drug-drug interaction**  
**(Top down + Bottom-up)**
  - (i) victim drugs-perpetrator drugs**
  - (ii) endogenous biomarker (CP-I) – rifampicin**
- 3-2) Simple bottom-up predictions dot not always work well.**
  - (i) prediction of hepatic clearance of highly protein bound drugs**  
**(albumin-mediated hepatic uptake mechanisms should be considered)**
  - (ii) Time-dependent inhibition (inhibitors for OATP1B and OCTs)**
- 4) Target-mediated drug disposition (TMDD); To obtain dose-dependent change in molecular target occupancies only from the plasma concentration time-profile**


# Drug-interaction between Cerivastatin and Gemfibrozil/CsA



日本では、重篤な副作用のためメーカー自主回収へ

薬事日報平成13年8月27日号

薬事日報平成13年8月15日号

WORLD HEALTH ORGANIZATION  ORGANISATION MONDIALE DE LA SANTE

QSM/MC/IEA.102 09 August 2001

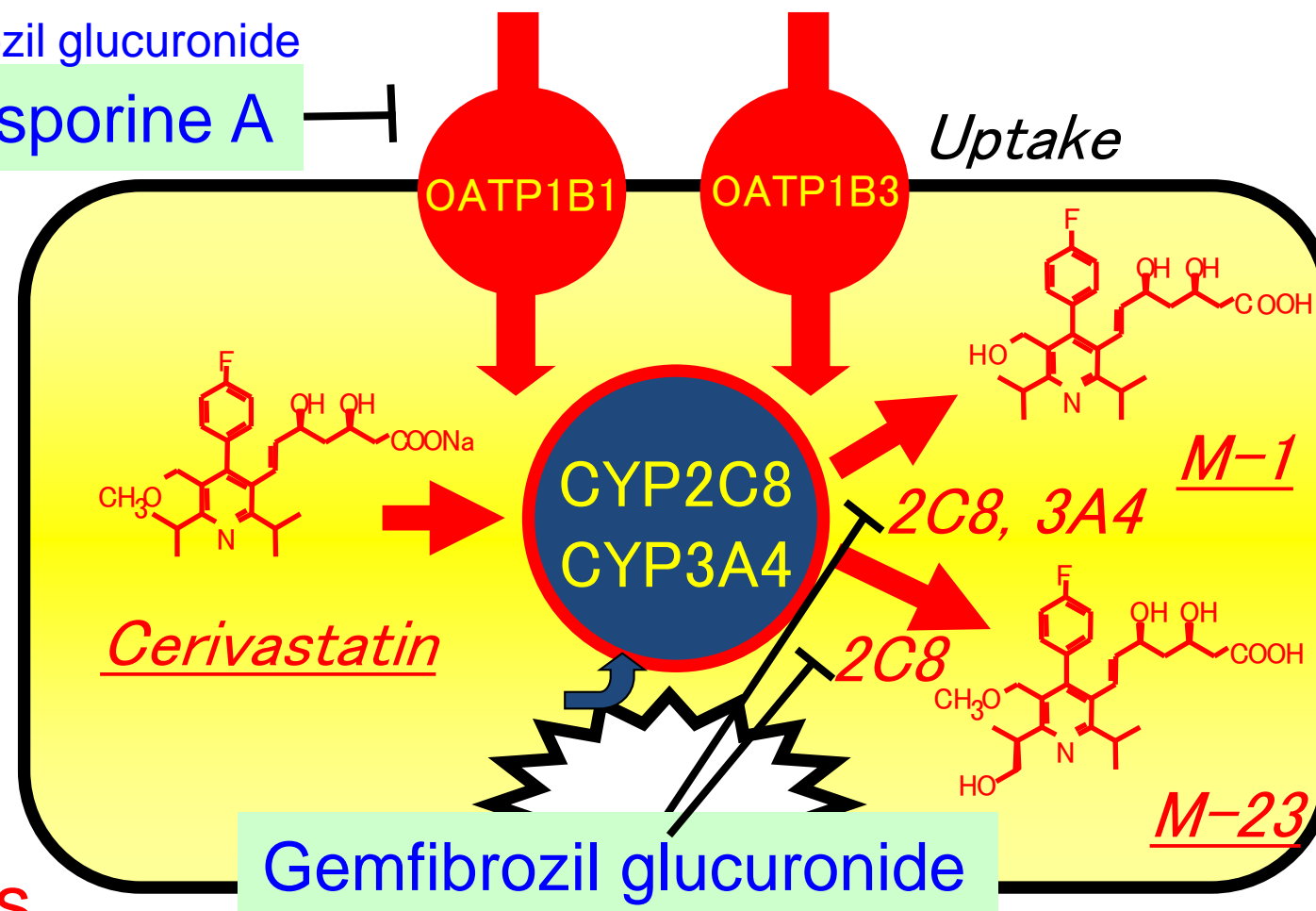
## Voluntary withdrawal of Cerivastatin – Reports of Rhabdomyolysis

Baycol (cerivastatin) was initially approved in the U.S. in 1997. It belongs to a group of cholesterol lowering drugs referred to as “statins”. While all statins could potentially cause this dangerous muscle reaction, rhabdomyolysis appears more frequent with cerivastatin, especially when used in high doses, in the elderly or, when taken along with gemfibrozil, another cholesterol lowering drug. In this connection it may be noted that Bayer has withdrawn all

- **52 patients died (US 31).**
- **Among 31 patients, 12 were given also gemfibrozil.**

Cerivastatin  
Dual substrates

Gemfibrozil-glucuronide  
Dual inhibitors



Shitara, Y. et al.  
J Pharmacol Exp Ther,  
304(2): 610-6 (2003)

Shitara, Y. et al.  
J Pharmacol Exp Ther,  
311(1): 228-36 (2004)

Shitara, Y. and Sugiyama Y.  
Pharmacol Ther,  
112(1): 71-105 (2006)

## Examples of substrates for uptake/efflux transporters and enzymes (1)

Substrates	Uptake transporter	Metabolic enzymes	Efflux transporter
Anti-Hyperlipidemic drugs (statins)			
atorvastatin	OATPs	CYP3A4	-
<b>cerivastatin</b>	OATPs	CYP2C8, 3A4	-
fluvastatin	OATPs	CYP2C9	-
pravastatin	OATPs	-	MRP2
rosuvastatin, pitavastatin	OATPs	-	BCRP
Anti-hypertension or -cardiovascular disease			
bosentan	OATPs	CYP3A4, 2C9	-
torasemide	OATPs	CYP2C9	-
telmisartan	OATP1B3	UGTs	-
valsartan	OATPs	-	MRP2
Anti-cancer drug			
docetaxel	OATP1B3	CYP3A4	-

## Examples of substrates for uptake/efflux transporters and enzymes (2)

Substrates	Uptake transporter	Metabolic enzymes	Efflux transporter
Anti-diabetes			
repaglinide	OATPs	CYP2C8, 3A4	-
nateglinide, glibenclamide	OATPs	CYP2C9, 3A4	
Anti-HCV			
simeprevir, grazoprevir	OATP1B1	CYP3A4	-
asunaprevir, danoprevir, paritaprevir	OATPs	CYP3A4	Pgp
Etc.			
Montelukast	OATP2B1	CYP2C8, 2C9, 3A4	-
maraviroc	OATP1B1	CYP3A4	Pgp
fexofenadine	OATPs	-	Pgp

**Understanding Extended Clearance Concept  
is very important (I have been training this concept  
to all the students , post-doc in my lab (UOT, RIKEN)**

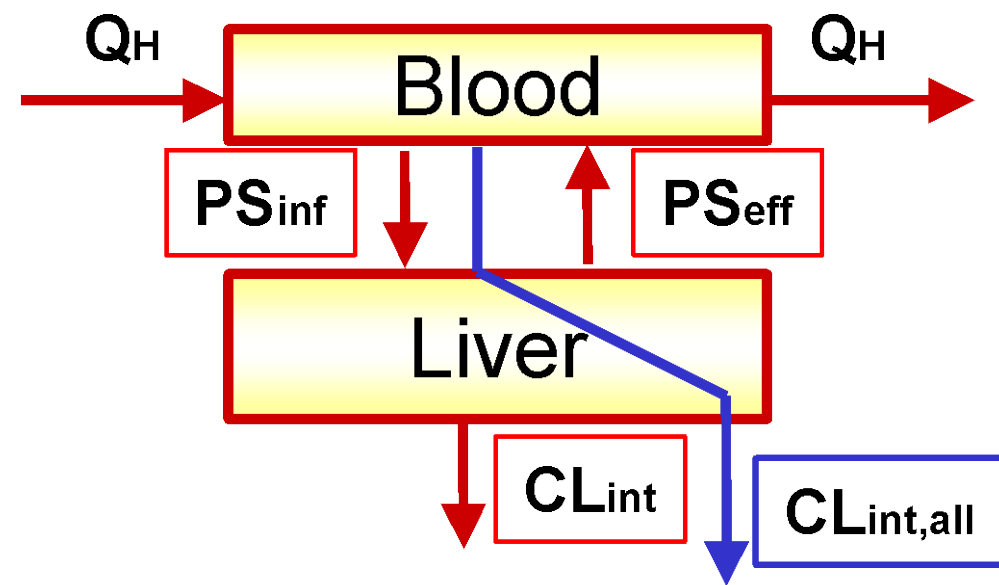
## “Extended Clearance Concept”

- enough to predict the change in **AUC and/or C<sub>ss</sub> both in plasma and tissue**  
(PG<sub>x</sub>, DDI (at least for static analyses,  
and not for dynamic analysis))

## “PBPK modeling”

- Appropriate model for describing **the drug conc-time course both in plasma and tissue as well as AUC, C<sub>ss</sub>**

# Overall Hepatic Intrinsic Clearance ( $CL_{int,all}$ )



Extended clearance concept

$$CL_{int,all} = PS_{inf} \times \frac{CL_{int}}{PS_{eff} + CL_{int}}$$

1) When  $PS_{eff} \ll CL_{int}$ , (Case-1)

$$CL_{int,all} = PS_{inf}$$

Uptake-limited (anionic drugs; statins, sartans)  
 $\beta = 1$

Degree of Sequestration ( $\beta$ )

2) When  $PS_{eff} \gg CL_{int}$ , (Case-2)

$$CL_{int,all} = \frac{PS_{inf}}{PS_{eff}} \times CL_{int}$$

Degree of Active Uptake ( $\alpha$ )

When  $PS_{inf} = PS_{eff}$  (passive diffusion),

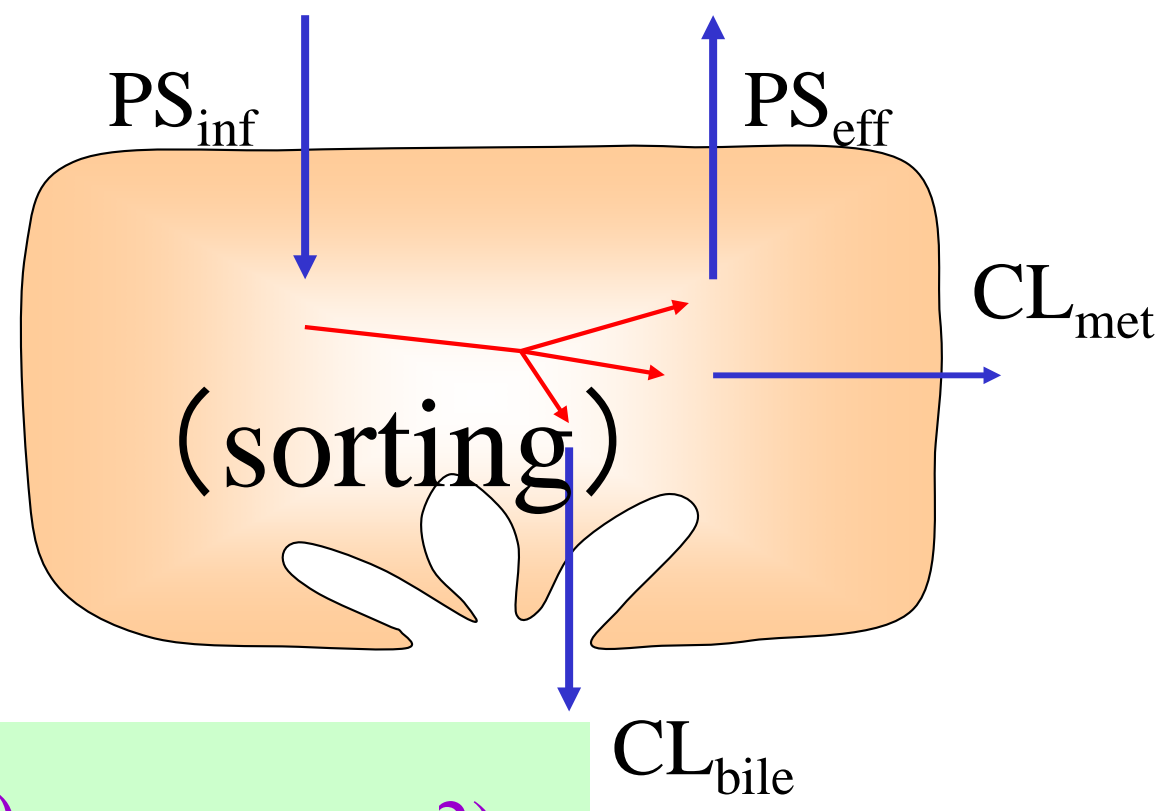
$$CL_{int,all} = CL_{int}$$

**Intrinsic Clearance-limited**  $\alpha = 1$

(lipophilic basic/neutral drugs; quinidine, diazepam)

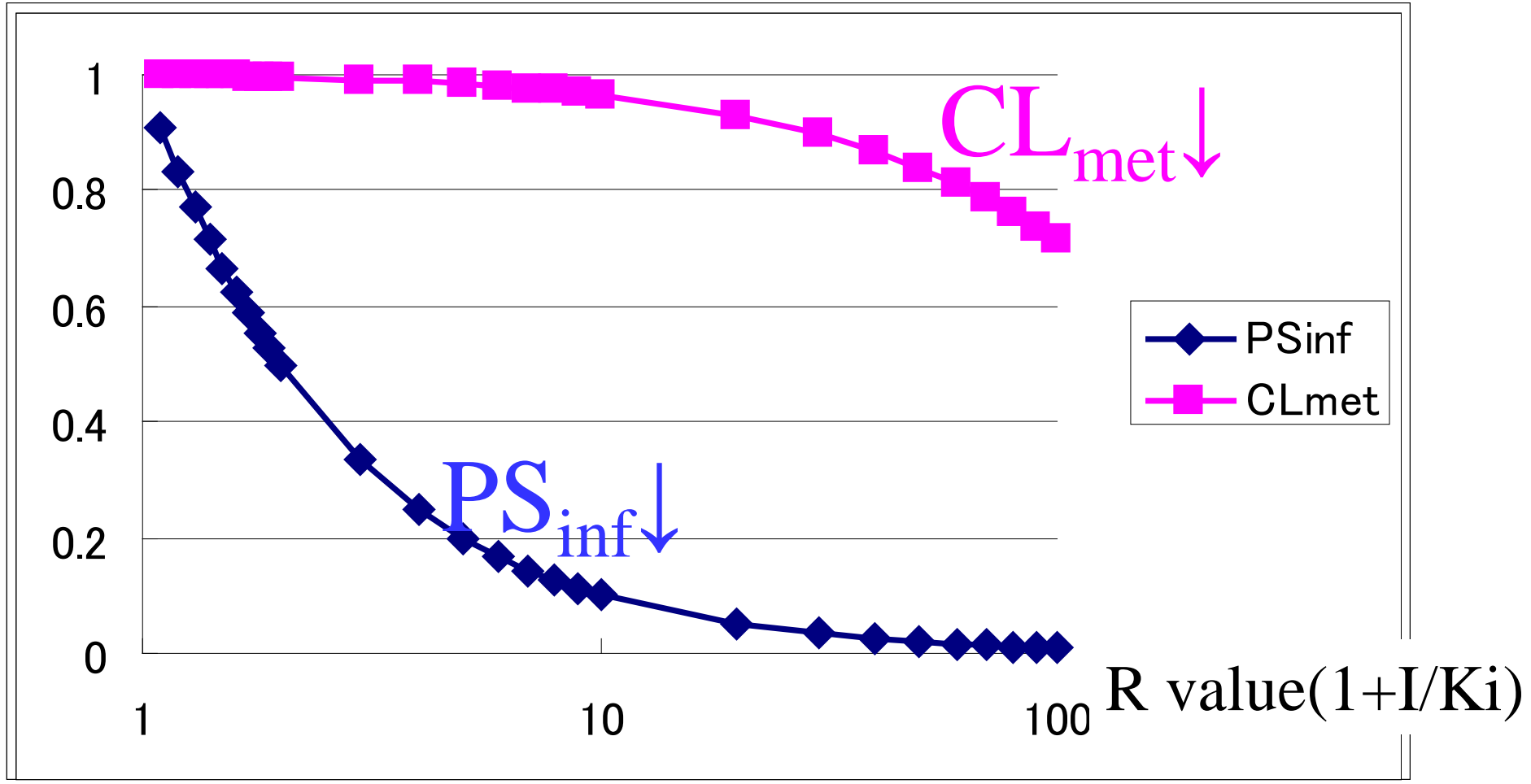
$$CL_{int,all} = PS_{inf} \times \frac{CL_{bile} + CL_{met}}{PS_{eff} + CL_{bile} + CL_{met}}$$

(sorting)



$PS_{inf} = 100, PS_{eff} = 2, CL_{bile} + CL_{met} = 500$  (Case-1) 2)

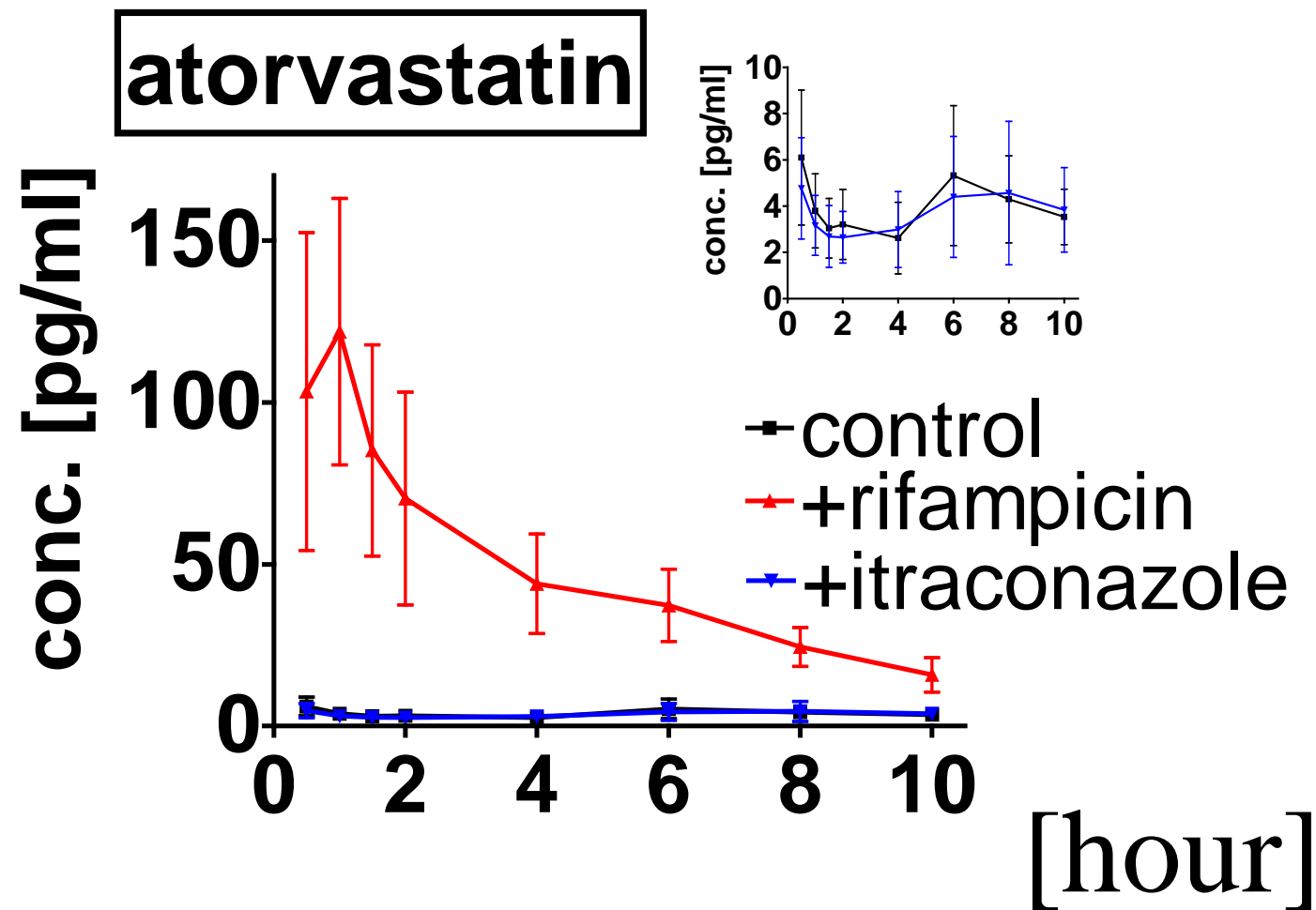
% of  $CL_{int,all}(cont.)$   
(1/AUC)



Impact of the function of each pathway on the overall intrinsic clearance



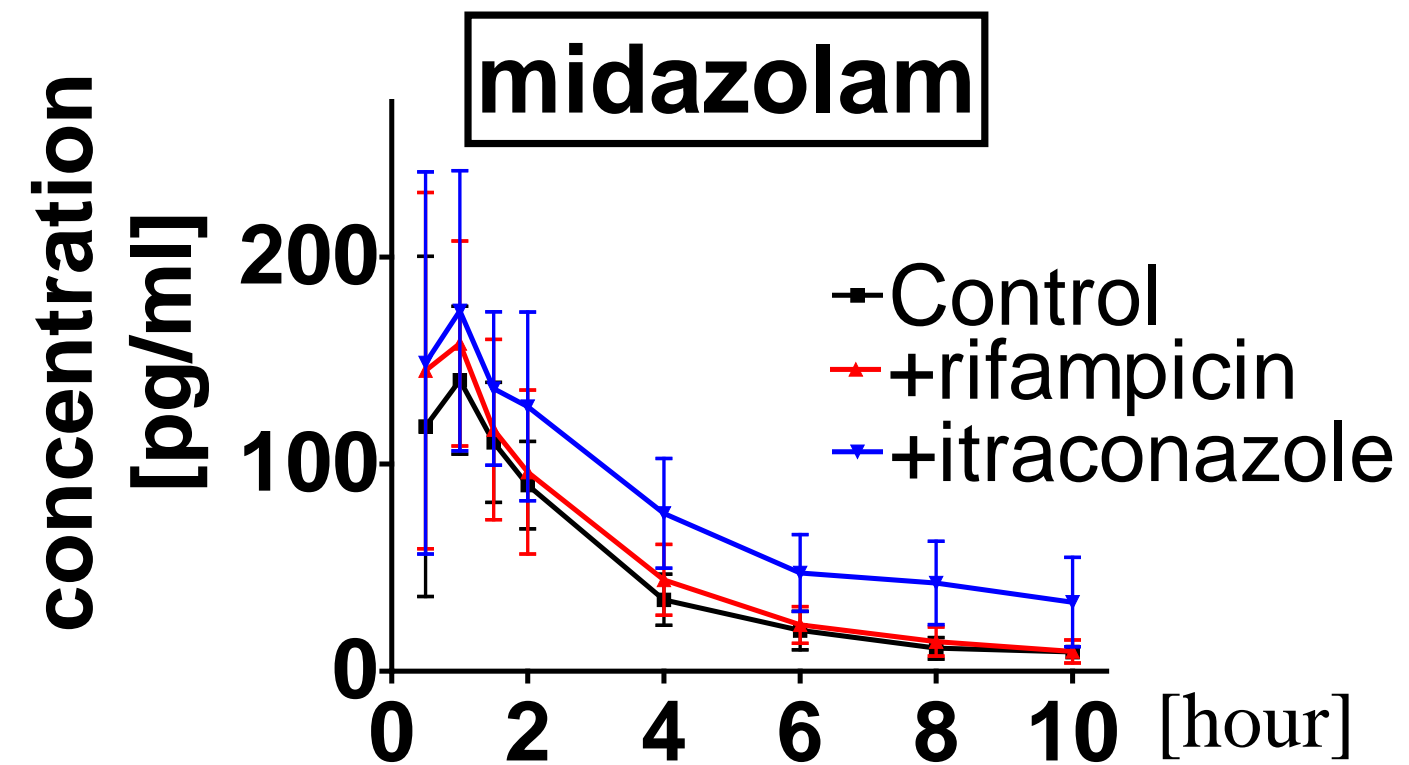
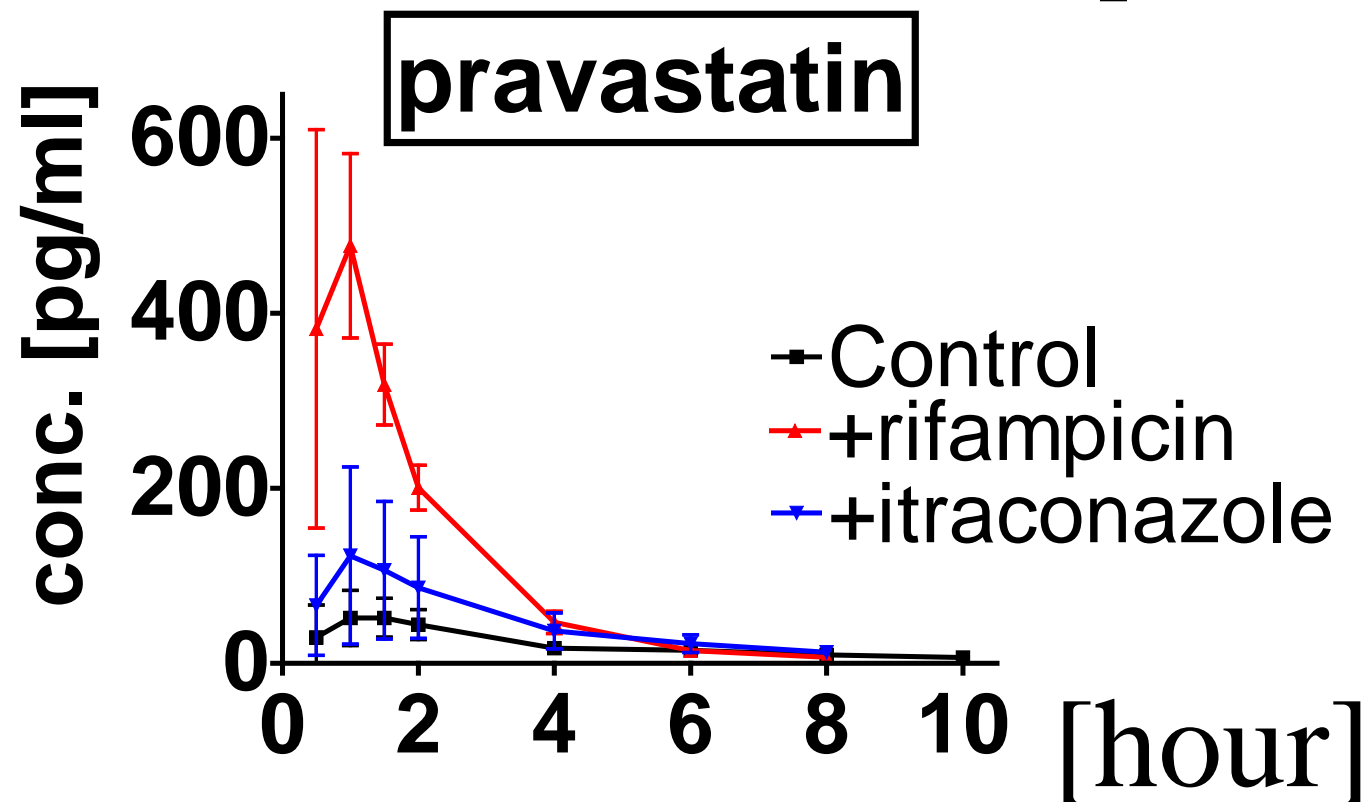
Plasma concentrations of atorvastatin and pravastatin were greatly increased by rifampicin, but not by itraconazole



AUC <sub>0-10</sub> [pg*hr/ml]	ATV	PRV (AUC <sub>0-8</sub> )	MDZ
Cont.	38.5 ±17.5	195 ±78.7	434 ±122
+RIF	439*** ±134	949*** ±179	471 ±168
+ITZ	36.0 ±19.2	386 ±254	755* ±276

\*\*\*: P<0.0005

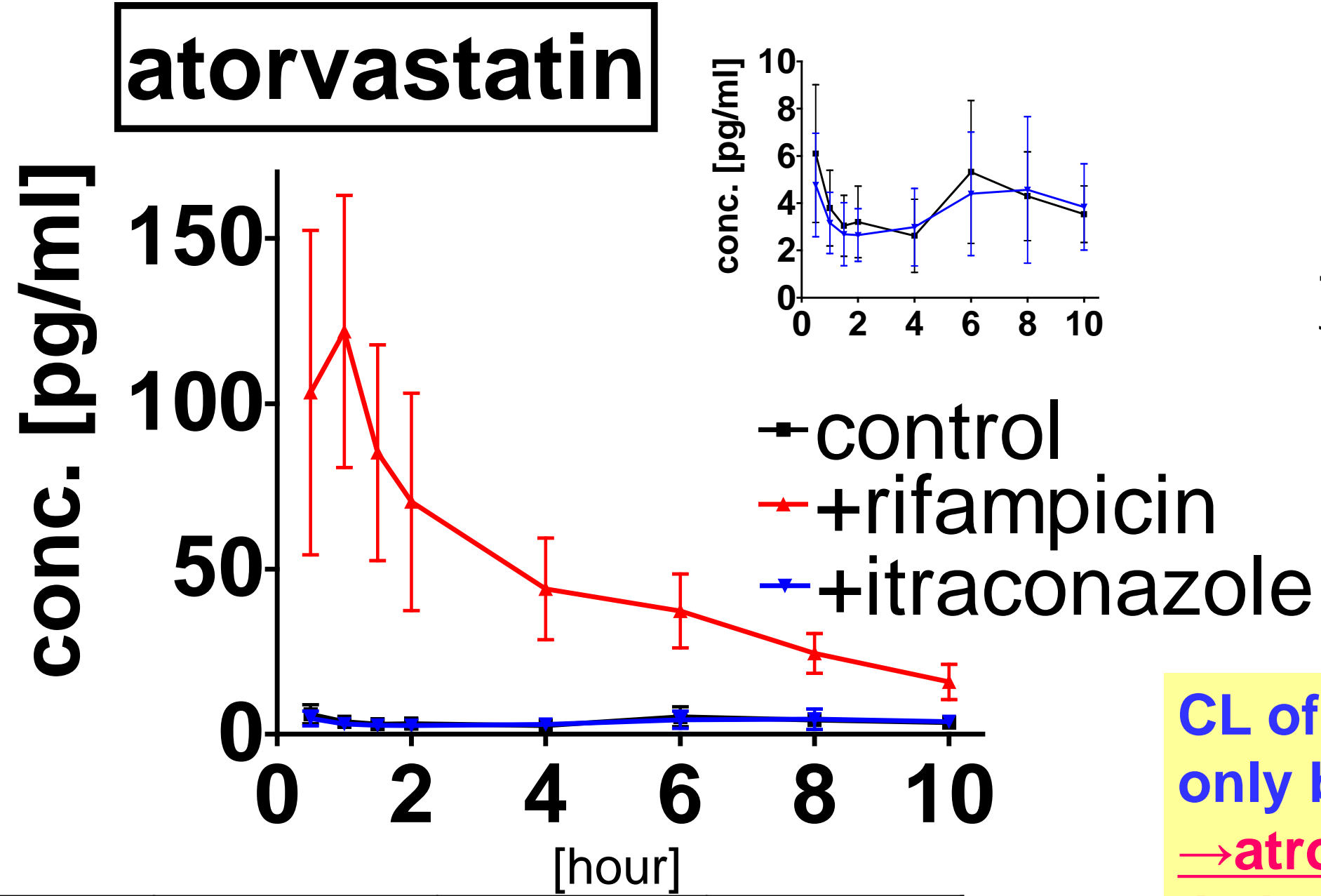
\*: P<0.05



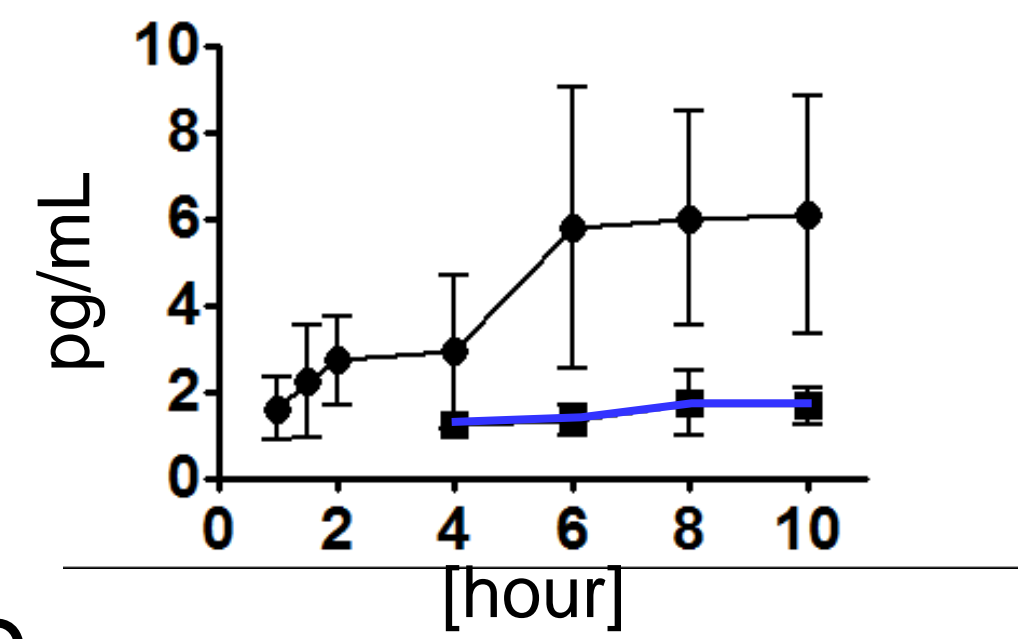
\*Doses of each substrates are 33µg

Maeda K et al., CPT (2011)

# Effects of rifampicin and itraconazole on the PK of atorvastatin



cf. 2-hydroxyatorvastatin



CL of atorvastatin was inhibited only by rifampicin.  
 → atorvastatin hepatic clearance is limited only by hepatic uptake

	Control	+RIF	+ITZ
AUC <sub>0-10</sub> [pg*hr/mL]	38.5 ±17.5	439*** ±134	36.0 ±19.2

Maeda K, Ikeda Y, Fujita T, Yoshida K, Azuma Y, Haruyama Y, Yamane N, Kumagai Y and Sugiyama Y. Identification of the rate-determining process in the hepatic clearance of atorvastatin in a clinical cassette microdosing study. Clin Pharmacol Ther 90:575-581 (2011).

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# The impact of OATP1B1 on the PK of drugs

~Plasma conc. of drugs is increased in subjects with OATP1B1\*15~

## HMG-CoA reductase inhibitors

pravastatin  
simvastatin acid  
pitavastatin  
atorvastatin  
rosuvastatin

## Anti-diabetes

repaglinide  
nateglinide  
glybenclamide

## Anti-allergic drug

fexofenadine

## Anti-pulmonary hypertension drug

Atrasentan  
bosentan

## Anti-cancer drug

irinotecan (SN-38)  
docetaxel, paclitaxel

## Chol-absorption inhibitor

ezetimibe

## Loop diuretics

Torasemide

## Angiotensin receptor antagonists

Olmесartan

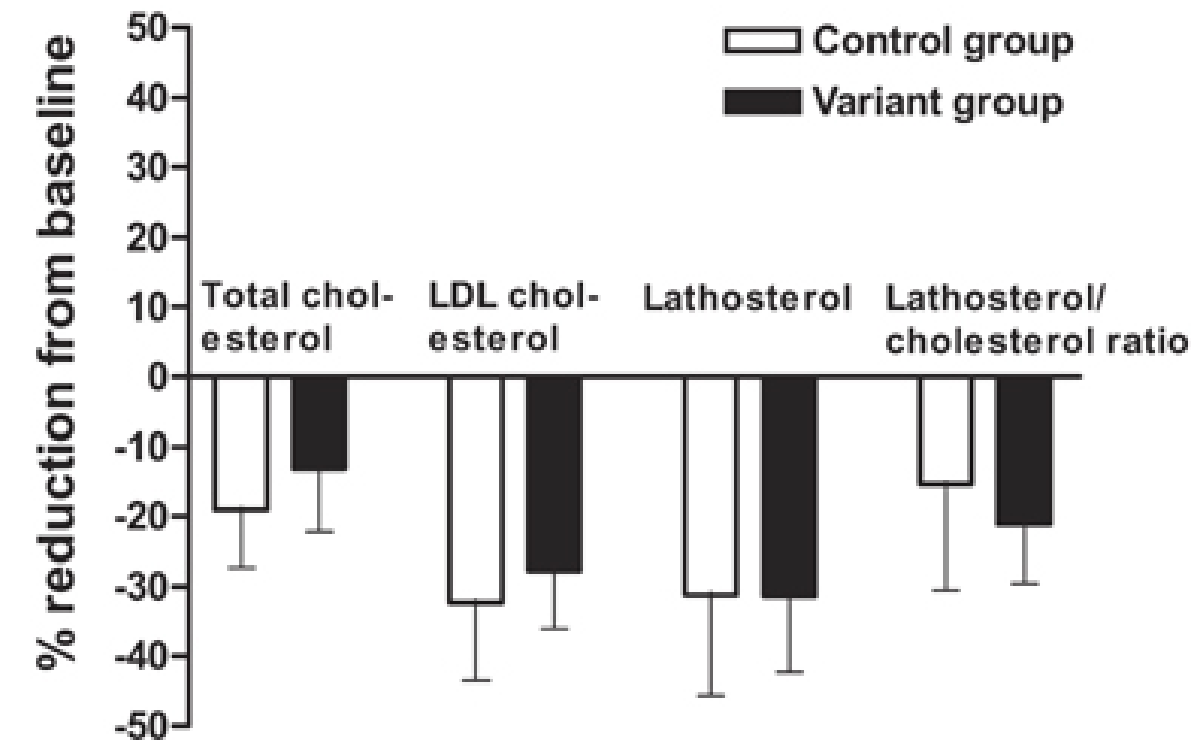
# Relationship between OATP1B1 genetic polymorphism and Pharmacological effect and adverse effect of statins

## ★ Pharmacological effect

Target; HMG CoA-reductase in the liver

No effect or small effect if any

(Igel M et al., Clin Pharmacol Ther, 79, 419-26 (2006))



## ★ Adverse Effect

Target; Muscle (via plasma)

Simvastatin-induced myopathy

strong correlation with OATP1B1 polymorphism

(SEARCH Collaborative Group et al., New Engl J Med, 359, 789-99 (2008))

Odds ratio of this SNPs for simvastatin-induced myopathy

521C/T vs T/T → 4.5 fold

521C/C vs T/T → 16.9 fold

# Summary

# All of these pharmacogenetic and DDI studies on OATP1B1 suggested that the hepatic uptake plays an important role in the plasma clearance of therapeutically important drugs (mostly anionic drugs; statins, ARA, ACE inhibitors, anti-HCV drugs, anticancer drugs, etc).

Why did this polymorphism and/or DDI affect **only side-effect (myopathy; muscle is a target tissue)**, and **not pharmacological effect (lipid lowering effect; liver is a target organ) ?**

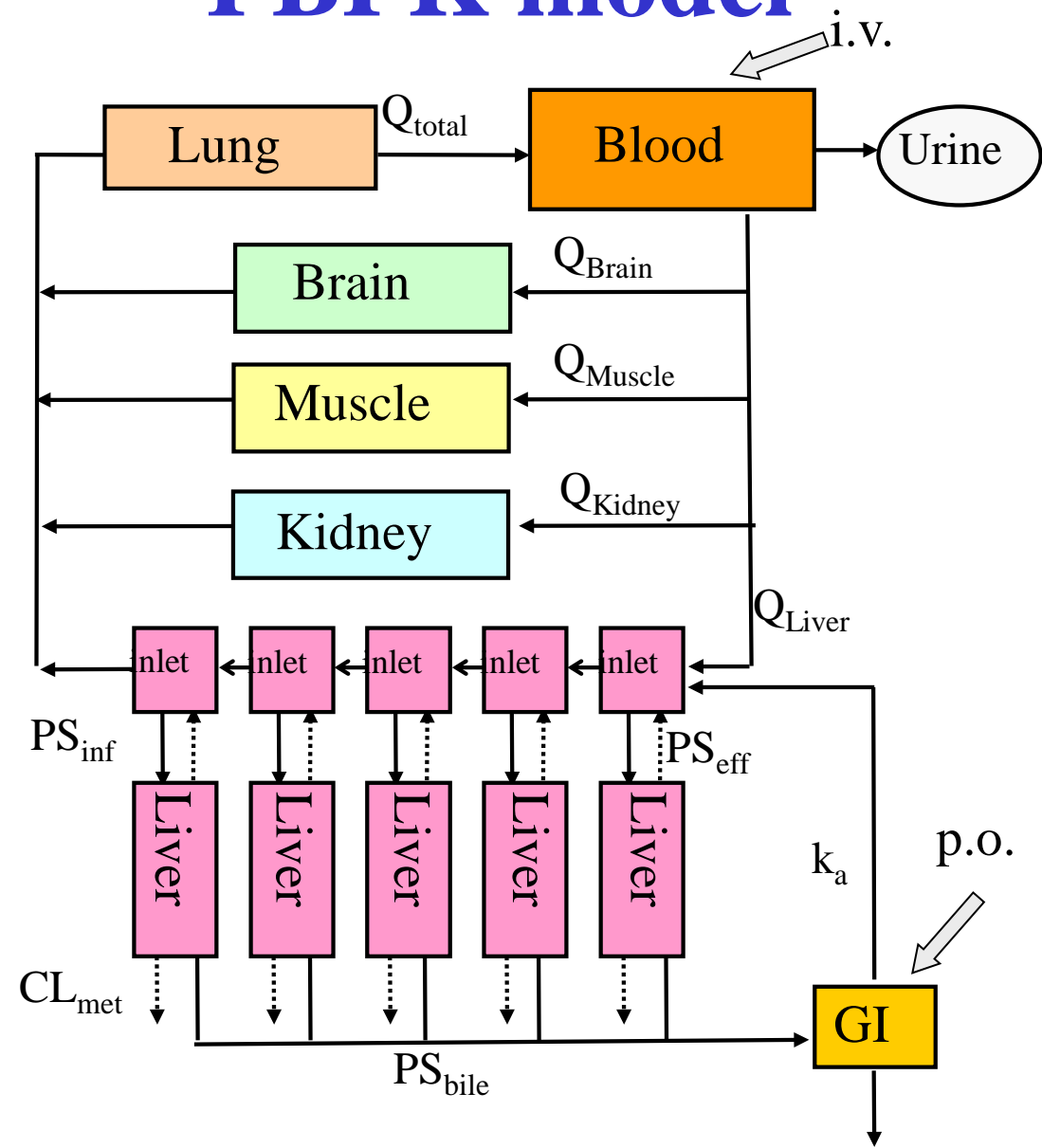
**To answer this question, it is important to estimate the exposure in the plasma (muscle ; side effect target) and in the liver (Pharmacological target) (statins, HCV drugs)**



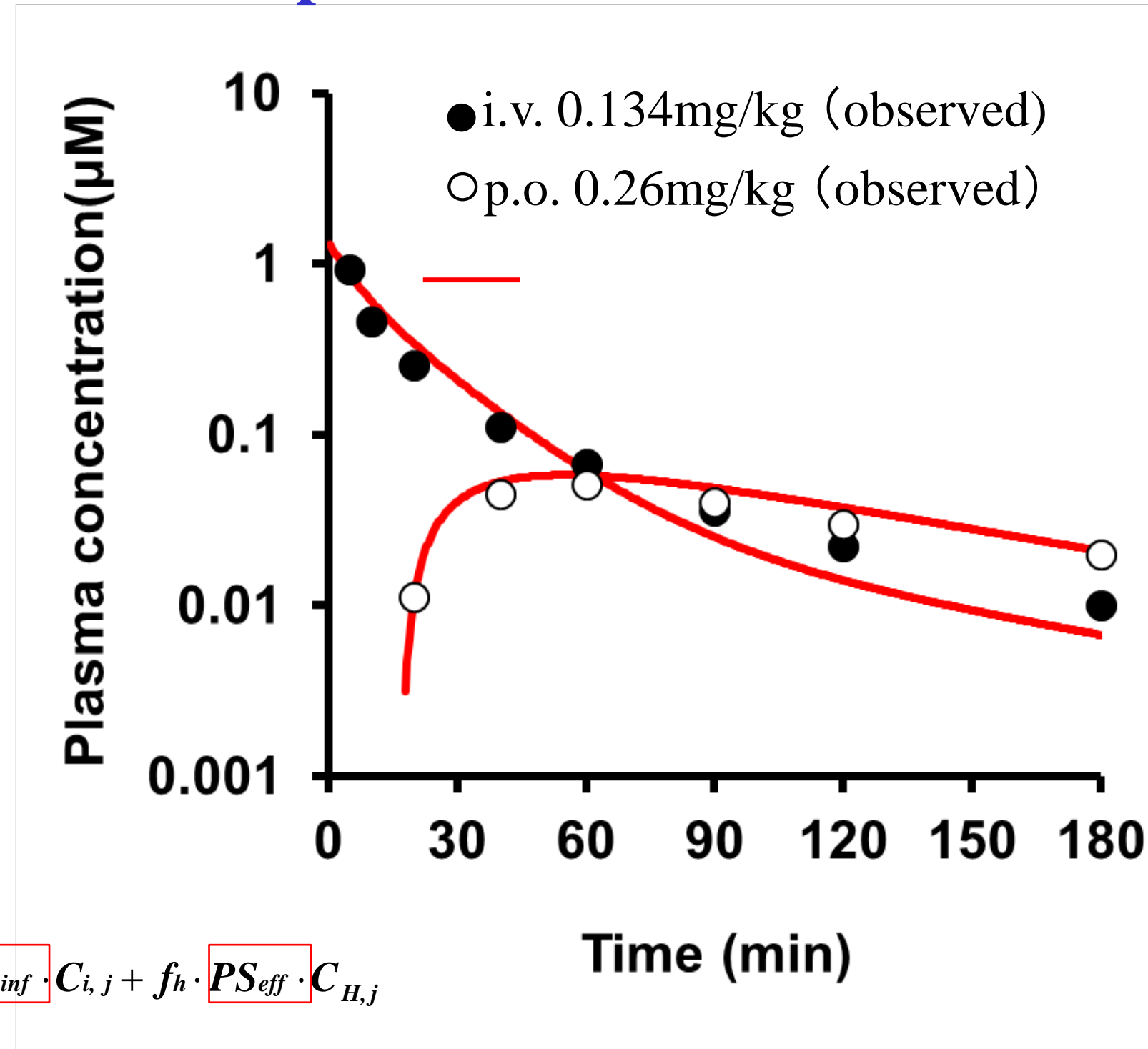
Watanabe T, Kusuhara H, Maeda K, Shitara Y and Sugiyama Y. Physiologically based pharmacokinetic modeling to predict transporter-mediated clearance and distribution of pravastatin in humans. **J Pharmacol Exp Ther** 328:652-662 (2009)

Yoshikado T, Yoshida K, Kotani N, Nakada T, Asaumi R, Toshimoto K, Maeda K, Kusuhara H, Sugiyama Y. Quantitative analyses of hepatic OATP-mediated interactions between statins and inhibitors using PBPK modeling with a parameter-optimization method. **Clin Pharmacol Ther.** 100:513-523(2016)

# PBPK model



# Plasma conc-time profile of pravastatin in human



**Blood** 
$$V_b \cdot \frac{dC_b}{dt} = Q_h(C_i - C_b) - CL_r \cdot C_b$$

**Capillary in the liver** 
$$V_{i,j} \cdot \frac{dC_{i,j}}{dt} = Q_h(C_{b,j} - C_{i,j}) - f_b \cdot PS_{inf} \cdot C_{i,j} + f_h \cdot PS_{eff} \cdot C_{H,j}$$

**Hepatocytes** 
$$V_{H,j} \cdot \frac{dC_{H,j}}{dt} = f_b \cdot PS_{inf} \cdot C_{i,j} - f_h \cdot (PS_{eff} + CL_{met} + PS_{bile}) \cdot C_{H,j}$$

  Extrapolated parameters based on in vitro (animal, human) and in vivo (animal)

## PBPK modeling of pravastatin disappearance

# Uptake-limited hepatic elimination of statins

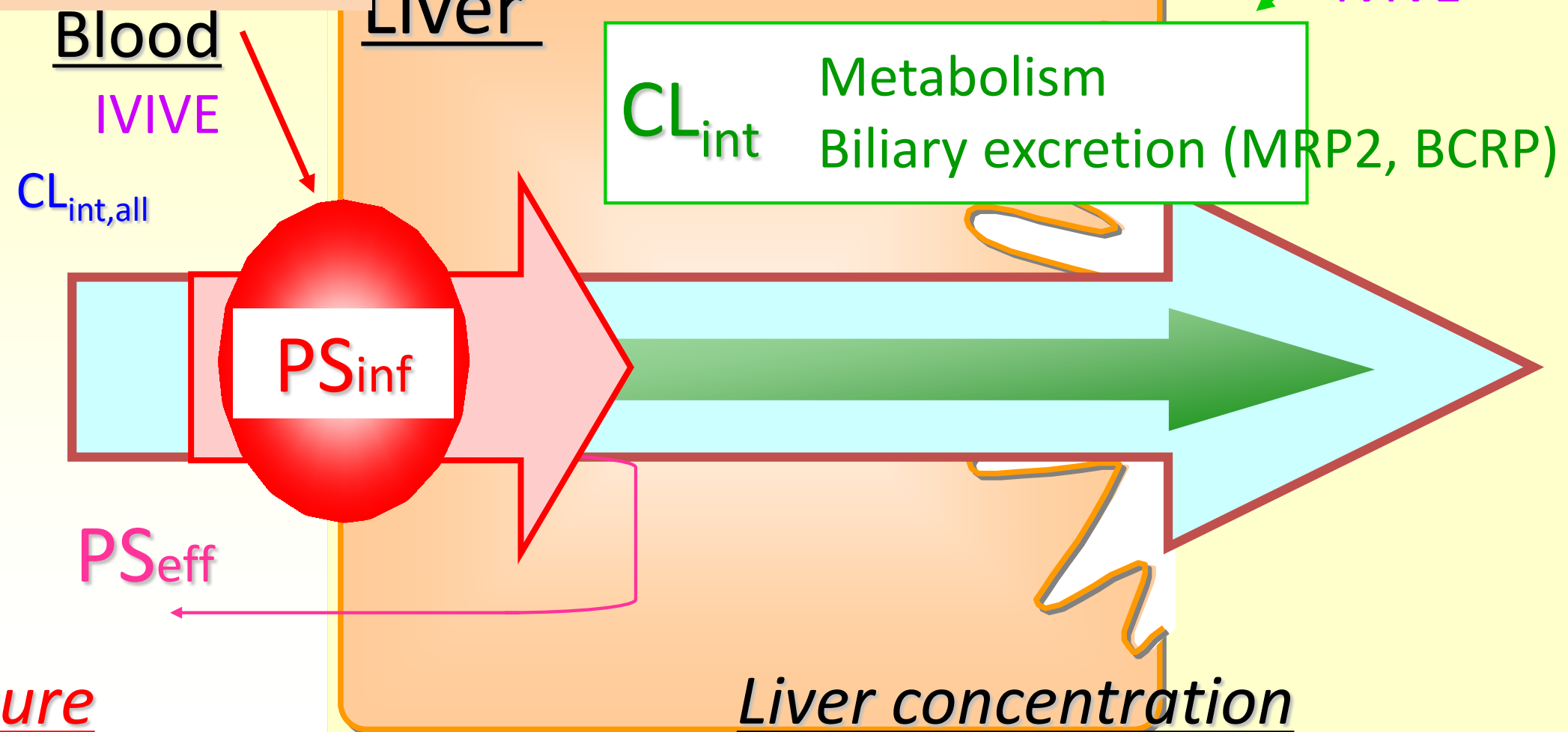
We need PET studies to know the drug exposure change in the liver

$$CL_{int,all} = PS_{inf} \times \frac{CL_{int}}{PS_{eff} + CL_{int}}$$

Microsome  
IVIVE

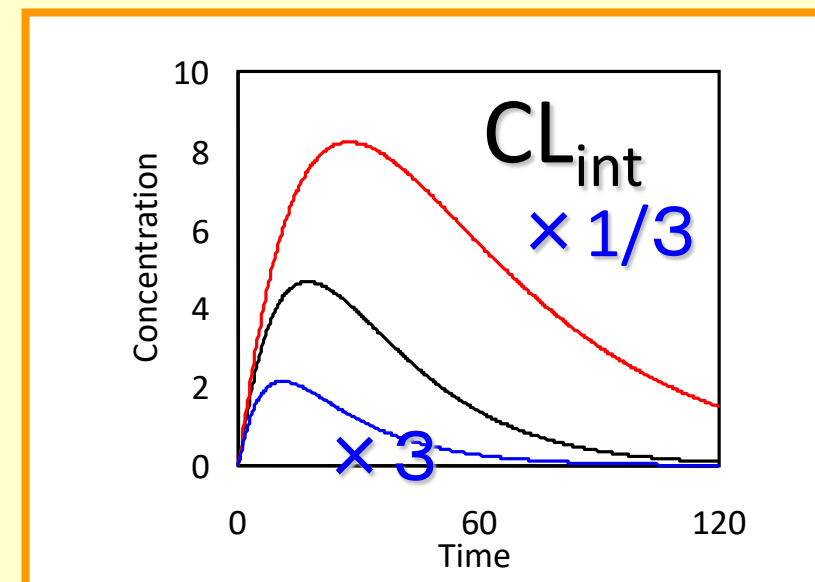
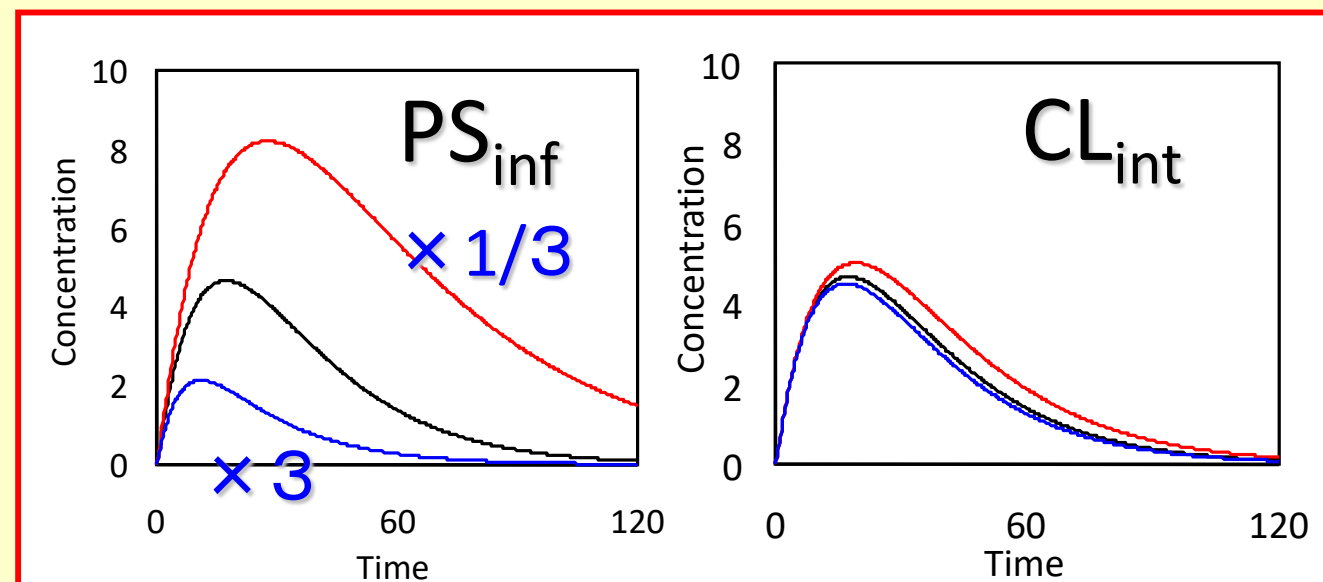
## Statins

Pravastatin  
Pitavastatin  
Atorvastatin  
Fluvastatin



## Systemic exposure

## Liver concentration





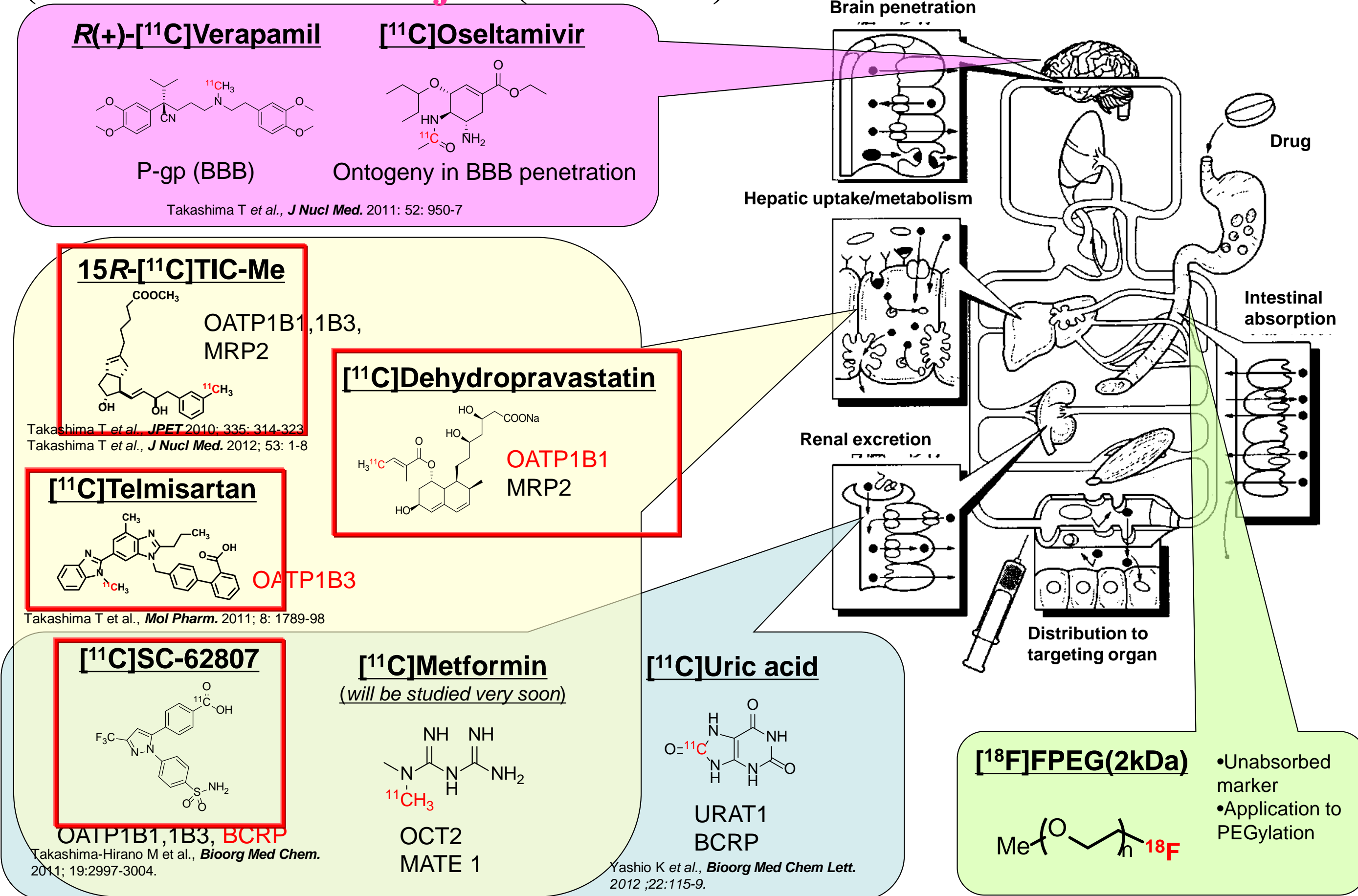
## Summary :

- 1) **Sensitivity analysis indicated that the change in hepatic uptake ability alters the plasma concentration profile sensitively (toxicity) and may not affect the profile in the liver, target tissue (pharmacological effect). GWAS for simvastatin in fact demonstrated it was the case.**
- 2) **Alteration in the biliary excretion ability (MRP2, BCRP) may affect the pharmacological effect (hepatic exposure) much more sensitively than that of the uptake, though there is little change in the plasma exposures.**  
**⇒ We have to confirm it by PET analyses**

**This prediction has been supported by several studies published by other groups (simvastatin GWAS study, rosuvastatin Jupiter trial)**

# Drug development with the Use of Microdosing Clinical Trial: Based on the Quantitative Prediction Technology of ADME

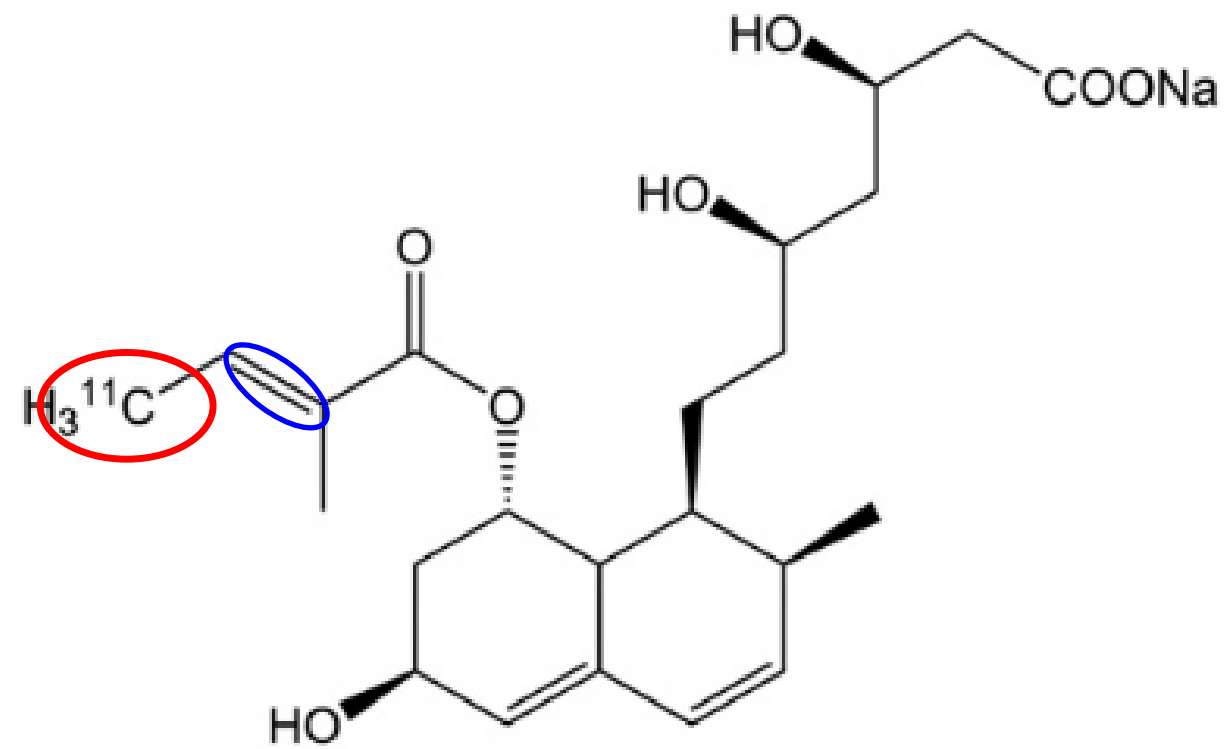
(**NEDO Research Project (2008-2011)** collaboration with Y.Watanabe



# PET imaging human studies

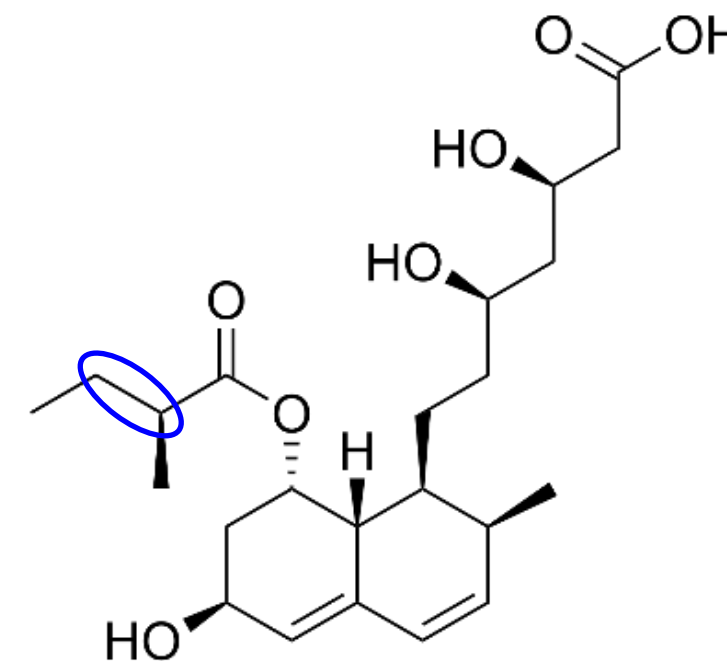
## [<sup>11</sup>C]DHP PET studies

[<sup>11</sup>C]DPV chemical structure



**Metabolism is minimized**

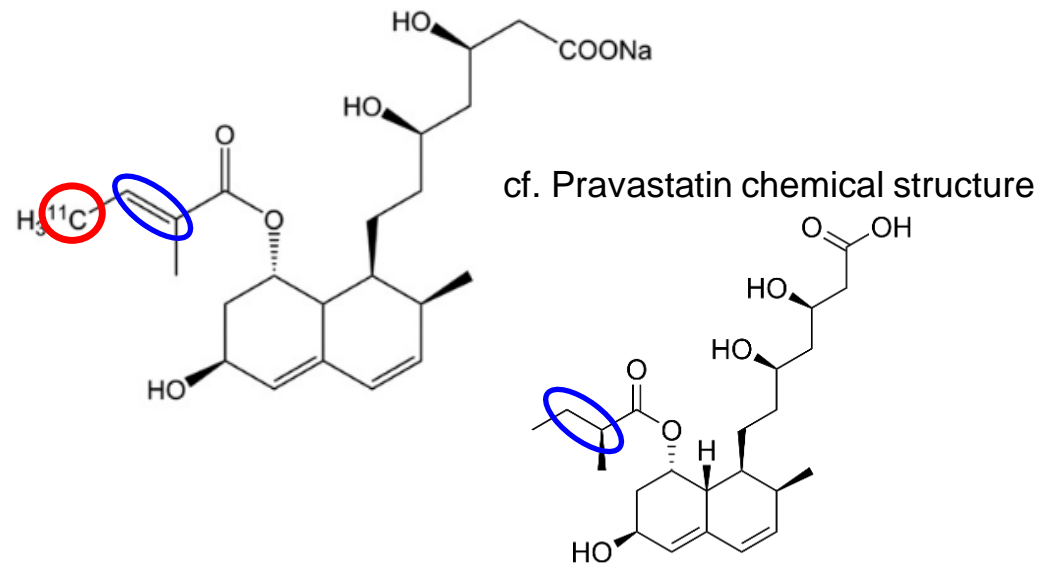
cf. Pravastatin chemical structure



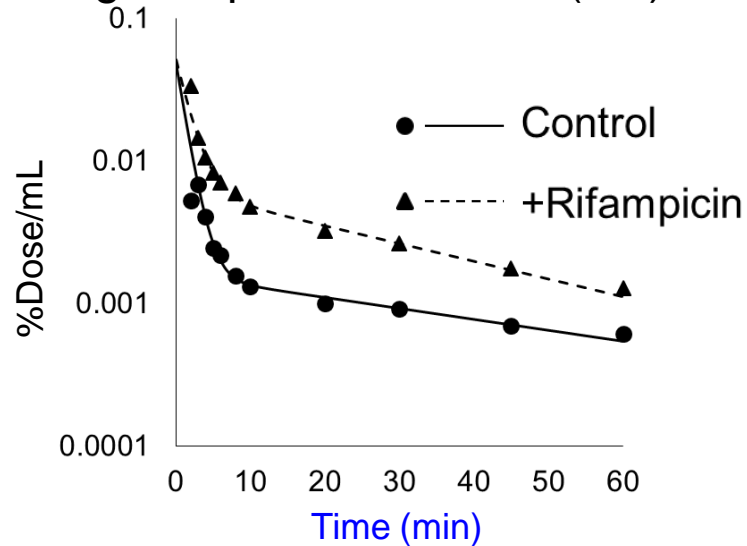
Kaneko K et al., A clinical quantitative evaluation of hepatobiliary transport of [<sup>11</sup>C]Dehydropravastatin in humans using positron emission tomography. *Drug Metab Dispos.* 46(5):719-728 (2018).

# [<sup>11</sup>C]DHP dynamic model fitting

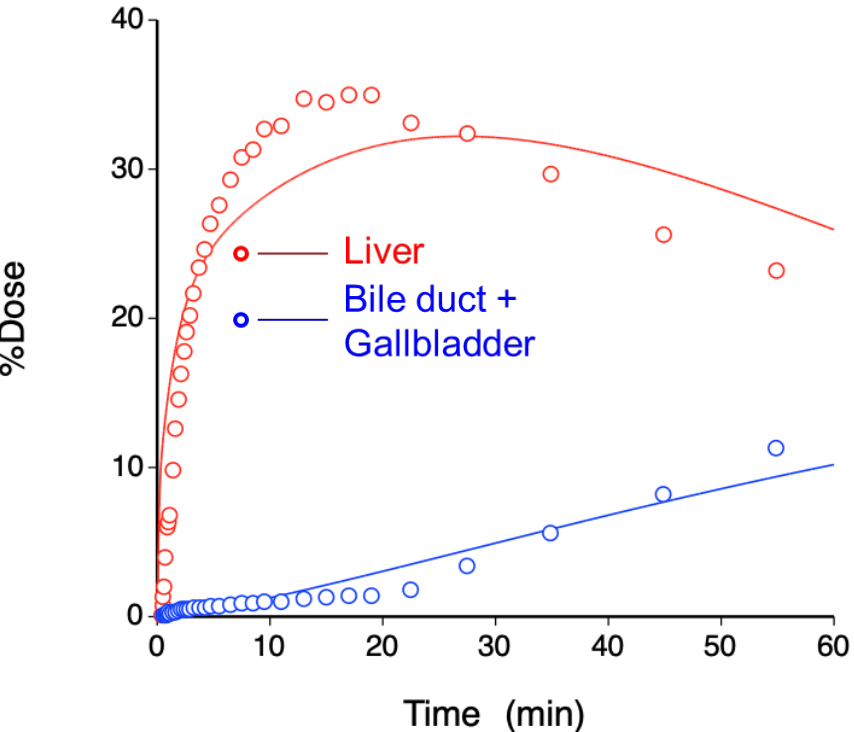
[<sup>11</sup>C]DPV chemical structure



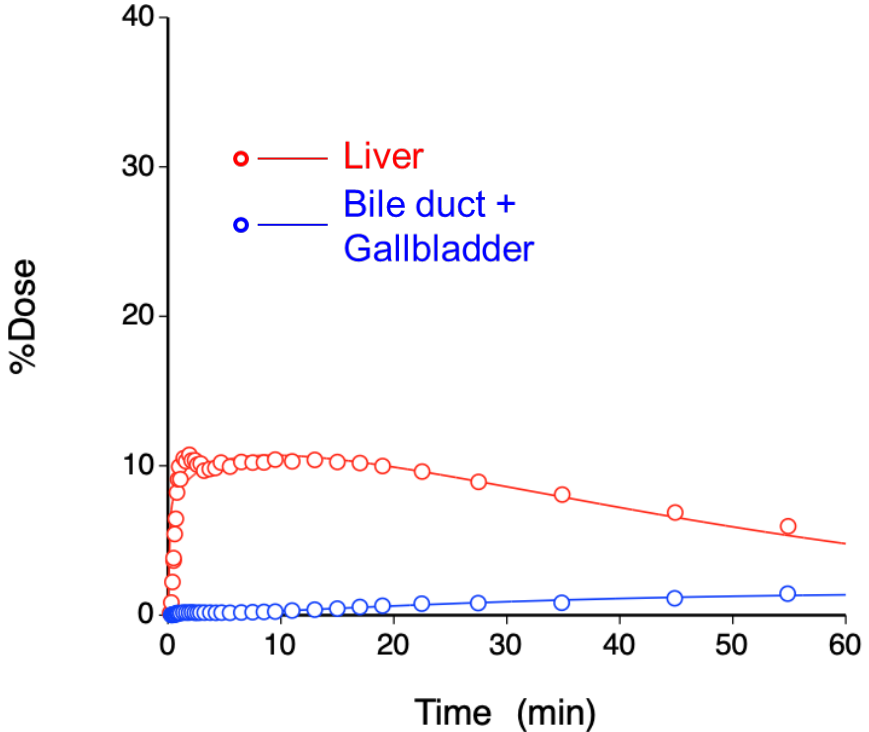
Fitted blood radioactivity-time profile of DPV using 2-exponential curve (#2)



Fitted radioactivity-time profile of DPV on **control phase** using dynamic model (#2)



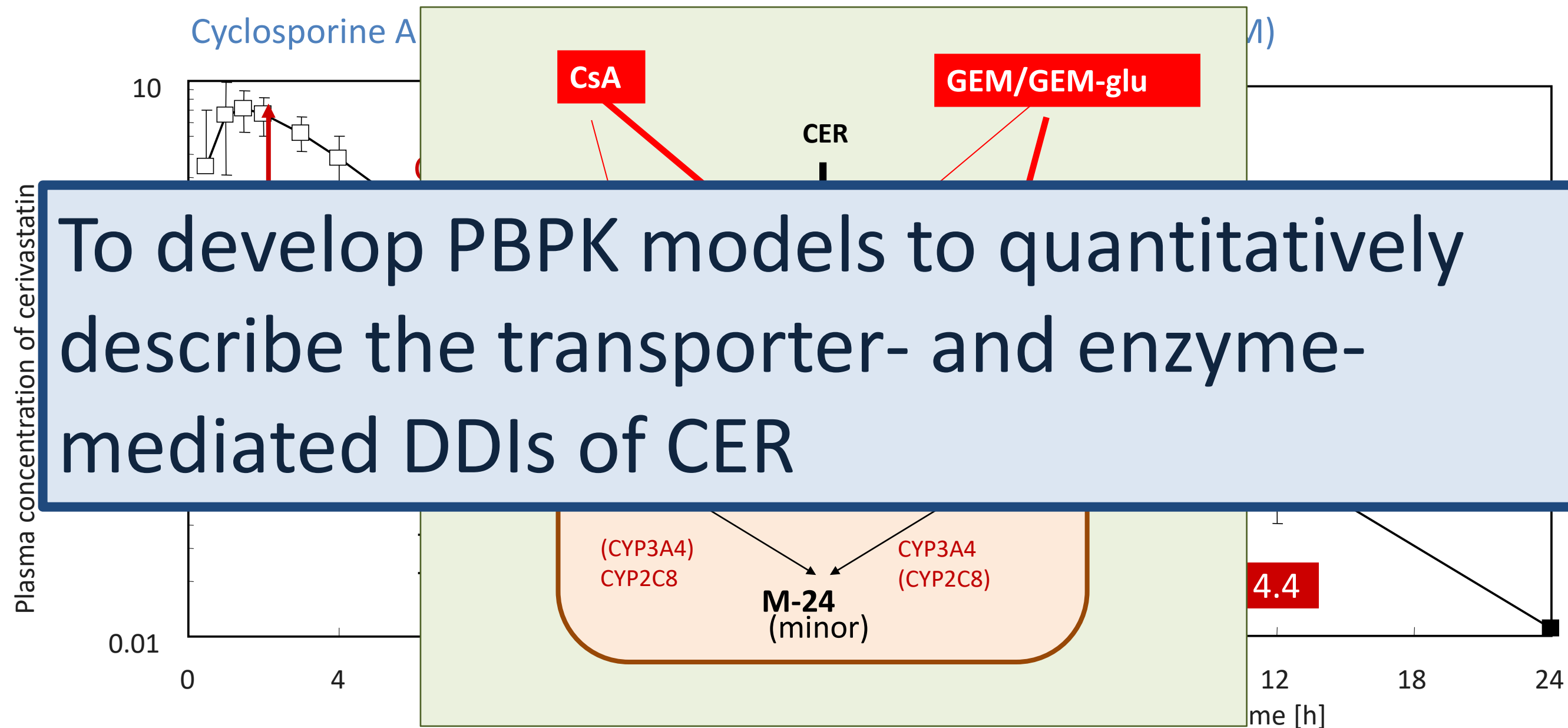
Fitted radioactivity-time profile of DPV on **+rifampicin phase** using dynamic model (#2)



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# Drug-Drug Interaction between CER and Cyclosporine A/Gemfibrozil



Mück W et al., *Clin Pharmacol Ther.*, (1999)

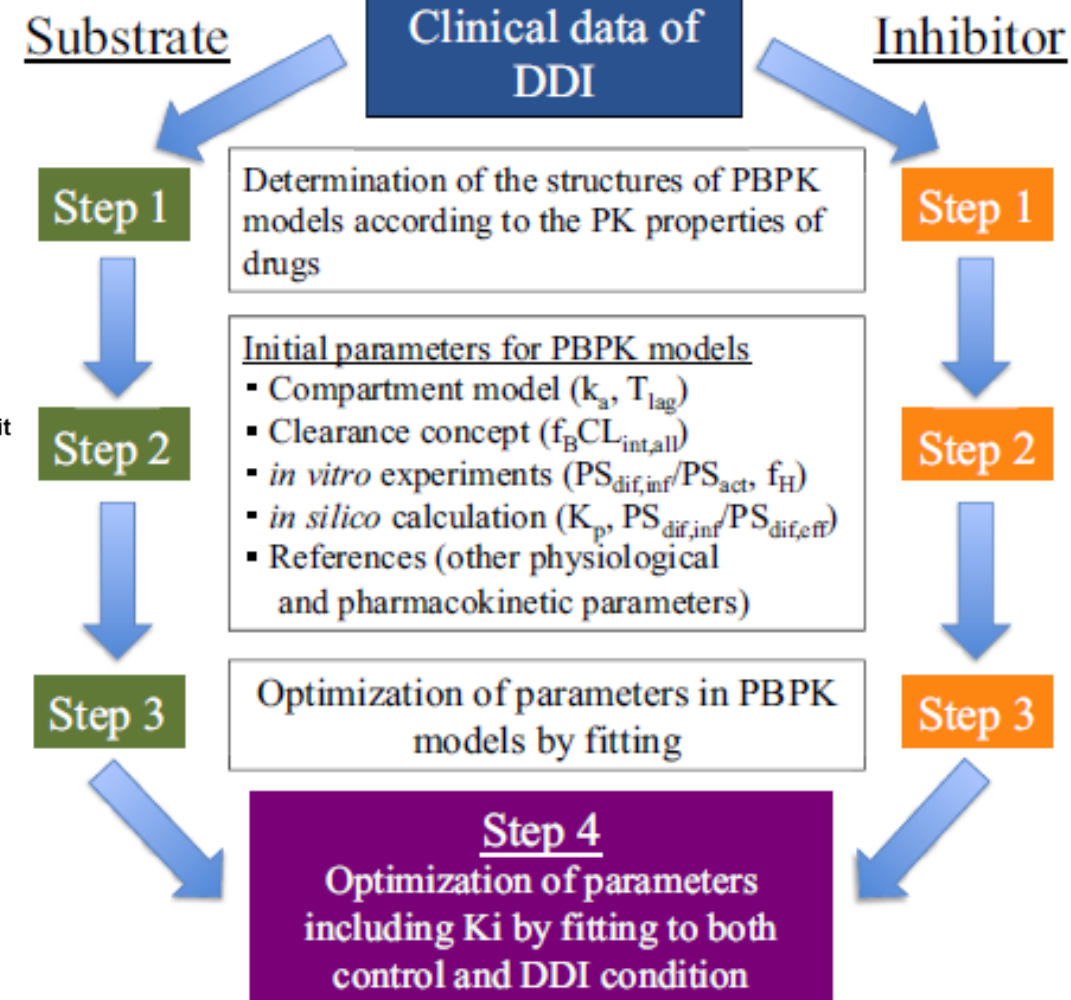
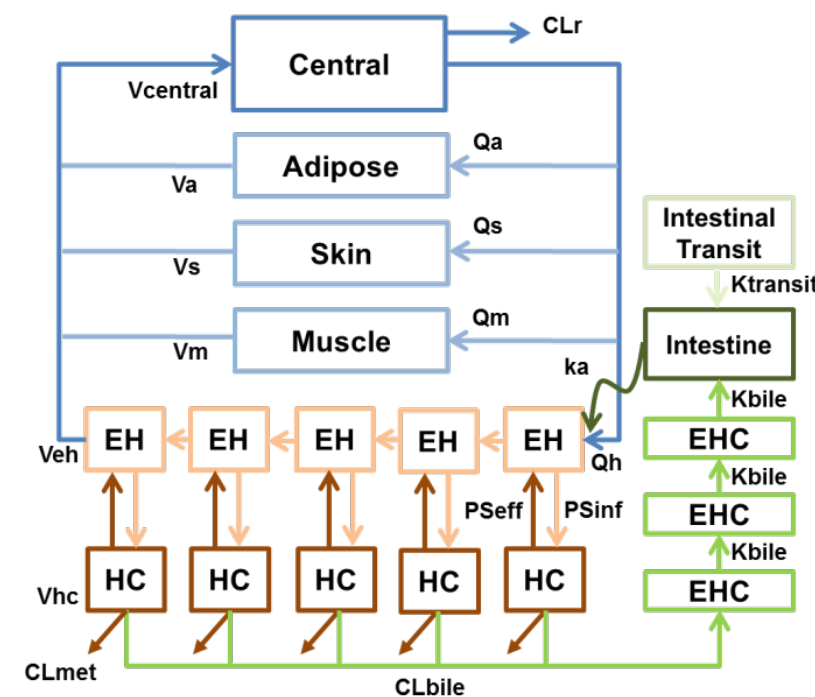
Backman JT et al., *Clin Pharmacol Ther.*, (2002)

$CL_h \downarrow \quad V_d \downarrow \quad T_{1/2} \rightarrow$

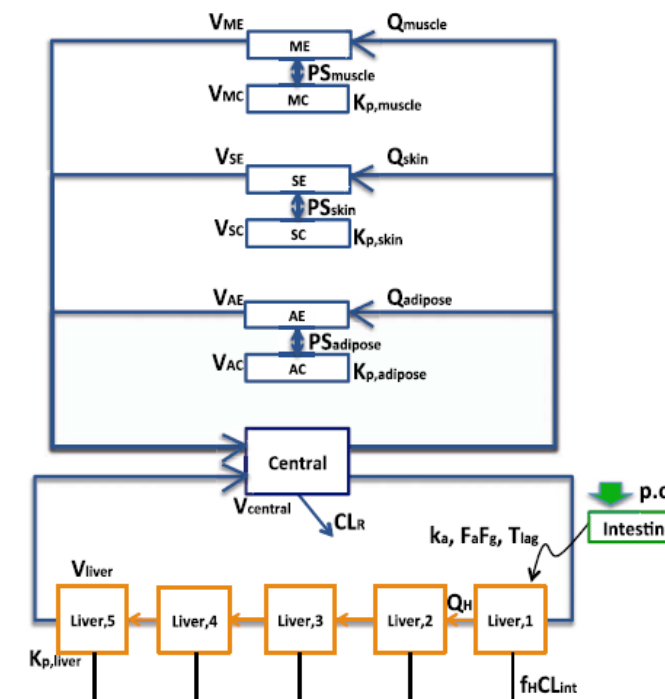
$CL_h \downarrow \quad V_d \rightarrow \quad T_{1/2} \downarrow$

# PBPK analyses of transporter/enzyme mediated complex DDI

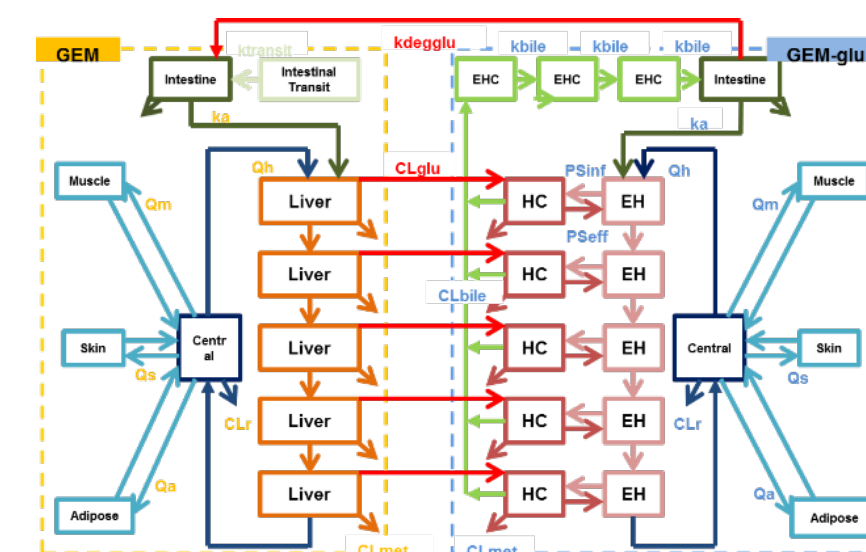
## Cerivastatin (CER)



## Cyclosporine A (CsA)



## Gemfibrozil (GEM)

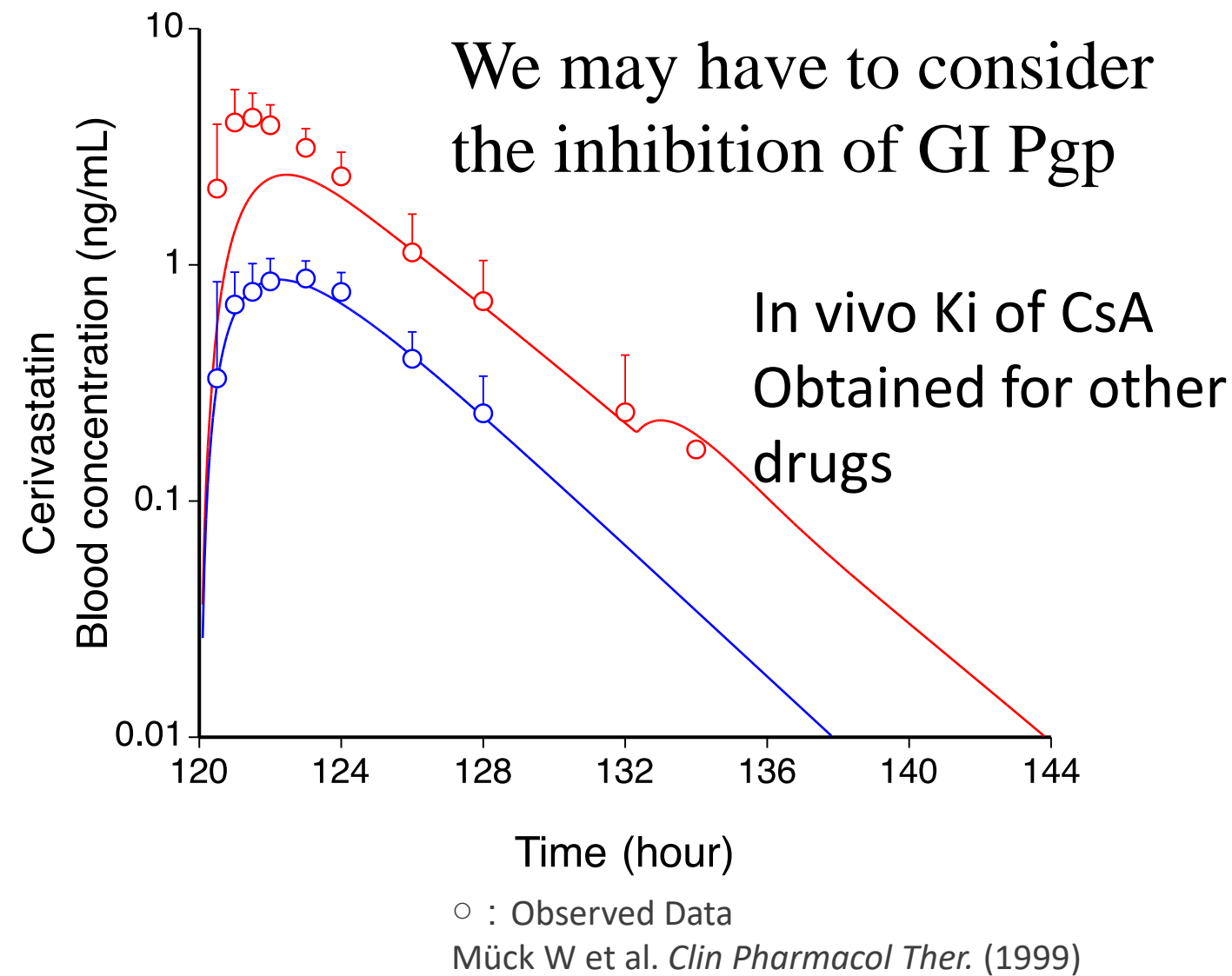


Integration of bottom-up approach  
And top-down approach is important

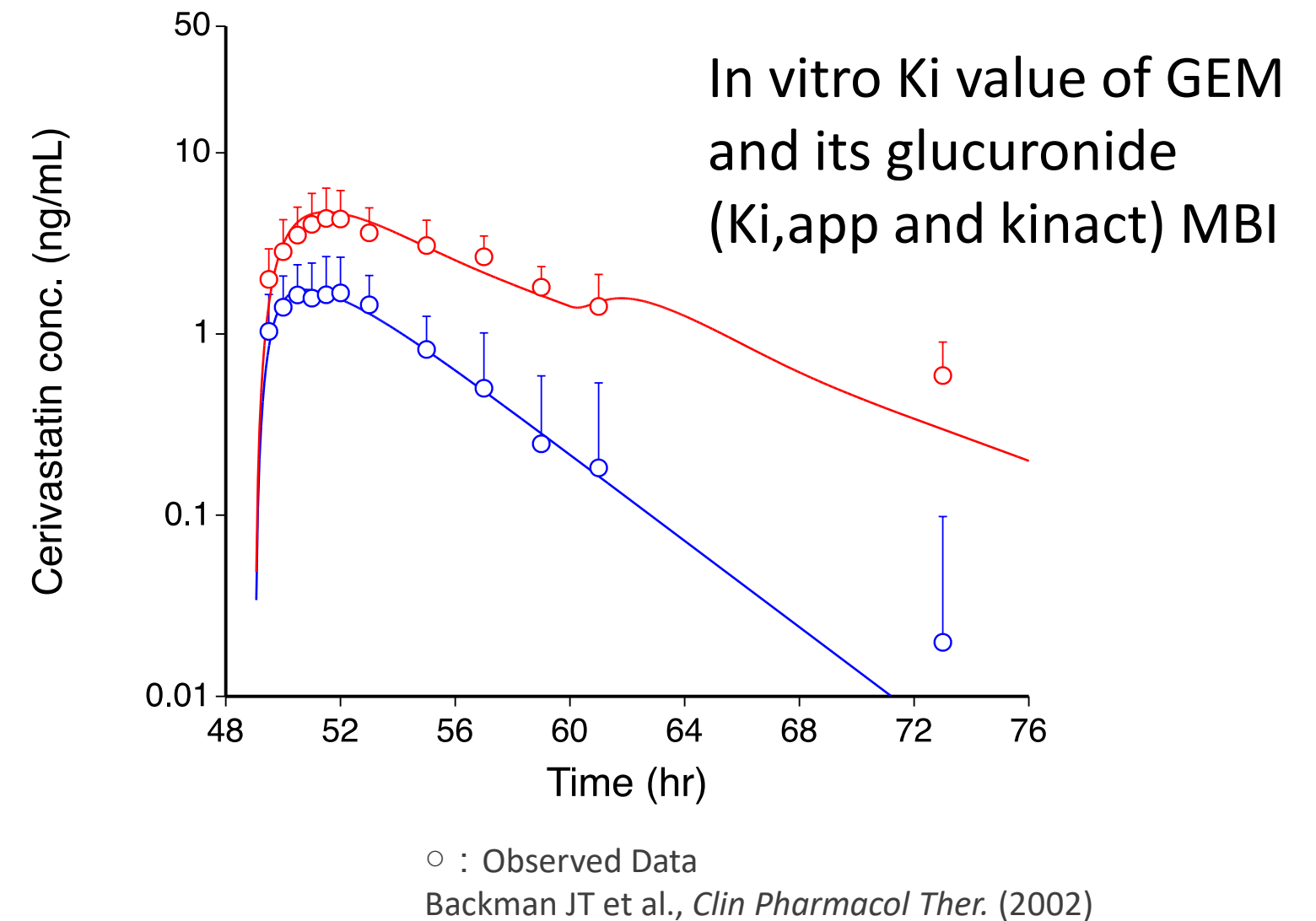
Yoshikado T et al.,  
*Clin Pharmacol Ther.*, (2016)

# Best simulation of CER vs CsA / CER vs GEM

## CER vs CsA



## CER vs GEM



## Present PBPK models were able to well reproduce the clinical DDIs

Yao Y, Toshimoto K, Kim SJ, Yoshikado T, Sugiyama Y. Quantitative Analysis of Complex Drug-Drug Interactions between Cerivastatin and Metabolism/Transport Inhibitors Using Physiologically Based Pharmacokinetic Modeling. *Drug Metab Dispos.* 46:924-933. (2018)



# Summary

- The concentration-time profiles for CER and GEM/GEM-glu described by PBPK models were well agreed with the clinically observed data.
- The present PBPK models were able to capture the clinical DDIs
  - CER and CsA : using  $x1/2$  *in vivo*  $K_i$  value for OATP1B1 which is previously reported (substrate-dependent  $K_i$ ). We may have to take into account the inhibition of intestinal Pgp to better describe this DDI
  - CER and GEM: using reported *in vitro*  $k_{inact}$  value (GEM-glu) and  $K_i$  value for OATP1B1 or CYP2C8 when  $fm_{2C8} = 0.85$  value was used



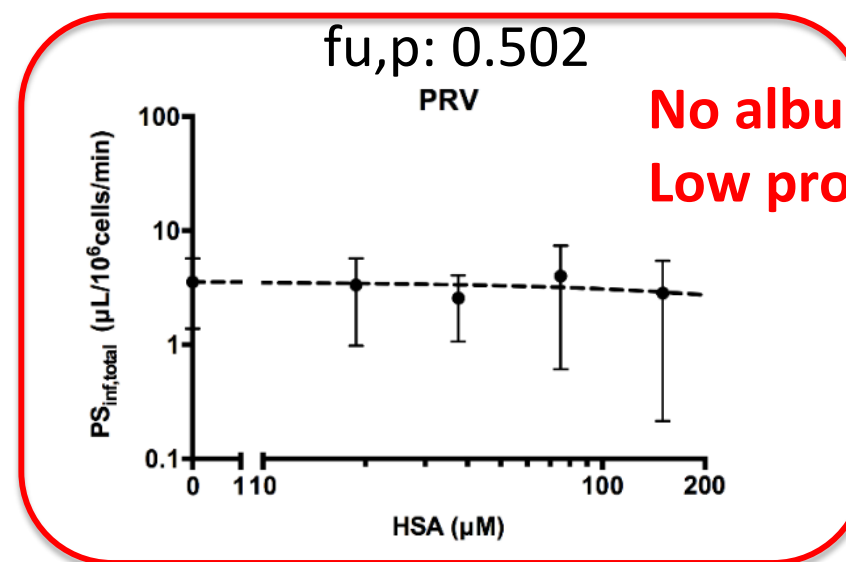
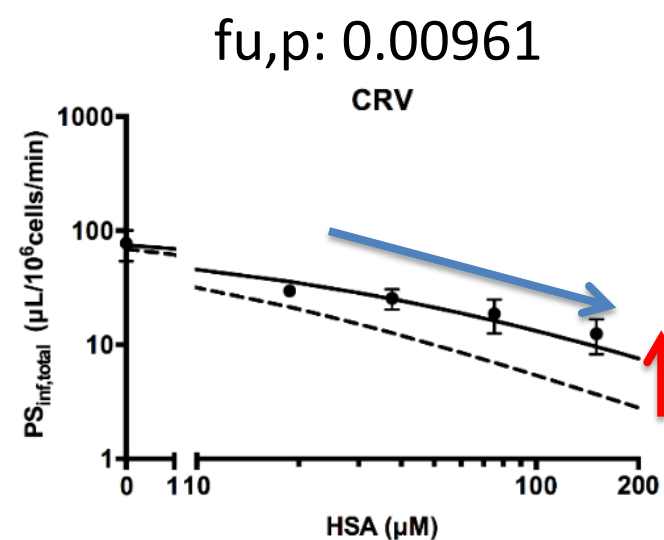
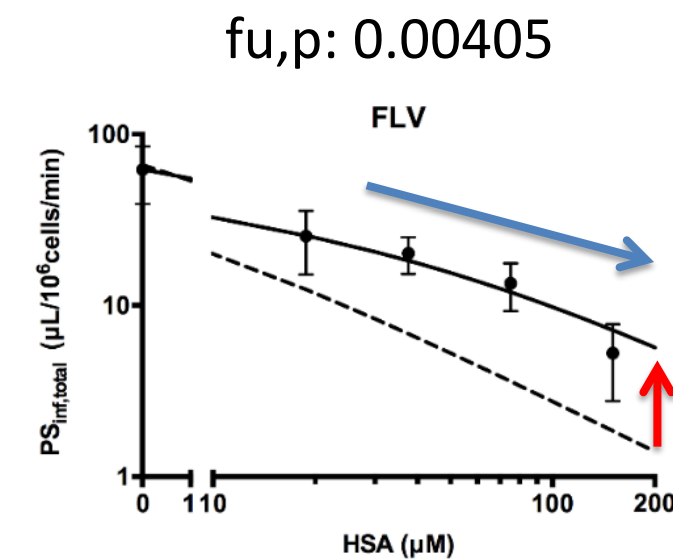
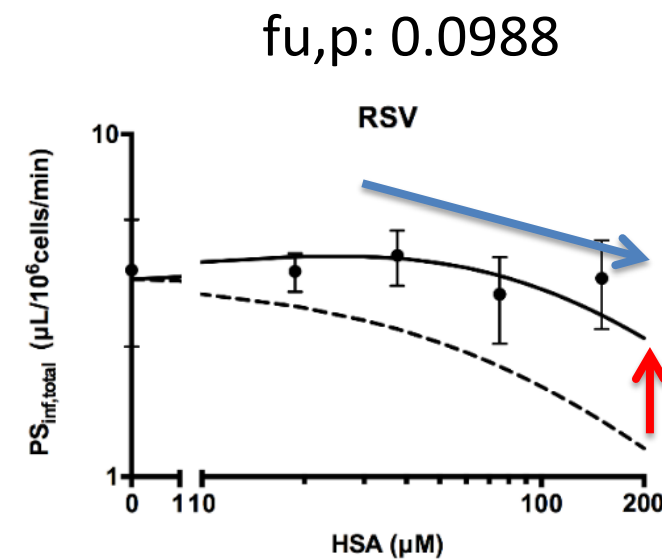
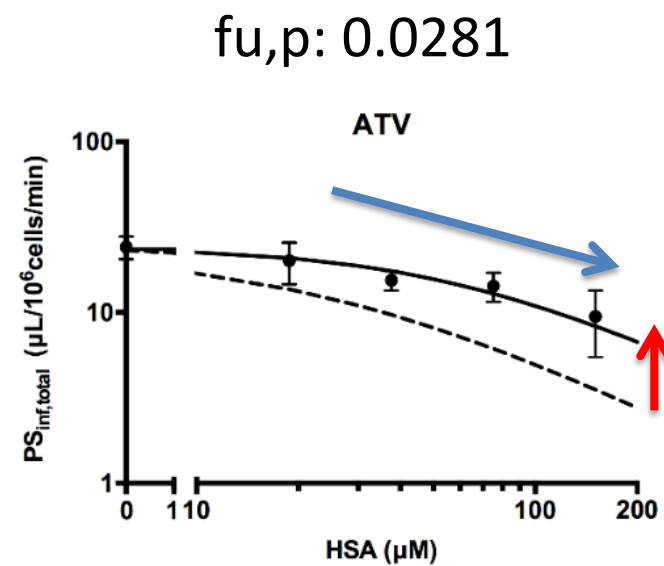
**Complex DDIs involving both transporters and metabolic enzymes could be quantitatively predicted by PBPK modeling (Bottom-up approach) based on the *in vitro* parameters ( $K_i$ ,  $k_{inact}$ )**

**We now know some cases where simple bottom up prediction (IVIVE) cannot be applied.**

- 1) Albumin-mediated hepatic uptake observed for highly protein bound drugs (OATP1B, OAT2 substrates)**
- 2) Time-dependent inhibition constant ( $K_{i,app}$  value) observed for some OATP1B inhibitors and OCT1 inhibitors**

# Albumin-mediated uptake of 10 OATPs substrates

## Uptake clearance for total drug in various HSA concentrations



No albumin effect,  
Low protein binding drug

Solid line: fitted data by Taso's model  
Broken line: free theoretical line

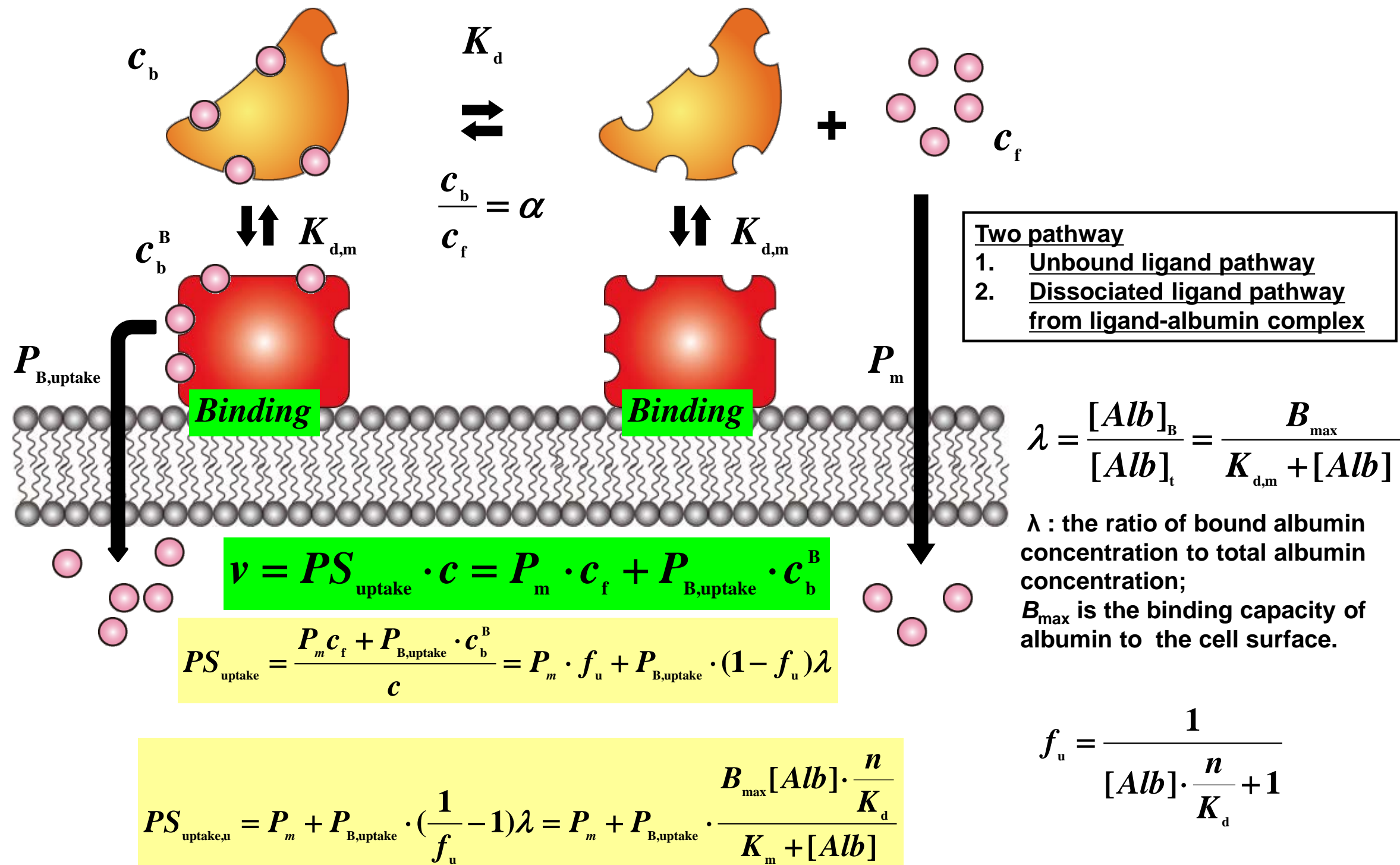
**Soo-JinKim, Kyeong-RyoonLee, Seiji Miyauchi, and Yuichi Sugiyama**

Extrapolation of In Vivo Hepatic Uptake Clearance from In Vitro Uptake Clearance by Suspended Human Hepatocytes (IVIVE) for Anionic Drugs with High Binding to Human Albumin: Improvement of IVIVE by Considering the “Albumin-Mediated” Hepatic Uptake Mechanism Based on the Facilitated-Dissociation Model

Drug Metab Dispos. 47(2):94-103 (2019).

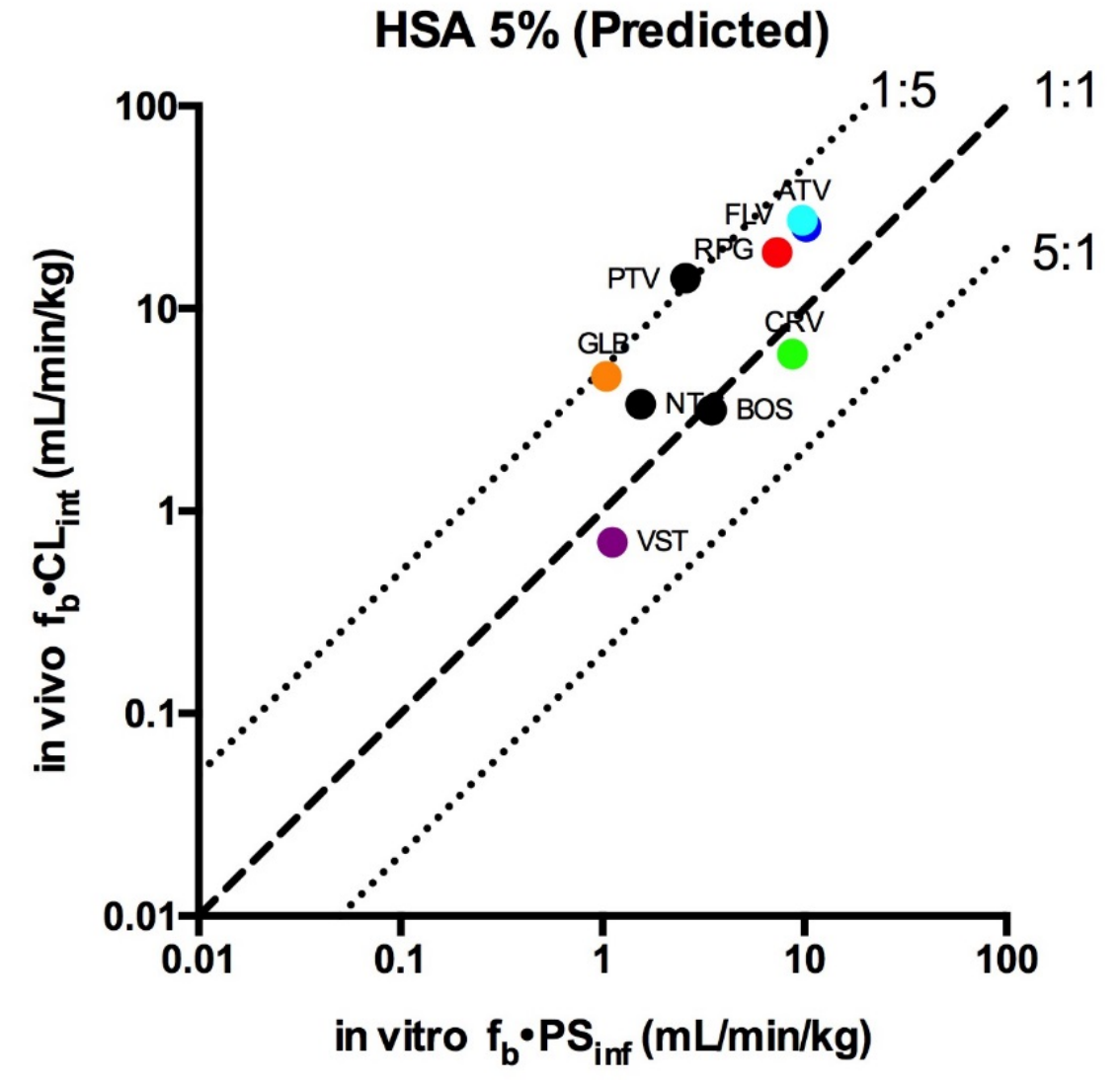
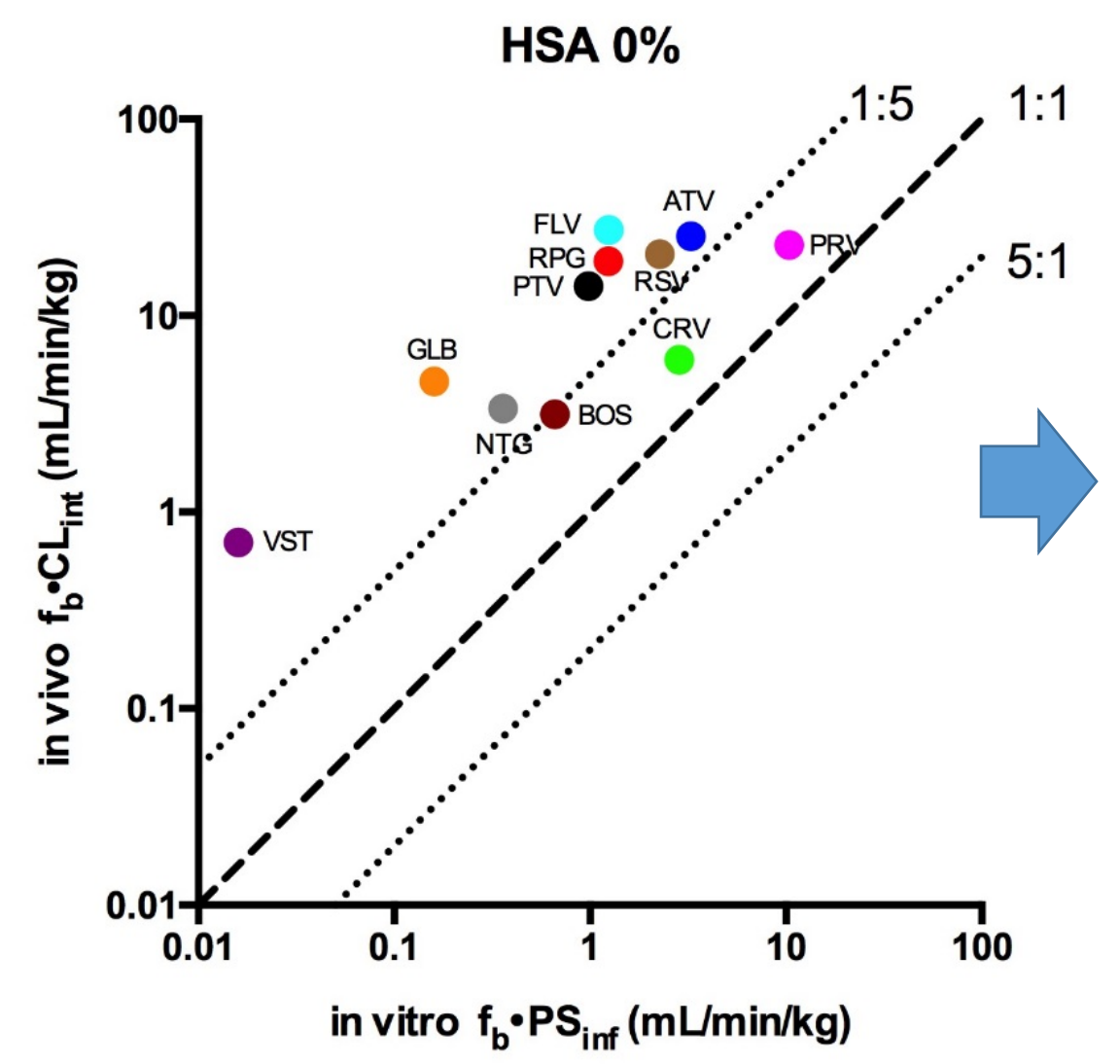
# The interaction of the albumin-ligand complex with the surface of hepatocytes enhances the dissociation of the ligand from albumin

Tsao SC, Sugiyama Y, Sawada Y, Iga T and Hanano M. Kinetic analysis of albumin-mediated uptake of warfarin by perfused rat liver. J Pharmacokinet Biopharm 16:165-181 (1988).



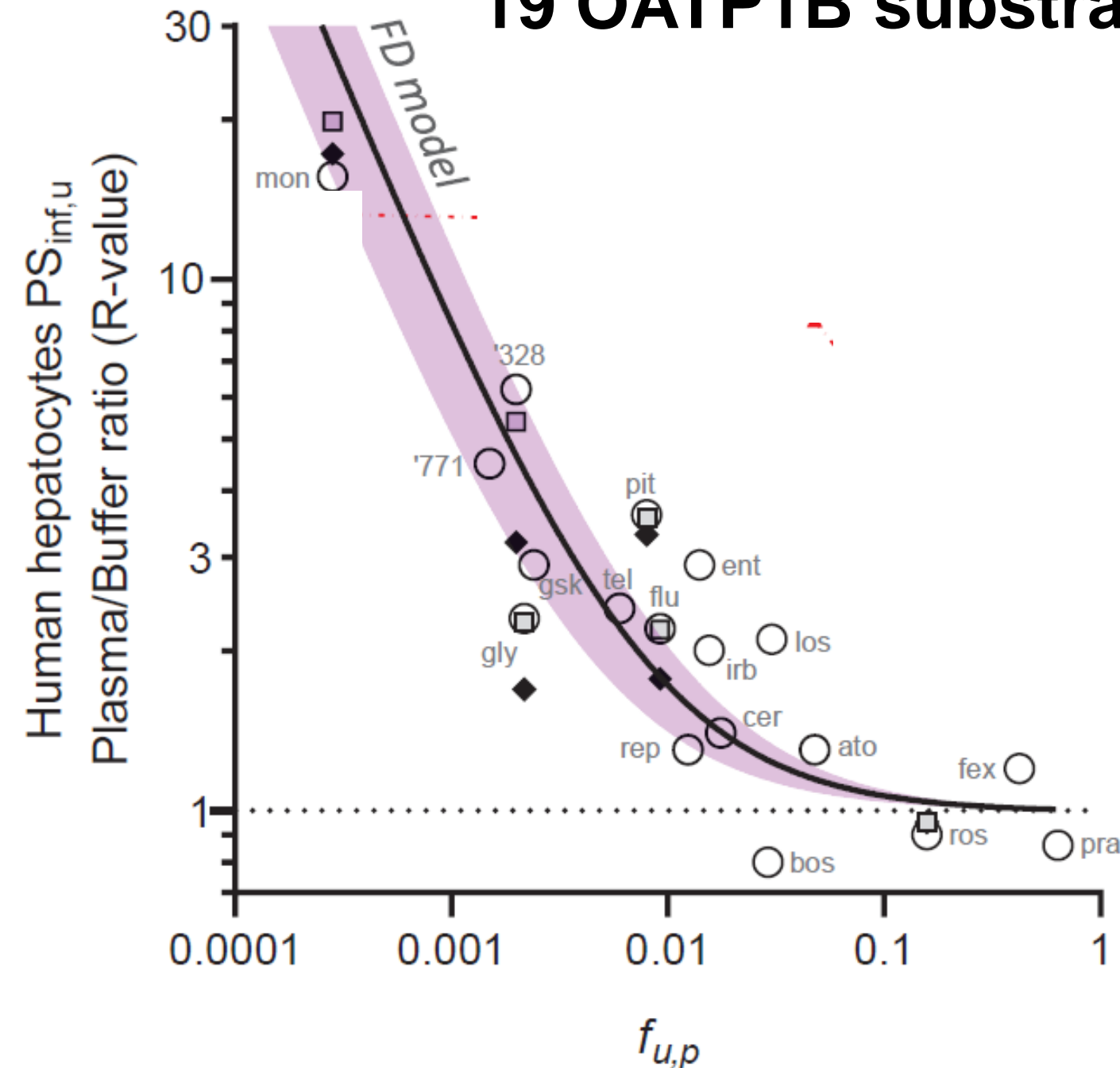
# Improvement of IVIVE of hepatic uptake clearance by taking into account of albumin-mediated hepatic uptake mechanism

IVIVE was improved by taking into account of the albumin-mediated hepatic uptake, though not perfect.



**9 compounds**  
 The  $CL_{uptake,u}$  (at 5% HSA) predicted by fitting with Taso model was used.

## 19 OATP1B substrates



### Facilitated-dissociation (FD) model

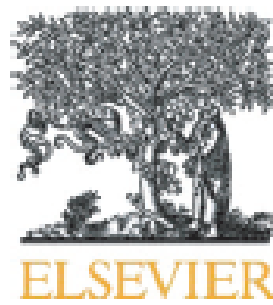
$$R = \frac{PS_{inf,u,plasma}}{PS_{inf,u,buffer}} = 1 + \left( \frac{1}{f_{u,p}} - 1 \right) \cdot \frac{r \cdot B_{max}}{K_{d,m} + [Alb]}$$

The parameter 'r' represents a ratio of uptake clearance of the unbound ligand dissociated from the ligand-albumin complex at cell surface to the uptake clearance of the unbound ligand dissociated in the plasma away from cell surface. **For data fitting, 'r · B<sub>max</sub>' was considered as a hybrid constant as they were individually un-identifiable.**

Yi-an Bi, Sangwoo Ryu, David A. Tess, A. David Rodrigues, Manthena V. S. Varma  
 Effect of Human Plasma on Hepatic Uptake of Organic Anion-Transporting Polypeptide 1B  
 Substrates: Studies using Transfected Cells and Primary Human Hepatocytes  
 DMD Fast Forward. Published on November 2, 2020 as DOI: 10.1124/dmd.

**We now know some cases where simple IVIVE cannot be applied.**

- 1) Albumin-mediated hepatic uptake observed for highly protein bound drugs (OATP1B, OAT2 substrates)**
- 2) Time-dependent inhibition constant ( $K_{i,app}$  value) observed for some OATP1B inhibitors and OCT1 inhibitors**



## Preincubation-dependent and long-lasting inhibition of organic anion transporting polypeptide (OATP) and its impact on drug-drug interactions



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<sup>a</sup> Pharmacokinetics, Dynamics and Metabolism, Translational Medicine and Early Development, R&D Operations, Sanofi, Tokyo, Japan

<sup>b</sup> Sugiyama Laboratory, RIKEN Innovation Center, RIKEN, Yokohama, Japan

### ARTICLE INFO

Available online 27 February 2017

#### Keywords:

OATP1B1

Drug–drug interactions

Hepatic uptake

Time-dependent inhibition

Long-lasting inhibition

Physiologically based pharmacokinetic model

Modeling & simulation

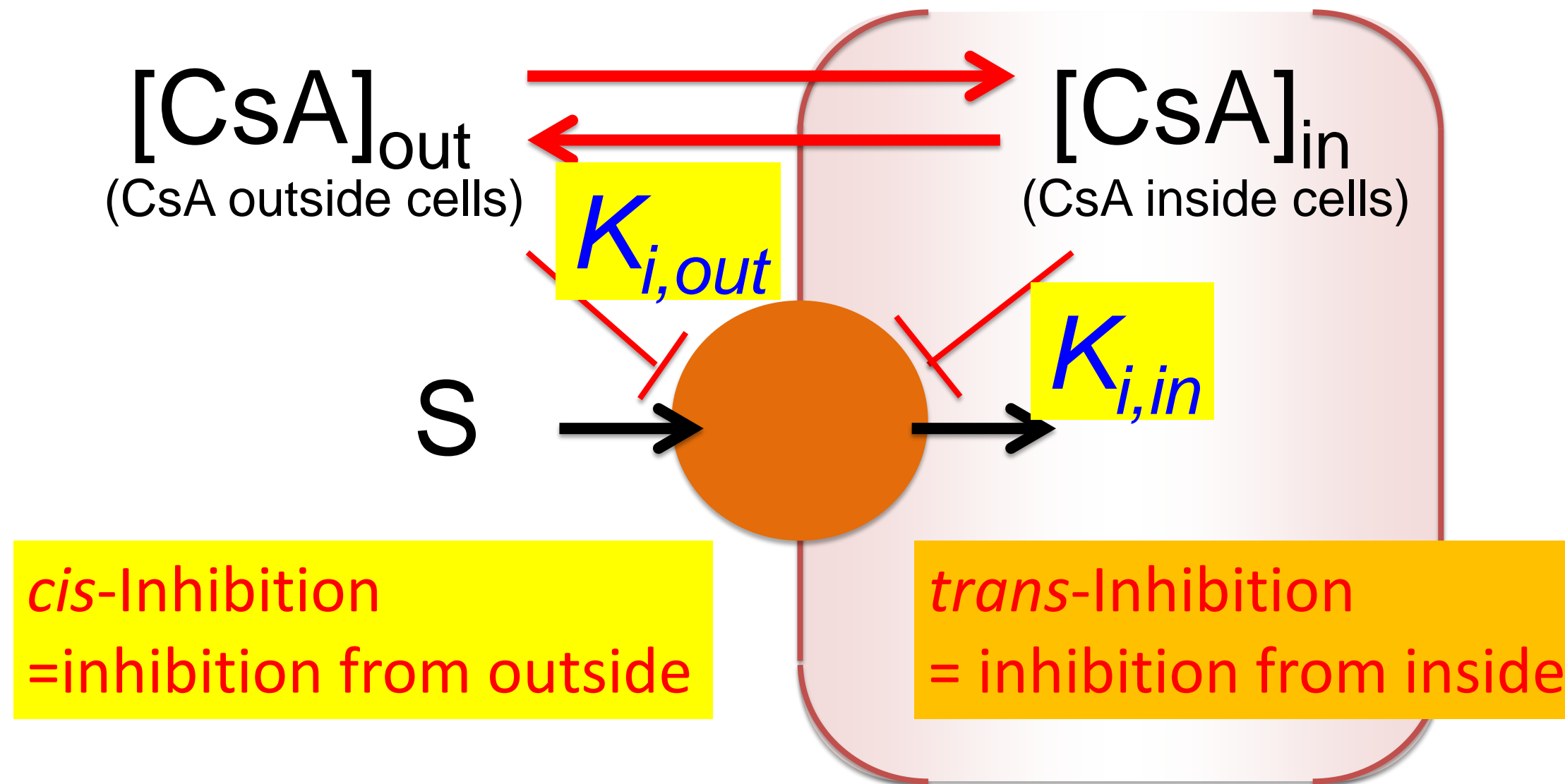
### ABSTRACT

Preincubation with cyclosporin A (CsA), a potent inhibitor of organic anion transporting polypeptide 1B1 (OATP1B1) and OATP1B3, enhanced its inhibitory effects on these transporters *in vitro*. A similar effect was observed upon preincubation with some other inhibitors. Removing these from the incubation media did not readily reverse the inhibition on OATP1B1 and OATP1B3. This preincubation-dependent long-lasting inhibition appeared to be related to CsA concentration in the cells in addition to that in the incubation media. Thus, we hypothesized that CsA inhibits OATP1B1 and OATP1B3 from inside (*trans*-inhibition) as well as outside (*cis*-inhibition) the cells and constructed the *cis*- and *trans*-inhibition model. The enhanced inhibitory effect of CsA on OATP1B1 observed after preincubation was quantitatively described using  $K_{i,out}$  and  $K_{i,in}$  as inhibition constants for *cis*- and *trans*-inhibitions, respectively. In addition, a long-lasting inhibition was also described by this model. Additional factors taken into consideration when simulating *in vivo* pharmacokinetic alterations by CsA are potential inhibition by AM1, a major metabolite of CsA, which has been reported to inhibit OATP1B1 and OATP1B3. Based on the physiologically based pharmacokinetic model incorporating *trans*- and *cis*-inhibition of OATP1B1 by CsA, the simulation showed that OATP1B1-mediated drug–drug interaction with CsA was suggested to be time-dependent also *in vivo* although further clinical studies are required for confirmation.

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# Cis- and trans-inhibition of CsA on OATP1B1



Uptake clearance of OATP1B1 substrates in the liver when co-administered with CsA:

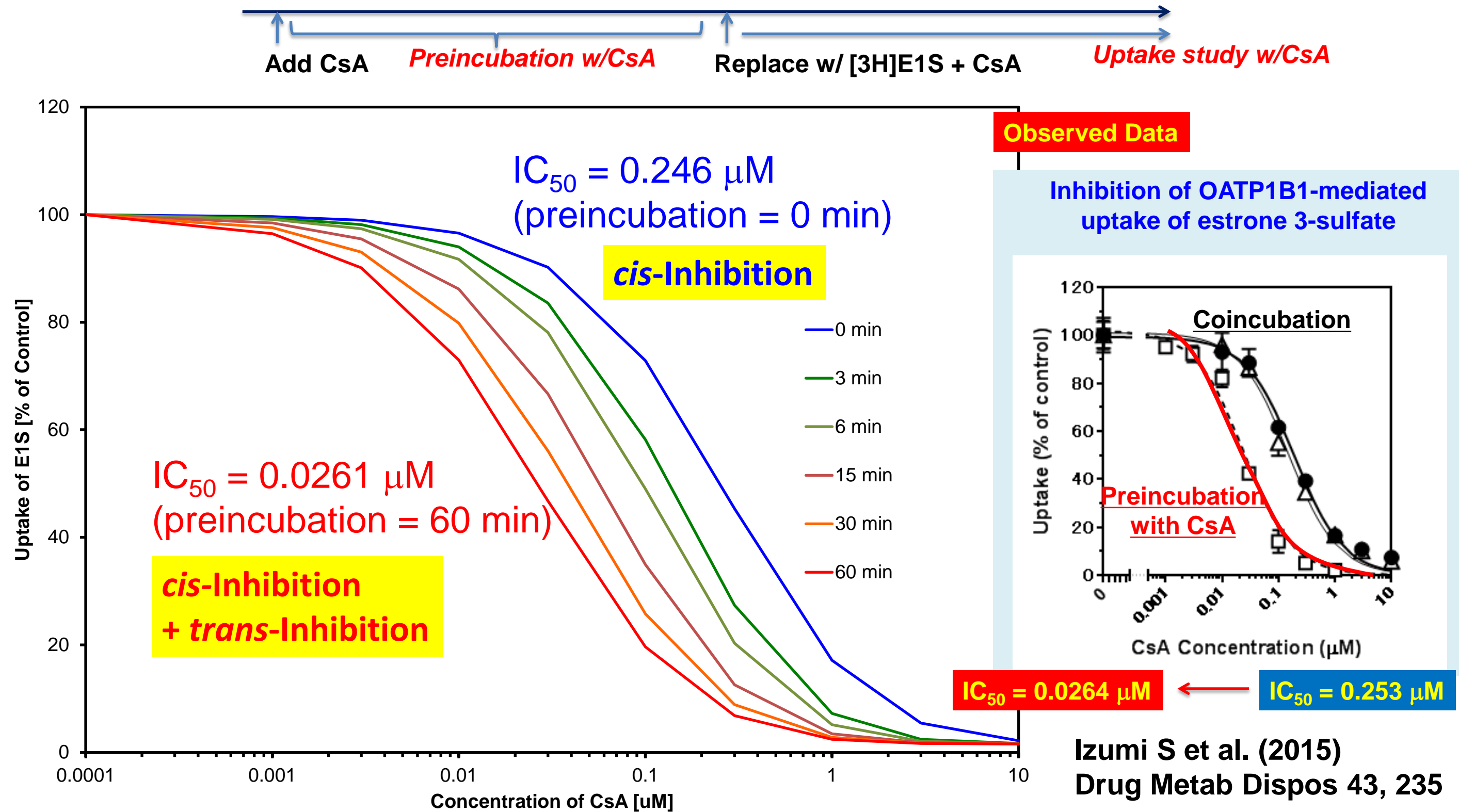
$$CL_{\text{uptake}}(+I) = \frac{V_{\text{max}} / (1 + f_{u,H} * I_H / K_{i,in})}{K_m * (1 + f_{u,B} * I_{EH} / K_{i,out}) + S}$$

*trans*-Inhibition (non-competitive)

*cis*-Inhibition (competitive)

\* $I_H$ ,  $I_{EH}$ : inhibitor (CsA) concentrations in the liver and at the extracellular space of the liver, respectively

# Simulation analysis of time-dependent enhancement effect of inhibition of OATP1B1 by CsA



$$\frac{1}{(1 + I_{out}/IC_{50,app})} = \frac{1}{[(1 + I_{out}/K_{i,out}) \times (1 + I_{u,in,unbound}/K_{i,in})]}$$

:with respect to time  $I_{u,in} \uparrow, K_{i,in} \rightarrow \Rightarrow IC_{50,app} \downarrow$

# **(1) A Systematic In Vitro Investigation of the Inhibitor Preincubation Effect on Multiple Classes of Clinically Relevant Transporters**

Péter Tátrai, Patrick Schweigler, Birk Poller, Norbert Domange, Roelof de Wilde, Imad Hanna,  
Zsuzsanna Gáborik, and Felix Huth

Solvo Biotechnology, Budapest, Hungary; Novartis Institutes for Biomedical Research, Basel,  
Switzerland and Novartis Institutes for Biomedical Research, East Hanover, New Jersey

**Drug Metab Dispos** 47:768–778, 2019

OATP1B1, OATP1B3, OAT1, OAT3, OCT1, OCT2, MATE1, and MATE2-K

**(Some inhibitors exhibited more than 200 folds decrease in apparent IC50 value by pre-incubation)**

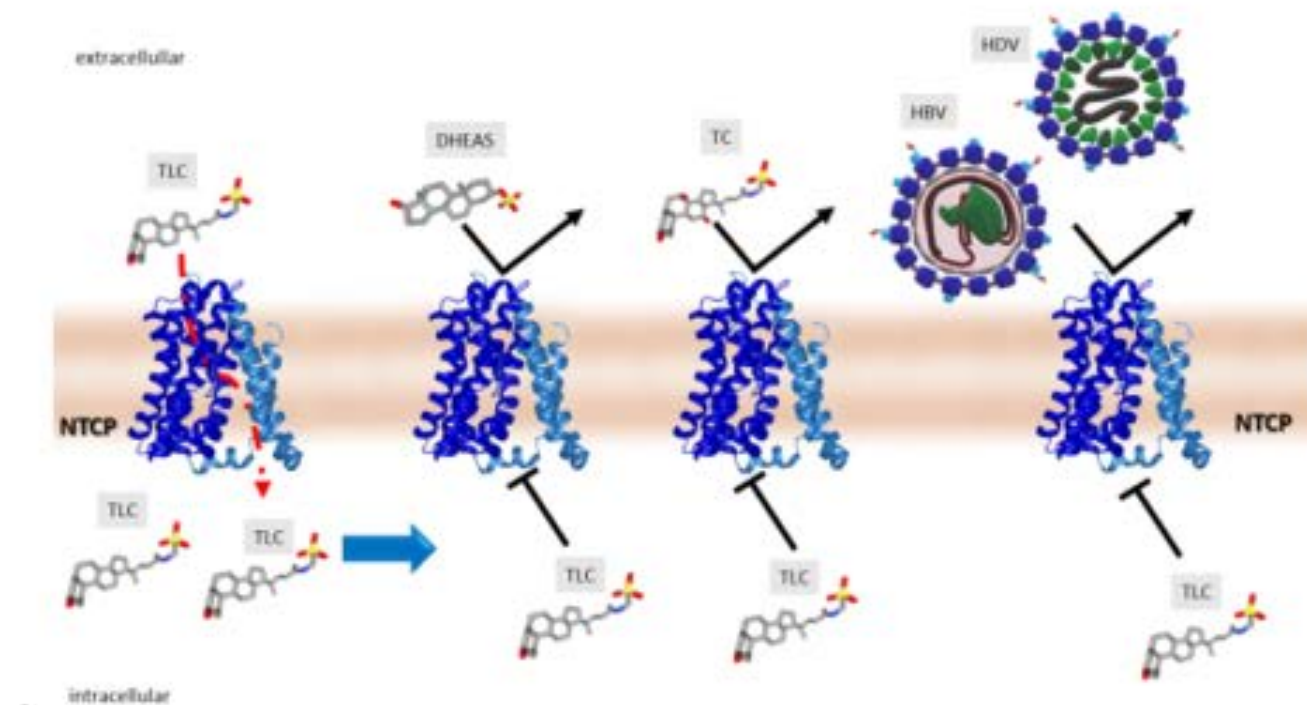
## **(2) Long-term *trans*-inhibition of the hepatitis B and D virus receptor NTCP by tauro lithocholic acid**

# Long-term *trans*-inhibition of the hepatitis B and D virus receptor NTCP by tauroolithocholic acid

Kira AAT Lowjaga, Michael Kirstgen, Simon F Müller, Nora Goldmann, Felix Lehmann, Dieter Glebe, Joachim Geyer

*Institute of Pharmacology and Toxicology, Faculty of Veterinary Medicine, Justus Liebig University Giessen, Germany*

**Am J Physiol Gastrointest Liver Physiol 320: G66–G80, 2021.**



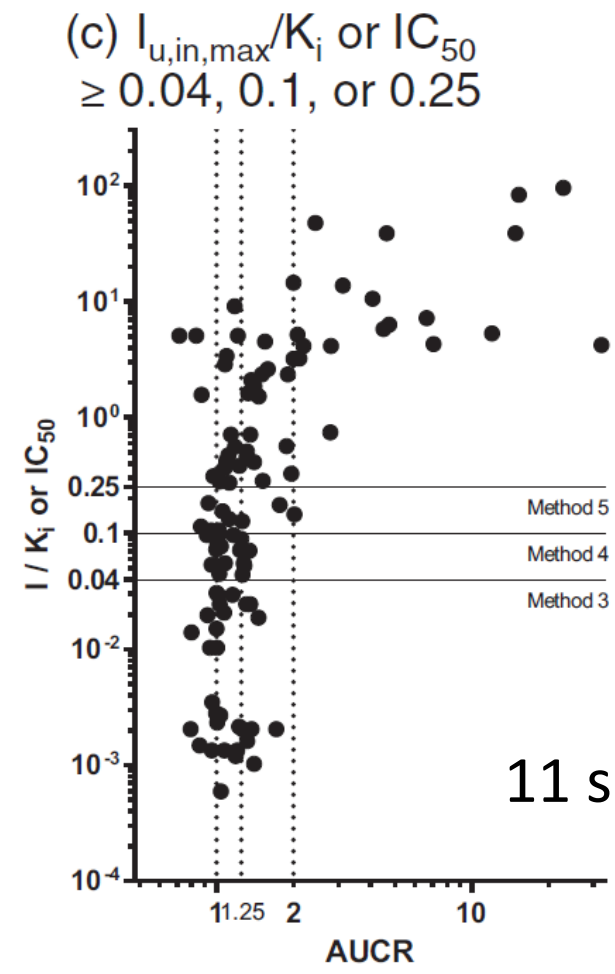
## What is the best translational approach until the mechanism is fully elucidated?

- 1) Benefit in using 'shifted' IC50 in the PBPK models as conservative approach?  
(Current recommendation by regulations)
- 2) More mechanism based PBPK modeling; distribution of inhibitor from extracellular space to intracellular sites  $\Rightarrow$  estimation of Permeability clearance, tissue binding,  $K_{i,in}$ ,  $K_{i,out}$   $\Rightarrow$  estimation of inhibiting effects which are changed with preincubation-time

# Contents

- 1) Introduction;  
Rate-determining process (focusing on the liver)  
(Uptake, efflux, elimination, metabolism) DDI
- 2) **PGx of OATP1B1: PBPK model based analysis**
- 3-1) **PBPK model based analysis of OATPs mediated drug-drug interaction**  
(Top down + Bottom-up)
  - (i) victim drugs-perpetrator drugs
  - (ii) endogenous biomarker (CP-I) – rifampicin**
- 3-2) Simple bottom-up predictions do not always work well.
  - (i) prediction of hepatic clearance of highly protein bound drugs  
(albumin-mediated hepatic uptake mechanisms should be considered)
  - (ii) Time-dependent inhibition (inhibitors for OATP1B and OCTs)
- 4) Target-mediated drug disposition (TMDD); To obtain dose-dependent change in molecular target occupancies only from the plasma concentration time-profile

# Concerns about the clinical DDI assessment: R-value



$$R\text{-value} = 1 + I_{u, in, max} / K_i \text{ or } IC_{50}$$

$$I_{u, in, max} = f_u * I_{max} + (k_a \times Dose \times F_a * F_g / Q_h)$$

If  $F_a * F_g$  values and  $k_a$  values are unknown, use 1 and  $0.1 \text{ min}^{-1}$   
 If  $f_u$  values are  $<0.1$  or undetermined, assume  $f_u = 0.01$

11 substrates and 61 inhibitors (total 106 studies)

Many false positive prediction

Method	1: $I_{max}/K_i \geq 0.1$	2: $I_{u,max}/K_i \geq 0.02$	EMA $R \geq 1.04$	4: $R \geq 1.1$	PMDA $R \geq 1.25$	FDA $R \geq 1.25$ $I_{max}/K_i \geq 0.1$
FN	12	17	8	12	15	17
FP	27	16	33	22	16	13
TN	28	39	22	33	39	42
TP	40	35	44	40	37	35
True positive rate	60%	69%	57%	65%	70%	73%
True negative rate	70%	70%	73%	73%	72%	71%

Vaidyanathan J et al., *J. Clin. Pharmacol.* 2016

**Current criteria may request pharmaceutical industry to conduct false positive DDI study, and overlook DDI risk.**

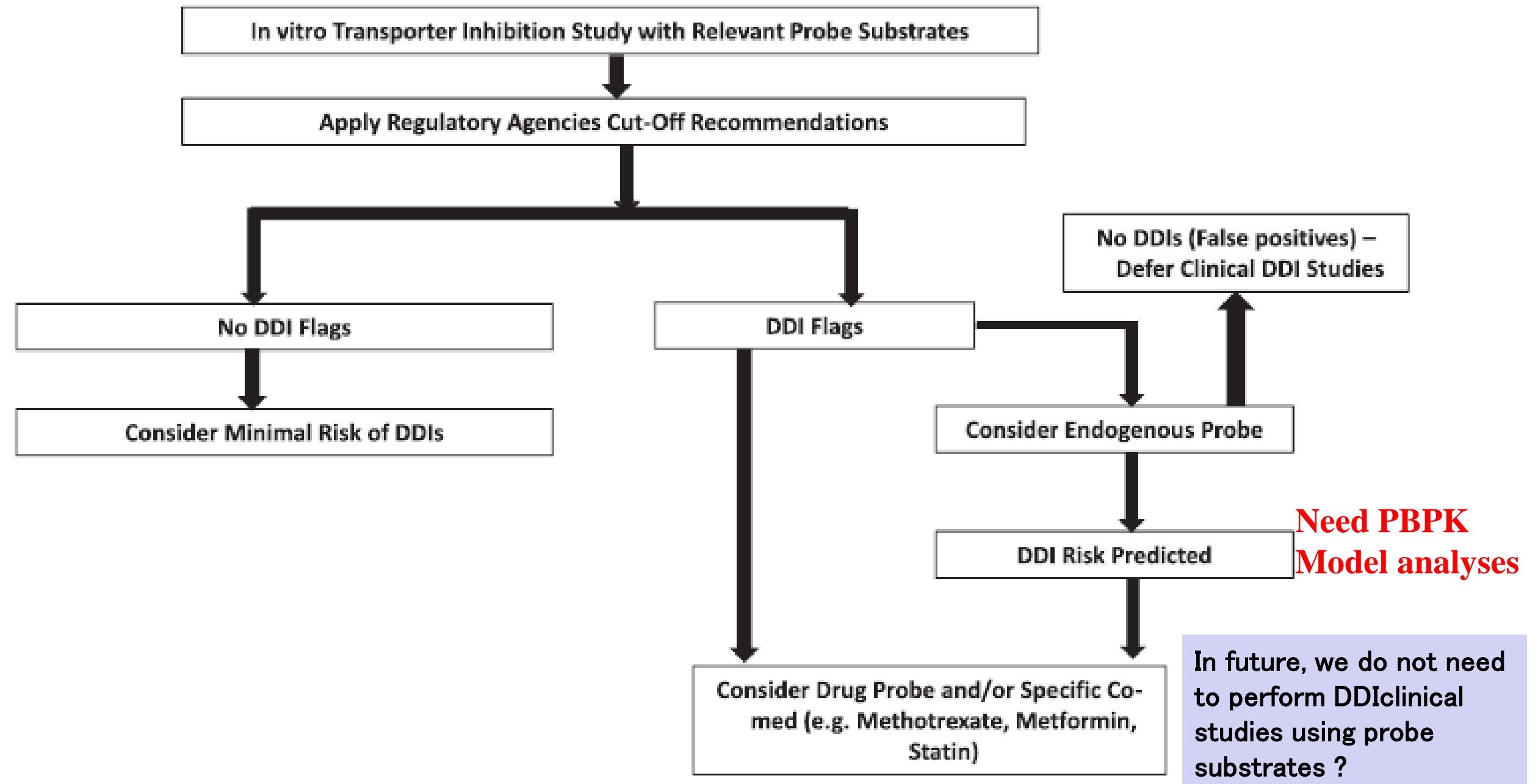
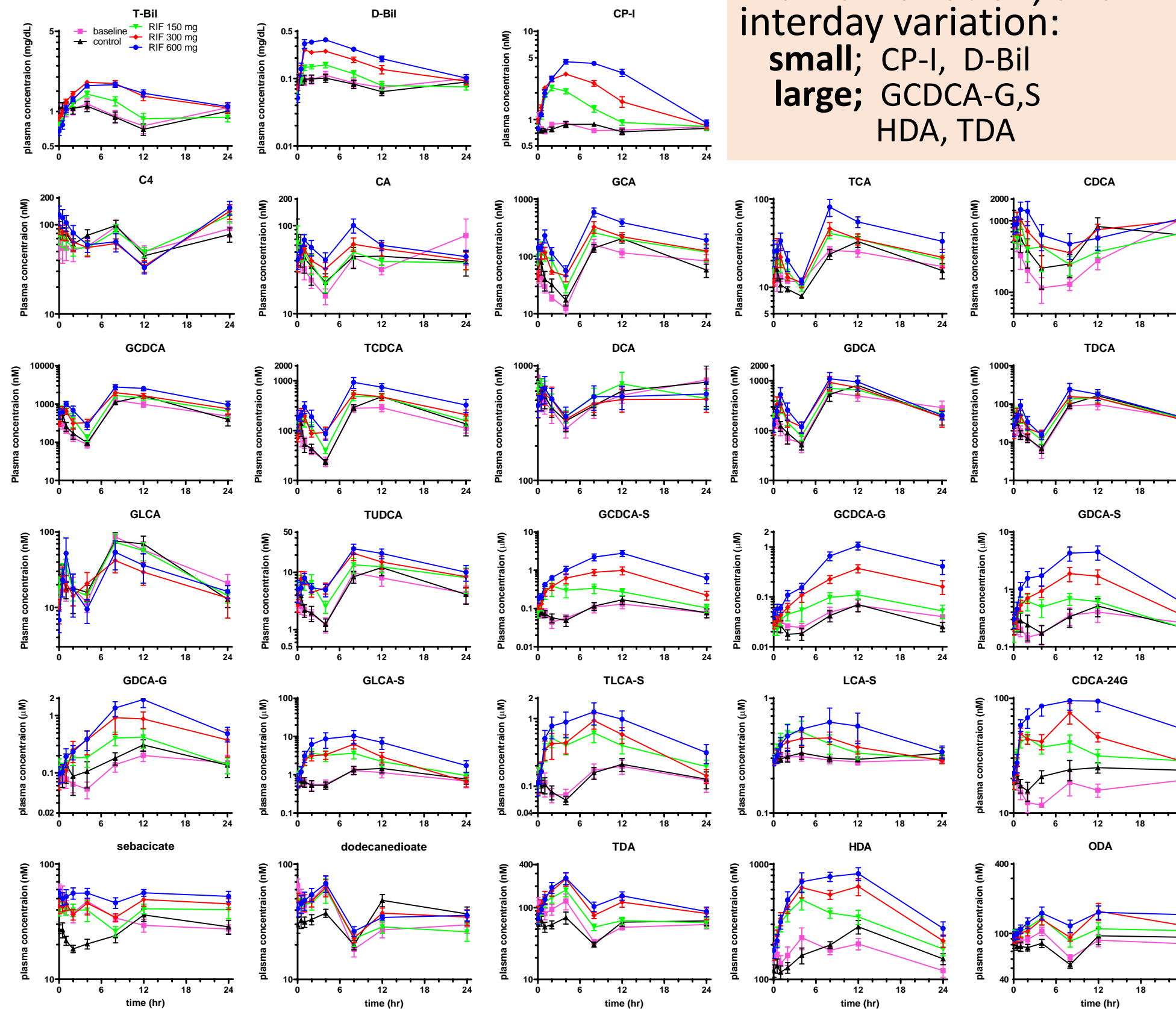


Figure 6 Potential inclusion of endogenous transporter markers in a regulatory decision tree. DDI, drug–drug interaction.

Rodrigues AD, Taskar KS, Kusuhara H, Sugiyama Y.  
**Clin Pharmacol Ther.103(3):434-448(2018)**



Diurnal variation, and interday variation:  
**small;** CP-I, D-Bil  
**large;** GCDCA-G,S  
 HDA, TDA



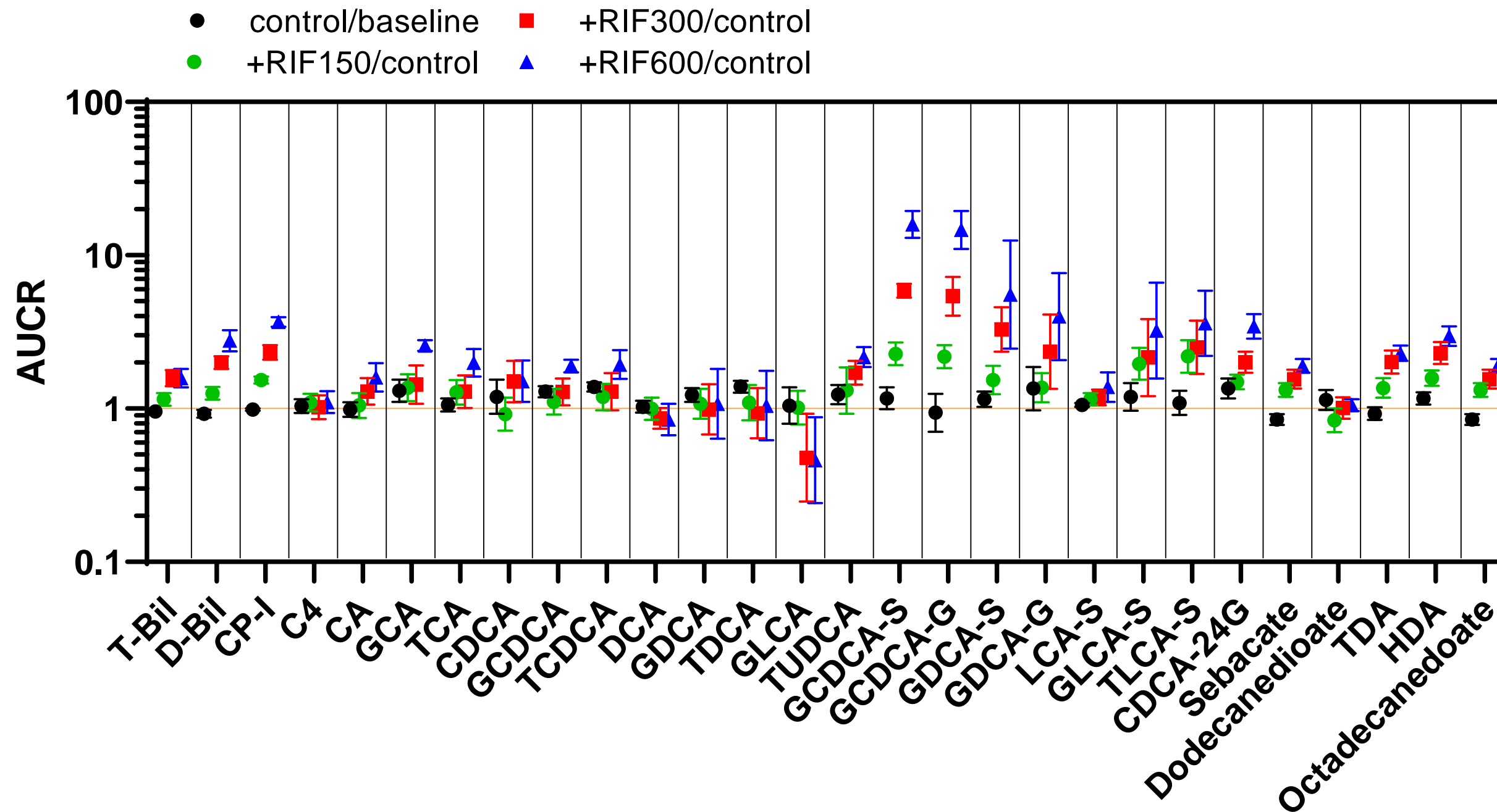
C4, 7-hydroxy-4-cholesten-3-one;  
 CA, cholate; CDCA, chenodeoxycholate; CDCA-24G,  
 chenodeoxycholate-24-glucuronide;  
 CP-I, coproporphyrin I;  
 D-Bil, direct bilirubins; DCA, deoxycholate;  
 GCDCA-G, glycochenodeoxycholate-3-glucuronide;  
 GCDCA-S, glycochenodeoxycholate-3-sulfate;  
 GDCA-G, glycodeoxycholate-3-glucuronide;  
 GDCA-S, glycodeoxycholate-3-sulfate;  
 HDA, hexadecanedioic acid;  
 LCA-S, lithocholate-3-sulfate; RIF, rifampicin;  
 T-Bil, total bilirubins

Mori D, Kimoto E, Rago B, Kondo Y, King-Ahmad A,  
 Ramanathan R, Wood LS, Johnson JG, Le VH,  
 Vourvahis M, Rodrigues AD, Muto C, Furihata K,  
 Sugiyama Y, Kusuhara H.

Dose-Dependent Inhibition of OATP1B by  
 Rifampicin in Healthy Volunteers:  
 Comprehensive Evaluation of Candidate  
 Biomarkers and OATP1B Probe Drugs.  
 Clin Pharmacol Ther. 107(4):1004-1013 (2020).

## Effect of rifampicin on the plasma concentration time profiles of 4 probe drugs and 28 endogenous substrates

Plasma concentrations of the endogenous substrates were determined at designated times in healthy volunteers treated with or without an oral dose of rifampicin (150, 300 and 600 mg).

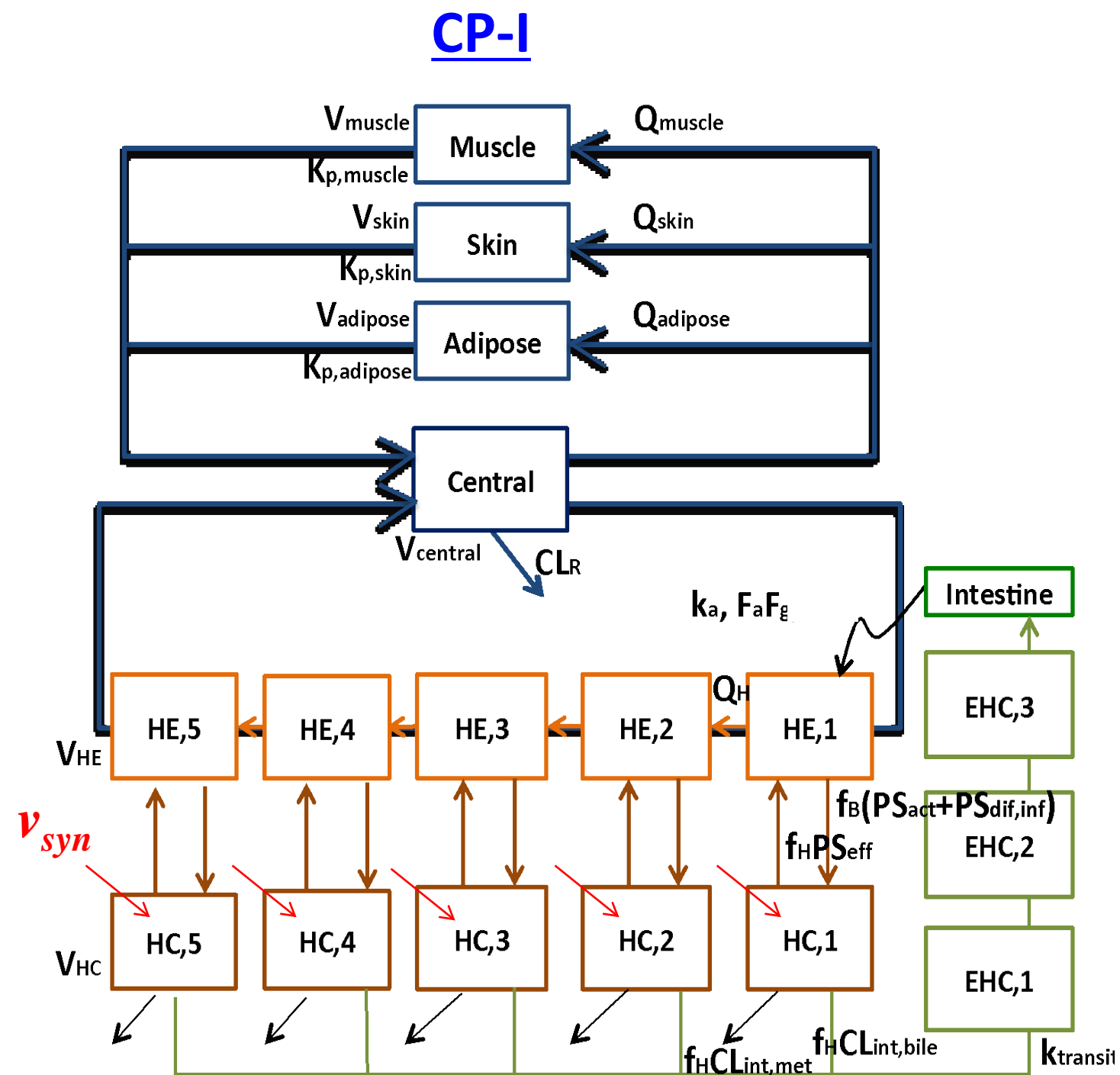


AUCR with 90% confidence interval assuming log normal distribution of various endogenous substrates with increasing dose of rifampicin. AUC<sub>0-24h</sub> was used to calculate AUCR.

Mori D, Kimoto E, Rago B, Kondo Y, King-Ahmad A, Ramanathan R, Wood LS, Johnson JG, Le VH, Vourvahis M, Rodrigues AD, Muto C, Furihata K, Sugiyama Y, Kusuhara H.

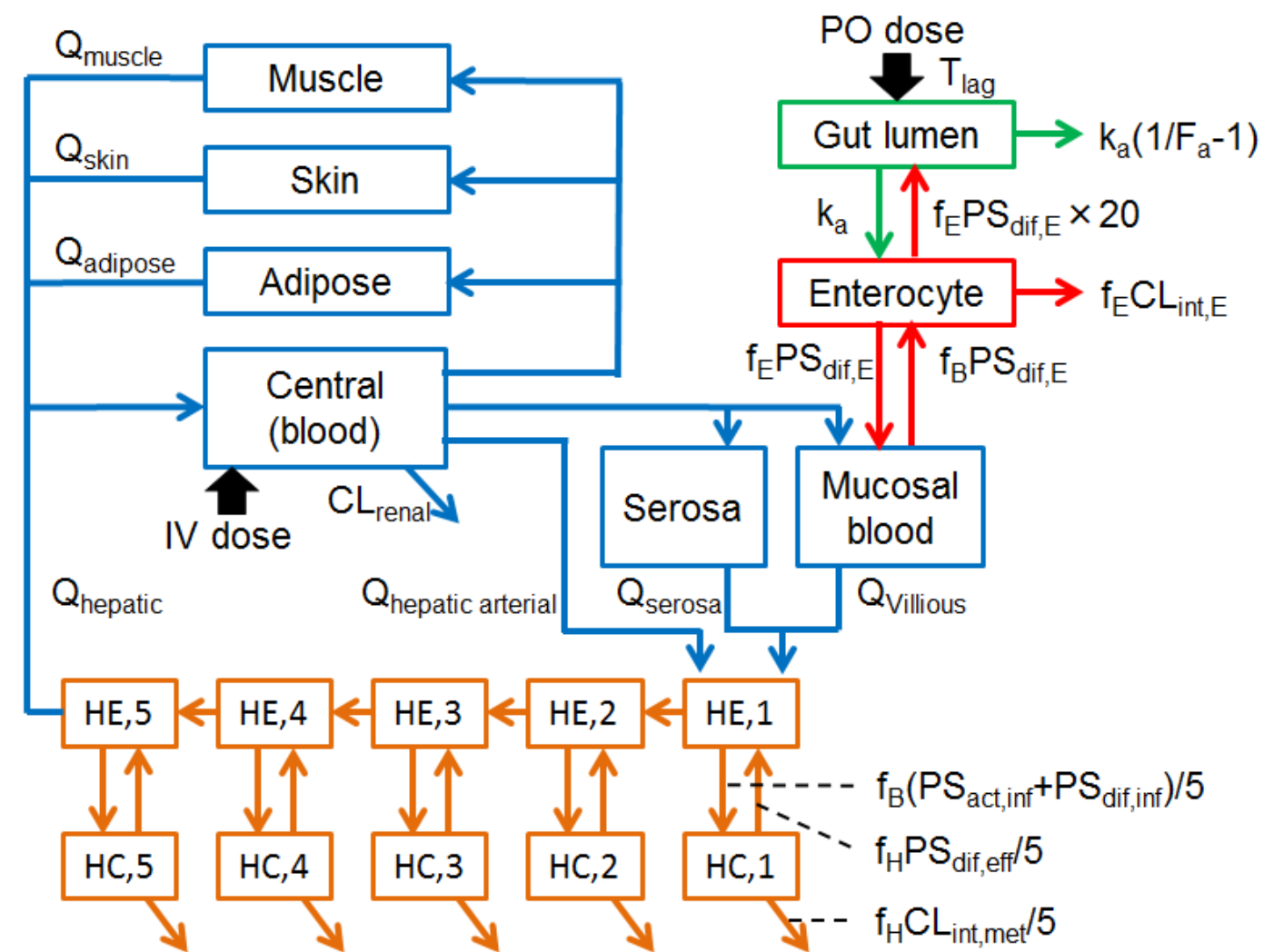
Dose-Dependent Inhibition of OATP1B by Rifampicin in Healthy Volunteers: Comprehensive Evaluation of Candidate Biomarkers and OATP1B Probe Drugs. *Clin Pharmacol Ther.* 107(4):1004-1013 (2020).

# Structure of PBPK models for CP-I and rifampicin



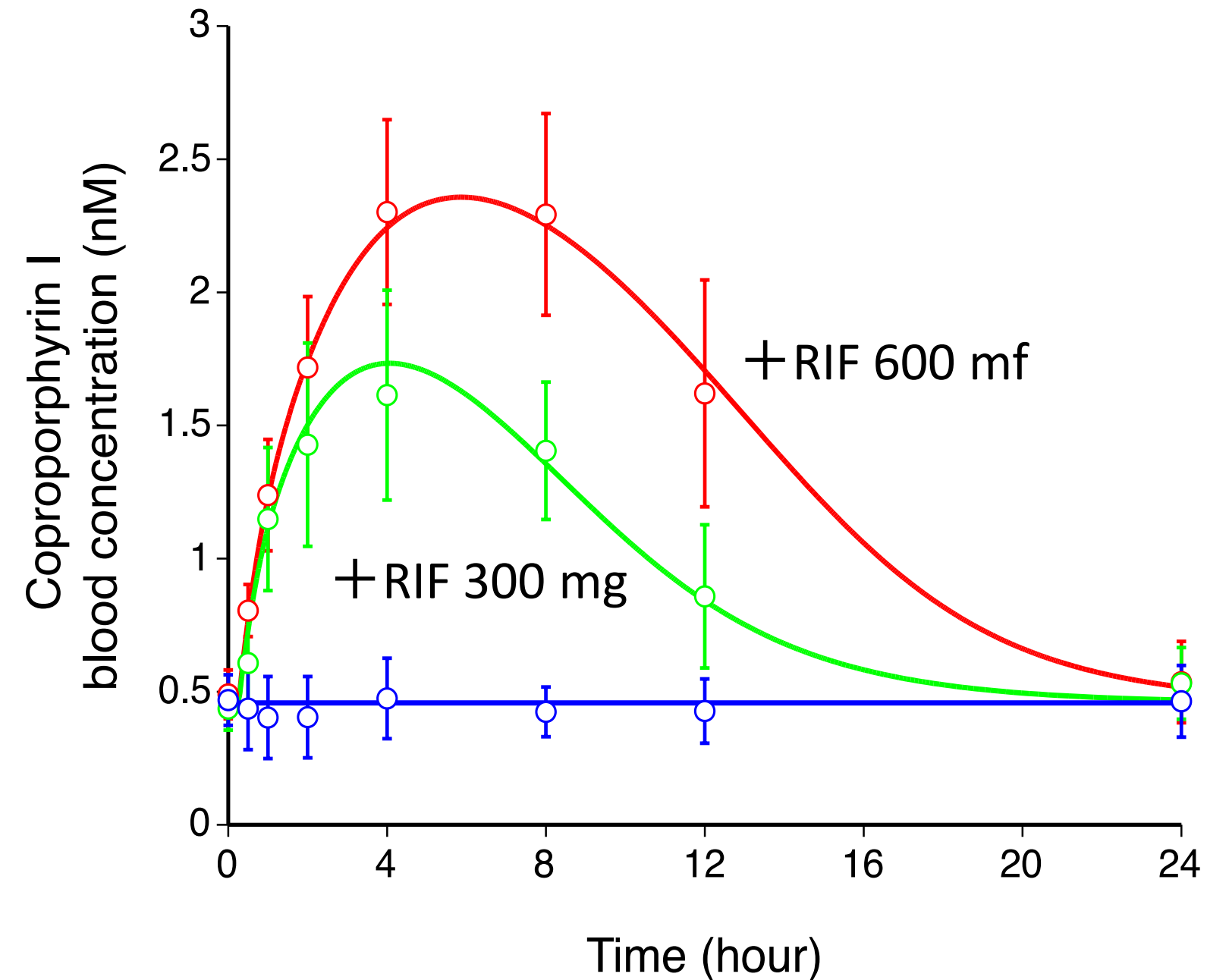
## Rifampicin

*Asaumi et al., CPT-PSP 7:186-196, 2018*



✓ The basic model structure for OATP1Bs substrates was reported previously (*Yoshikado et al., Clin Pharmacol Ther 100:513-523, 2016*).

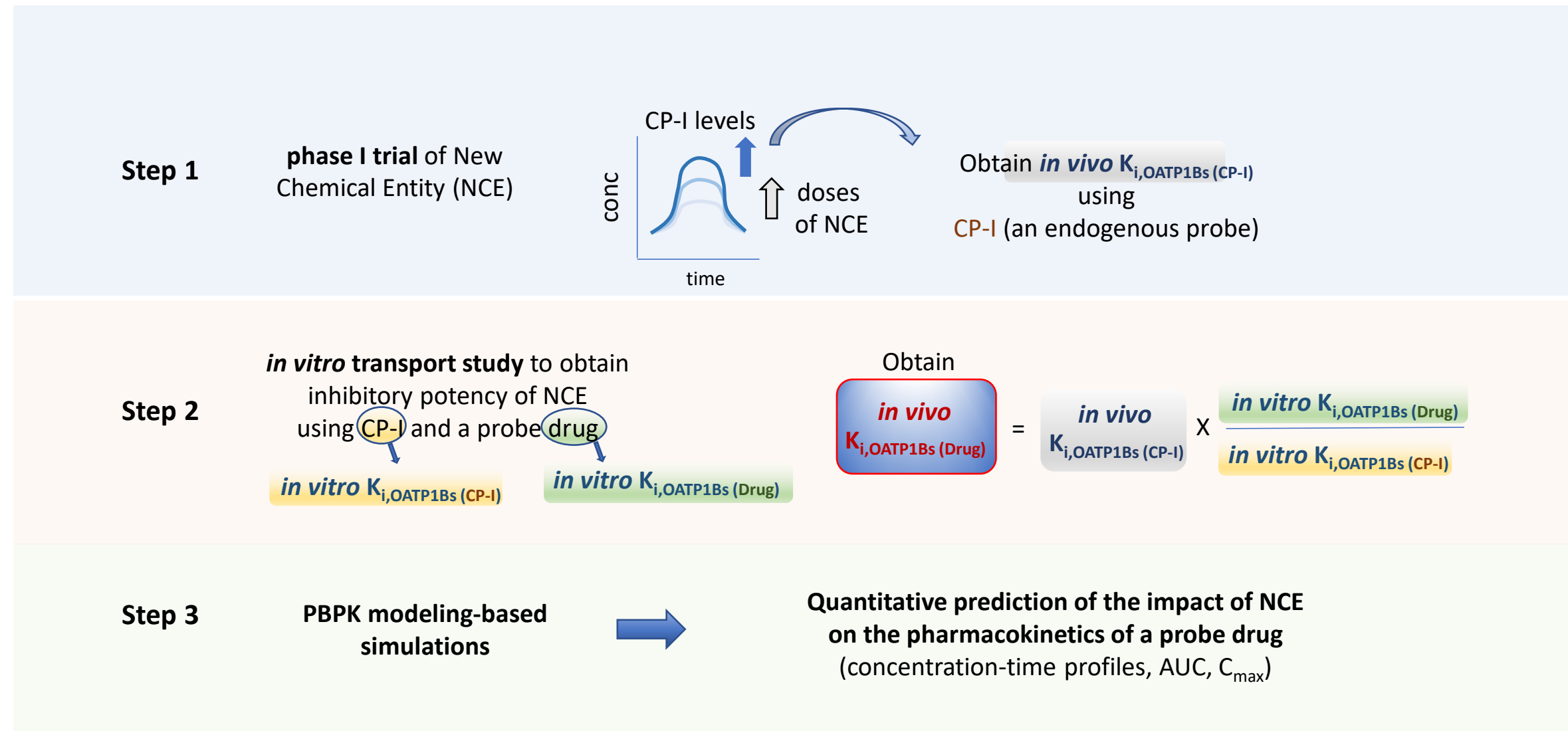
✓ The biosynthesis rate ( $v_{syn}$ ) of CP-I is incorporated.



**Simultaneously fitted blood concentration–time profiles of CP-I in the absence and presence of RIF after parameter optimization using the PBPK model incorporating **the inhibition of OATP1Bs and MRP2.****

Yoshikado T, Toshimoto K, Maeda K, Kusuhara H, Kimoto E, Rodrigues AD, Chiba K, Sugiyama Y. PBPK Modeling of Coproporphyrin I as an Endogenous Biomarker for Drug Interactions Involving Inhibition of Hepatic OATP1B1 and OATP1B3. *CPT Pharmacometrics Syst Pharmacol.* 7(11):739-747 (2018).

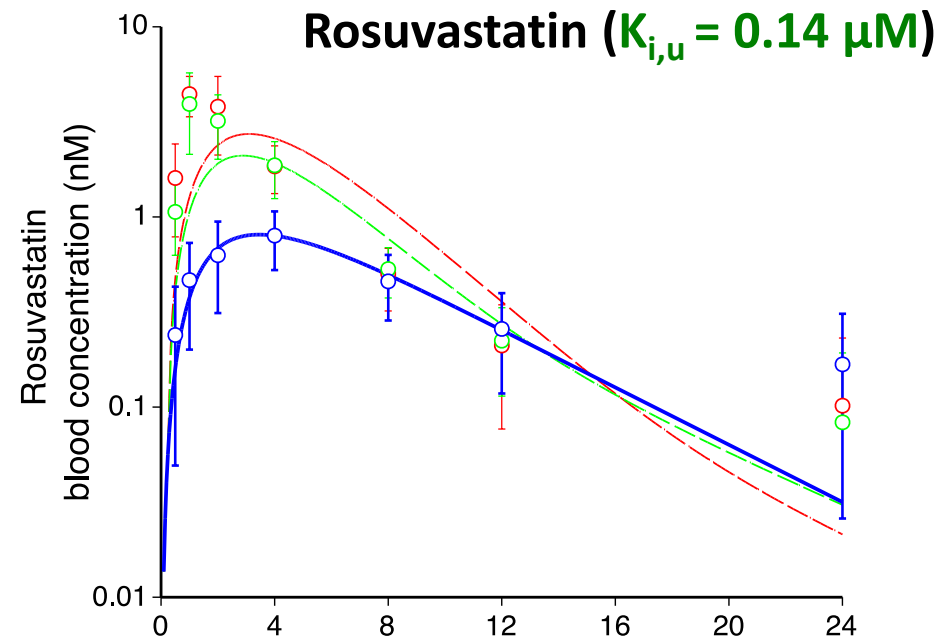
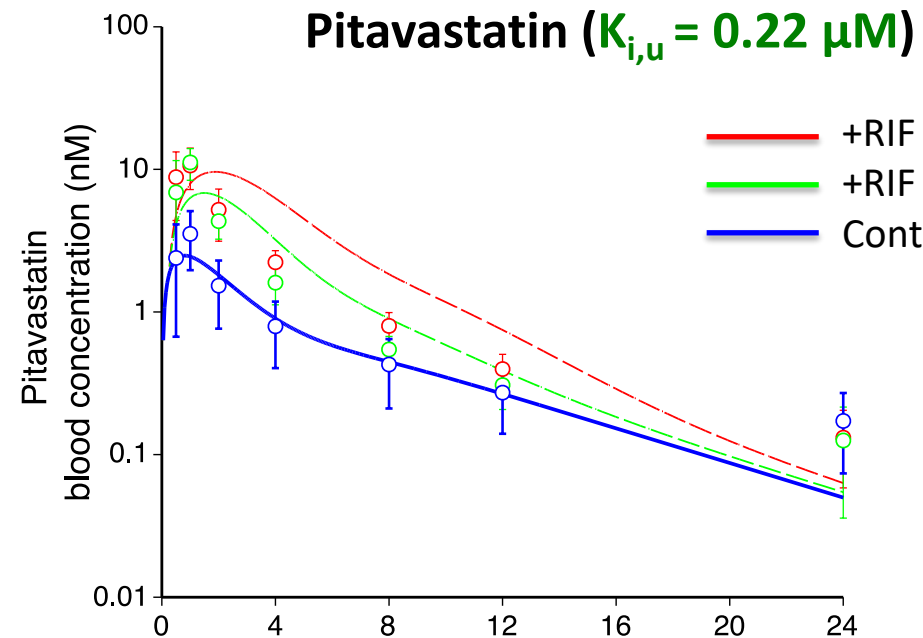
# Strategy to predict DDI for a probe substrate using CP-I as an endogenous biomarker



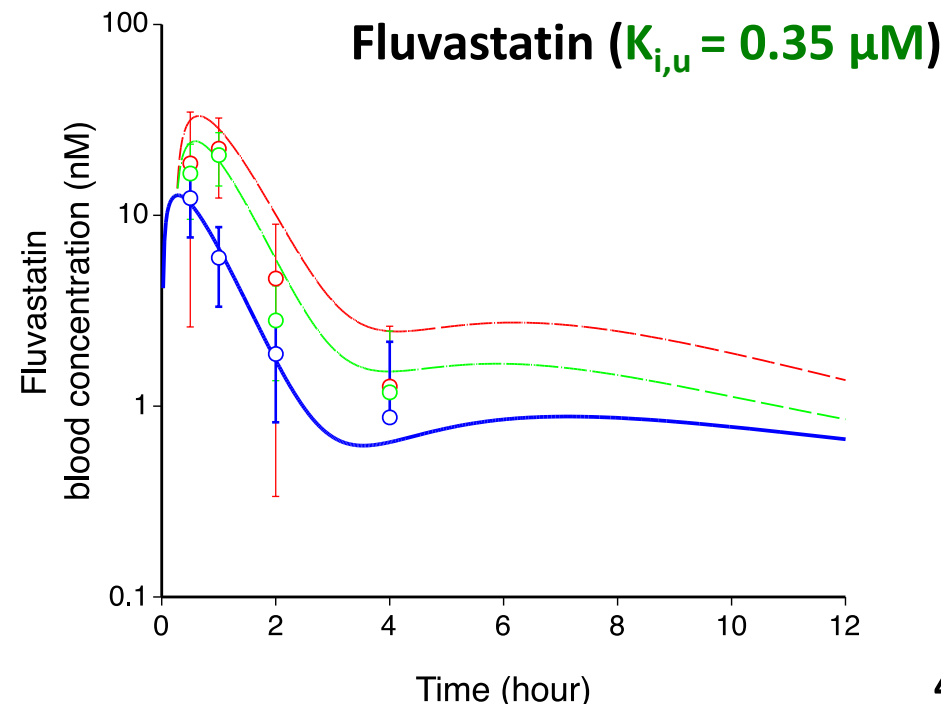
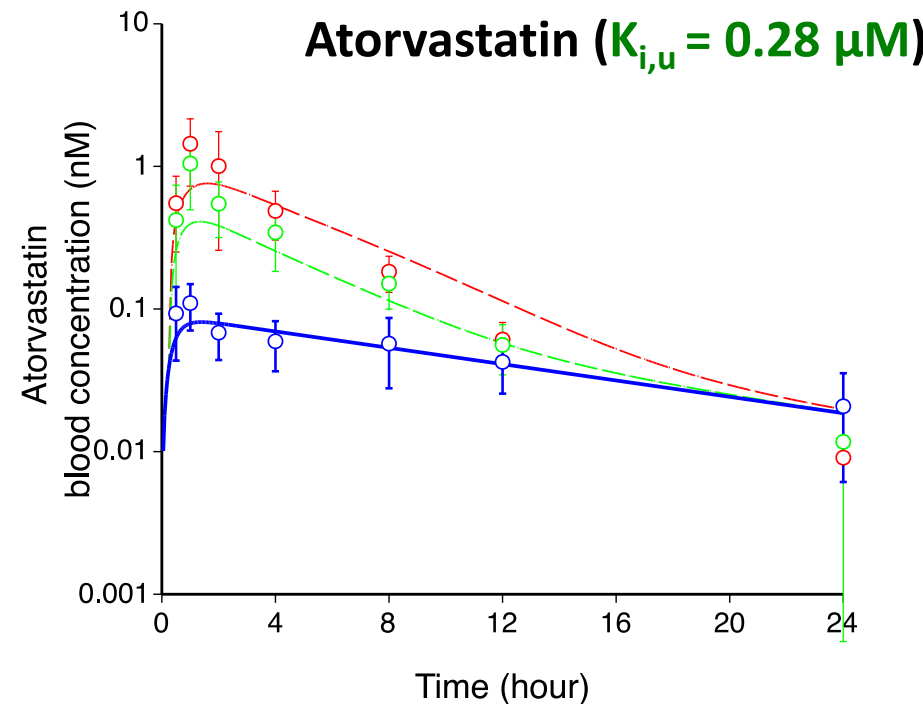
Yoshikado T, Toshimoto K, Maeda K, Kusuhara H, Kimoto E, Rodrigues AD, Chiba K, Sugiyama Y. [PBPK Modeling of Coproporphyrin I as an Endogenous Biomarker for Drug Interactions Involving Inhibition of Hepatic OATP1B1 and OATP1B3.](#)

CPT Pharmacometrics Syst Pharmacol. 7:739-747 (2018)

Prediction of the effect of RIF on blood concentration-time profiles of statins  
 (Correction of in vivo  $K_{i,uOATP1B_S}$  based on substrate-dependent difference of in vitro  $K_{i,u}$ )



Observations: *Takehara I et al., Pharm Res., 35:138 (2018)*

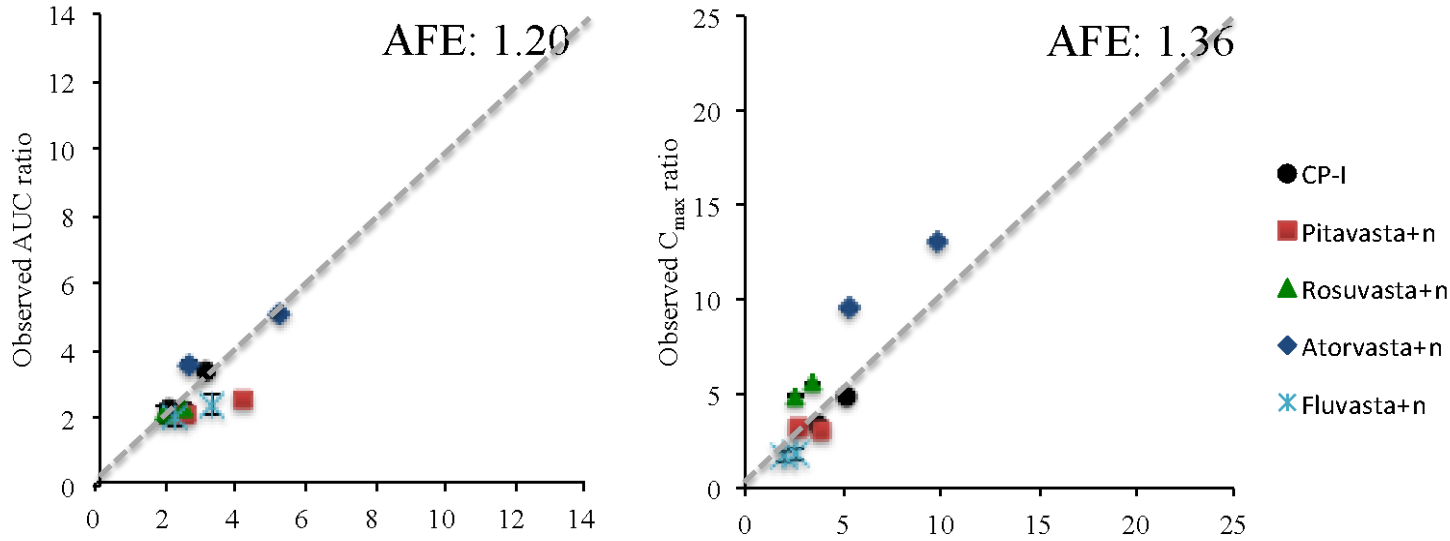


# Predicted and observed AUC ratios and $C_{max}$ ratios for statins using our PBPK models

## With Taking substrate-dependent $K_{i,u,OATP1Bs}$ into consideration

Upper panel

$K_i(\text{CP-I}) \neq K_i(\text{statins})$



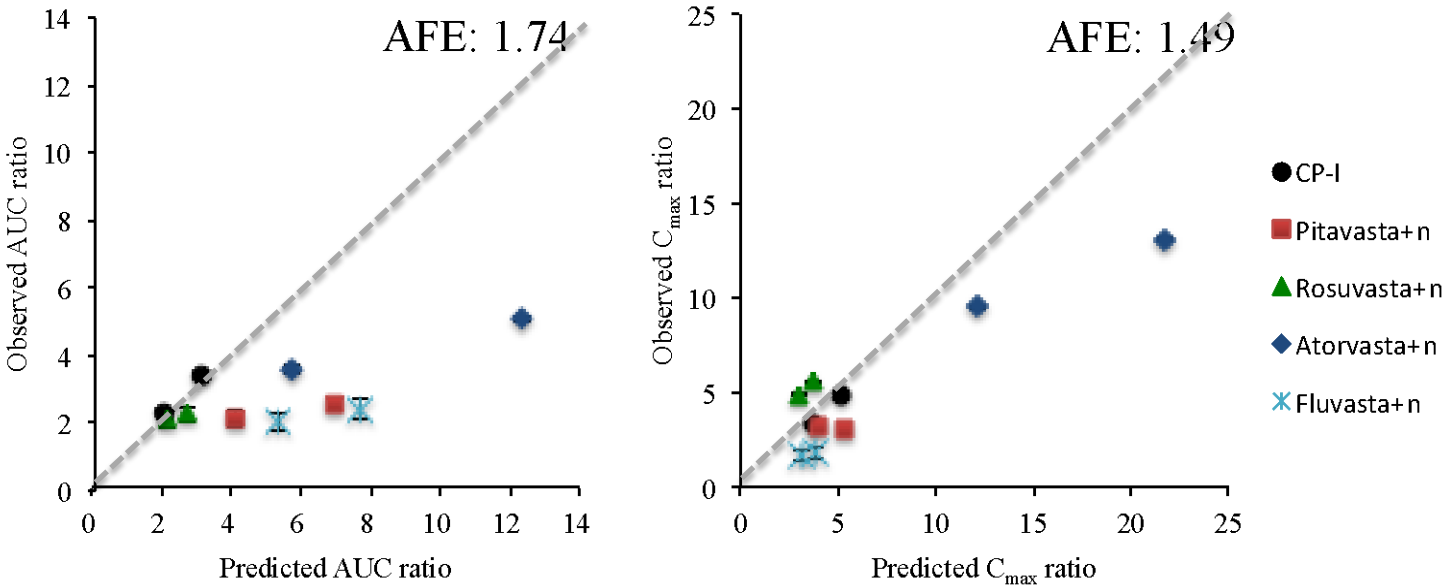
Observations: *Takehara I et al., Pharm Res., 35:138 (2018)*

AFE:  
Average fold errors

## Without taking substrate-dependent $K_{i,u,OATP1Bs}$ into consideration

Lower panel

$K_i(\text{CP-I}) = K_i(\text{statins})$



# Summary

The PBPK modeling approach provides an insightful understanding of the mechanisms governing changes in the plasma conc. of an endogenous biomarker (CP-I) for OATP1Bs and MRP2 (CP-I) and enables complex analyses of the dose-dependent inhibitory effects of RIF on the hepatic OATP1Bs/MRP2-mediated transport of CP-I.

It also lead to the successful prediction of RIF interaction with several probe substrates (statins) for OATP1Bs.

K<sub>i</sub> value for MRP2 (as a biliary excretion transporter of CP-I) cannot be determined only from the top-down analyses, and the value obtained by PET imaging analyses of different probe (TIC-Me (PGI<sub>2</sub> receptor imaging in the brain)) was used to obtain K<sub>i,MRP2</sub> (0.87  $\mu$ M).



# Perspective

Emphasize the importance of the use of multiple biomarkers (coproporphyrine I, bilirubin glucuronide and glycochenodeoxycholic acid sulfate) to assess the OATP1Bs mediated interaction of new NCE as perpetrators in their phase 1 clinical studies.

Some of recent manuscripts indicate the advantage and validity of the use of coproporphyrine I as a biomarker of OATP1Bs function in vivo. However, it is not so easy to confirm that your NCE does not modulate other transporters and enzymes which will be responsible for the biosynthesis, intestinal absorption and biliary excretion and renal clearance of this biomarker. In fact, rifampicin and cyclosporine which are well known to inhibit OATP1Bs mediated hepatic uptake are also known to affect P-gp/BCRP in the intestine, BSEP and MRP2/BCRP in the liver.

**Use of multiple biomarkers will ultimately increase the confidence in our prediction of clinical DDI using biomarkers from pharmaceutical and regulatory perspectives.**

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- 4) **Target-mediated drug disposition (TMDD); To obtain dose-dependent change in molecular target occupancies only from the plasma concentration time-profile**

# Target-mediated Drug Disposition (TMDD)

✓ Introduced by Dr. Gerhard Levy (1994)

✓ Type of **nonlinear PK**

✓ When drugs bind to a target with **high affinity** and **to a significant extent** (relative to dose), part of the initial dose is rapidly acquired by the target sites and only then the drug will distribute to other tissues.

Pharmacologic target-mediated drug disposition

Gerhard Levy, PharmD *Amherst, N.Y.*

(*Clin Pharmacol Ther*, 1994)

*Well-recognized with biologics  
(e.g, monoclonal antibodies),  
but earliest examples were in fact  
**small-molecule drugs**  
(more cases being reported/identified lately)*



Gerhard Levy, PharmD  
(1928-2017)

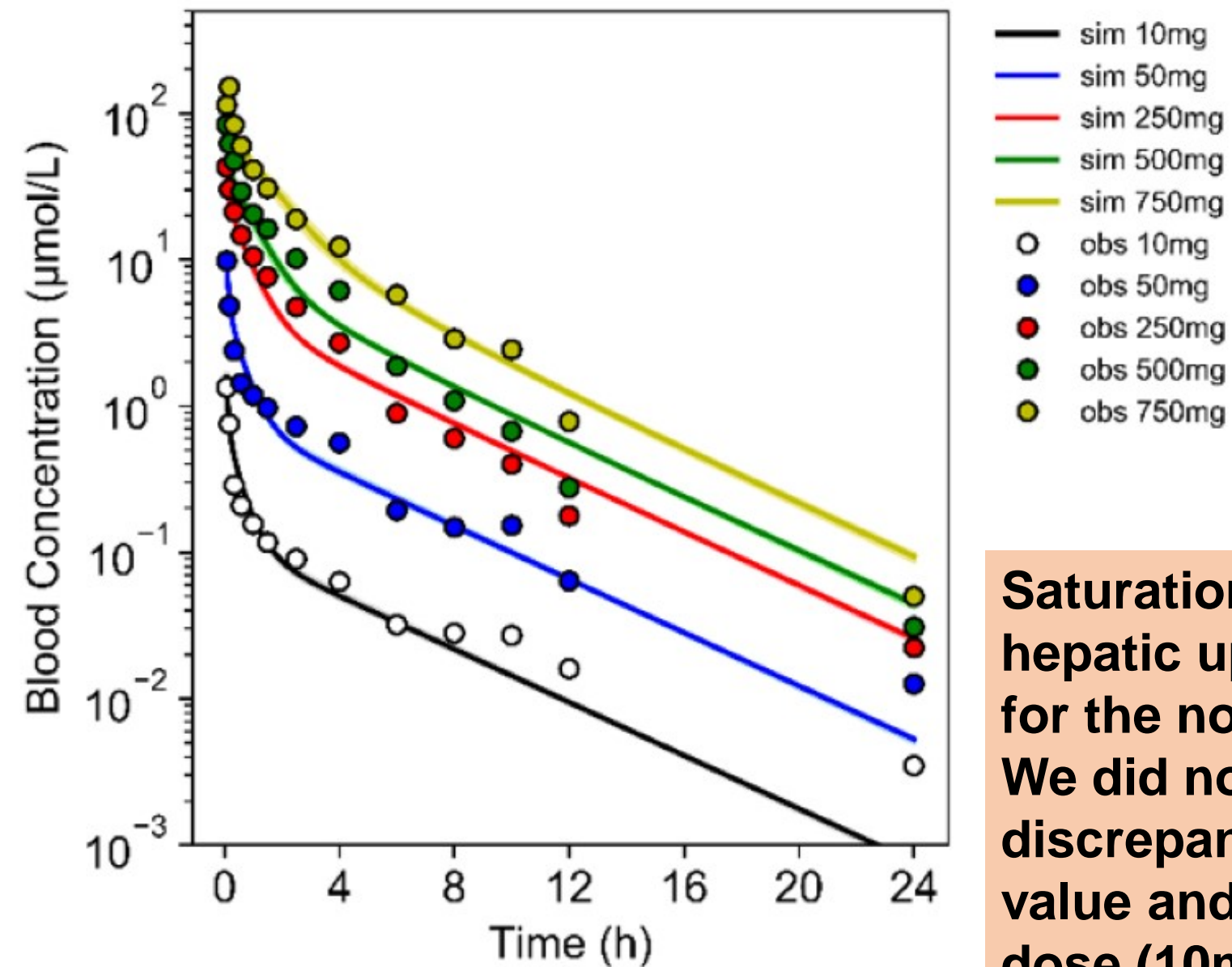
# Can the prediction of dose-dependent molecular target occupancy be possible from the phase-1 clinical studies ?

Just measuring the blood concentration of drugs over a wide range of dose without measuring the tissue concentration such as PET imaging

This is the methodology presented in a recent NRDD review(1) and in a recent original article on bosentan (2)

(1) Burt T, Young G, Lee W, Kusuhara H, Langer O, Rowland M, Sugiyama Y.  
Phase 0/microdosing approaches: time for mainstream application in drug development?  
Nat Rev Drug Discov. 19(11):801-818 (2020).

(2) Koyama S, Toshimoto K, Lee W, Aoki Y, and Sugiyama Y.  
Revisiting nonlinear bosentan pharmacokinetics by PBPK modeling: Target binding, albeit not a major contributor to nonlinearity, can offer prediction of target occupancy  
Drug Metab Dispos., in press



Volz A-K, Dingemans J, Krause A, and Lehr T (2019) Target-mediated population pharmacokinetic modeling of endothelin receptor antagonists. *Pharm Res* 37:2.

**Saturation of OATP1B mediated hepatic uptake mostly accounted for the non-linear PK of bosentan. We did not analyze such a discrepancy between observed value and fitted line at the lowest dose (10mg)**

## Our previous publication

**Sato M, Toshimoto K, Tomaru A, Yoshikado T, Tanaka Y, Hisaka A, Lee W, Sugiyama Y.**

Physiologically Based Pharmacokinetic Modeling of Bosentan Identifies the Saturable Hepatic Uptake as A Major Contributor to Its Nonlinear Pharmacokinetics. *Drug Metab Dispos.* 46(5):740-748 (2018).

(44)

## Recent studies on the analyses of non-linear PK of bosentan

Sato M, Toshimoto K, Tomaru A, Yoshikado T, Tanaka Y, Hisaka A, Lee W, and Sugiyama Y  
Physiologically based pharmacokinetic modeling of bosentan identifies the saturable hepatic uptake as a major contributor to its nonlinear pharmacokinetics.

Drug Metab Dispos 46: 740–748 (2018)

**(Major mechanism for non-linear PK: Hepatic uptake (OATP1B))**

Li R, Niosi M, Johnson N, Tess DA, Kimoto E, Lin J, Yang X, Riccardi KA, Ryu S, El-Kattan AF, et al.  
A study on pharmacokinetics of bosentan with systems modeling, part 1: translating systemic plasma concentration to liver exposure in healthy subjects.

Drug Metab Dispos 46: 346–356 (2018).

**(Major mechanism for non-linear PK: Multiple mechanism, OATP1B, CYPs, TMDD)**

Volz A-K, Krause A, Haefeli WE, Dingemans J, and Lehr T:

Target-mediated drug disposition pharmacokinetic–pharmacodynamic model of bosentan and endothelin-1.

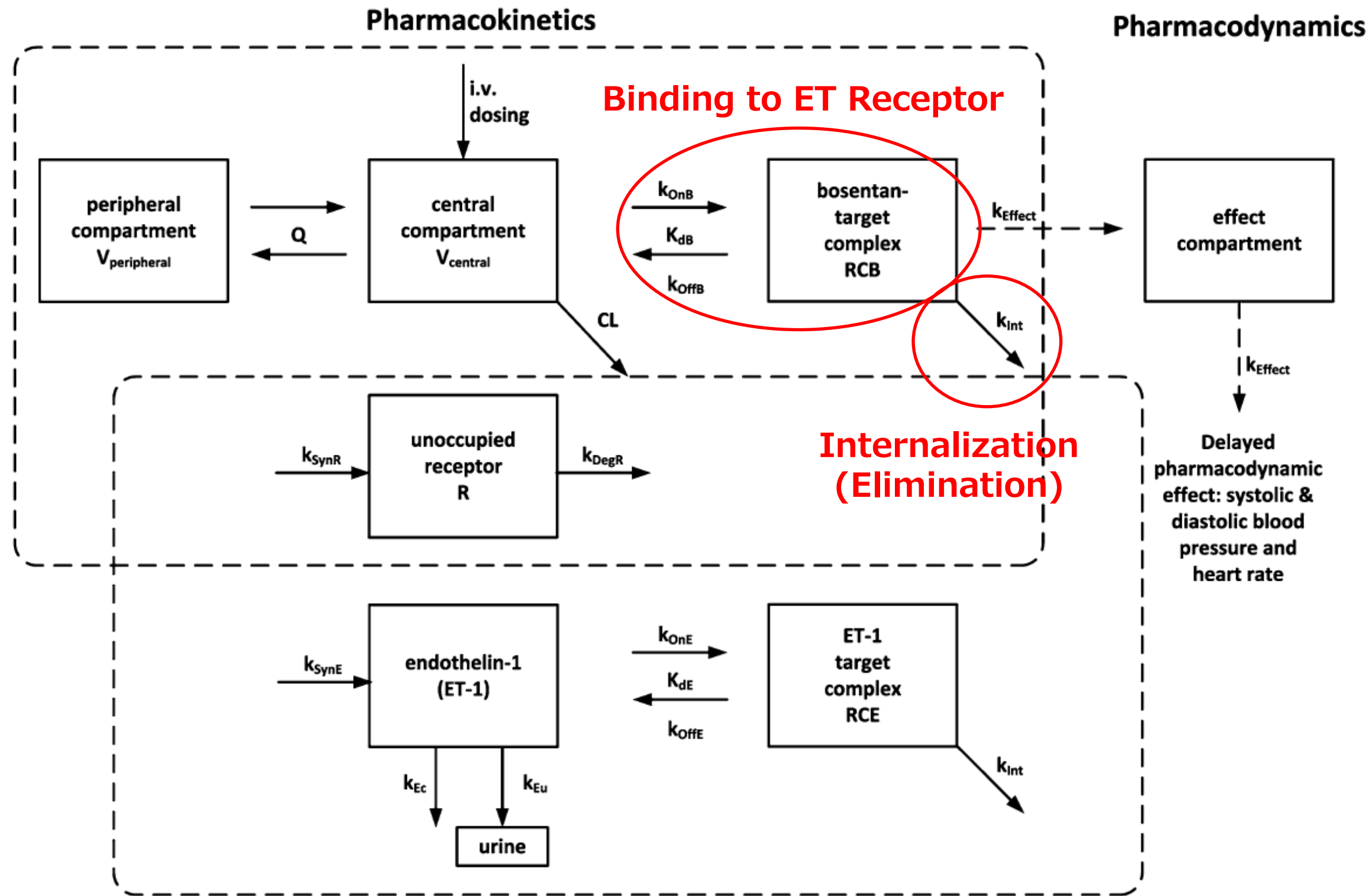
Clin Pharmacokinetics 56:1499–1511 (2017).

Volz A-K, Dingemans J, Krause A, and Lehr T :

Target-mediated population pharmacokinetic modeling of endothelin receptor antagonists.

Pharm Res 37:2 (2019) .

**(Major mechanism for non-linear PK: Target binding followed by the internalization(TMDD))**



- ◆ Population PK/PD Model considering only TMDD.
- ◆ There is no evidence that bosentan can be internalized and eliminated after binding to ET receptor. (Endothelin can be internalized and degraded.)

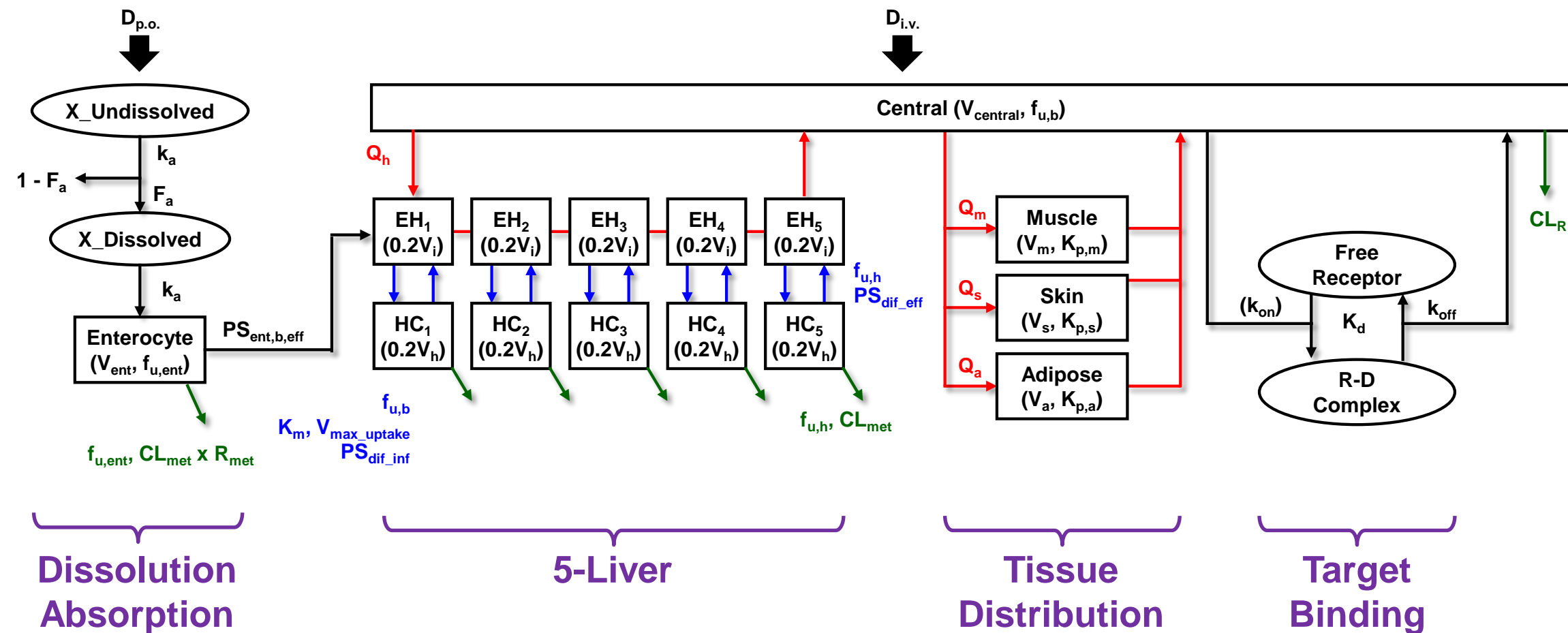
# PBPK Model Structure of Bosentan

Koyama S, Toshimoto K, Lee W, Aoki Y, and Sugiyama Y.

Revisiting nonlinear bosentan pharmacokinetics by PBPK modeling:

Target binding, albeit not a major contributor to nonlinearity, can offer prediction of target occupancy

Drug Metab Dispos., 49:298-304 (2021)



- ◆ Combined dissolution/absorption, 5 liver, tissue distribution, and target binding.
- ◆ Target binding is directly connected to central compartment.
- ◆ Considered saturable hepatic uptake and target binding, and non-saturable metabolism.
- ◆ Assuming linear and dissolution-rate limited absorption.



# Cluster Gauss-Newton method in comparison to conventional methods

## Conventional methods

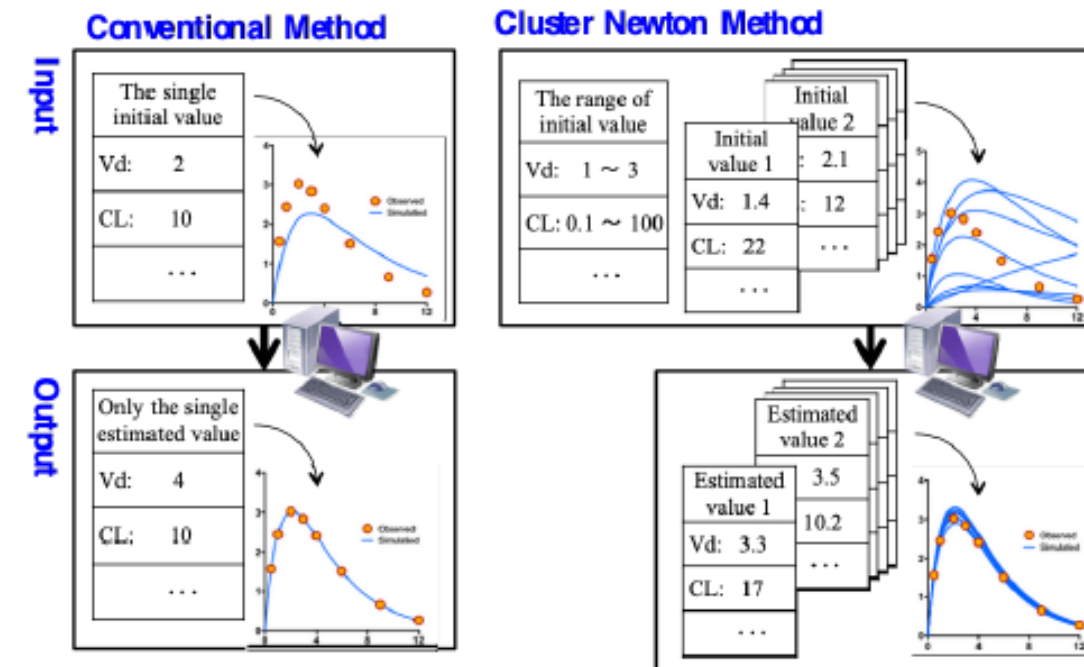
- Requires **appropriate initial iterate** for the parameters.
- Obtains **only a single set** of optimised solutions.
- Computationally **expensive**.
  - Need extra model evaluations (e.g. for derivative computation)
  - Need to restart with different initial parameters

**Requires a lot of experience!**

## Cluster Gauss-Newton method

- Only requires **setting wide ranges** of initial iterates.
- Obtains **multiple sets** of optimised solutions.
- Computationally **cheap and robust**.
  - No need for extra model evaluation.
  - Can estimate many unknown parameters in a complex model

**Requires less experience.**



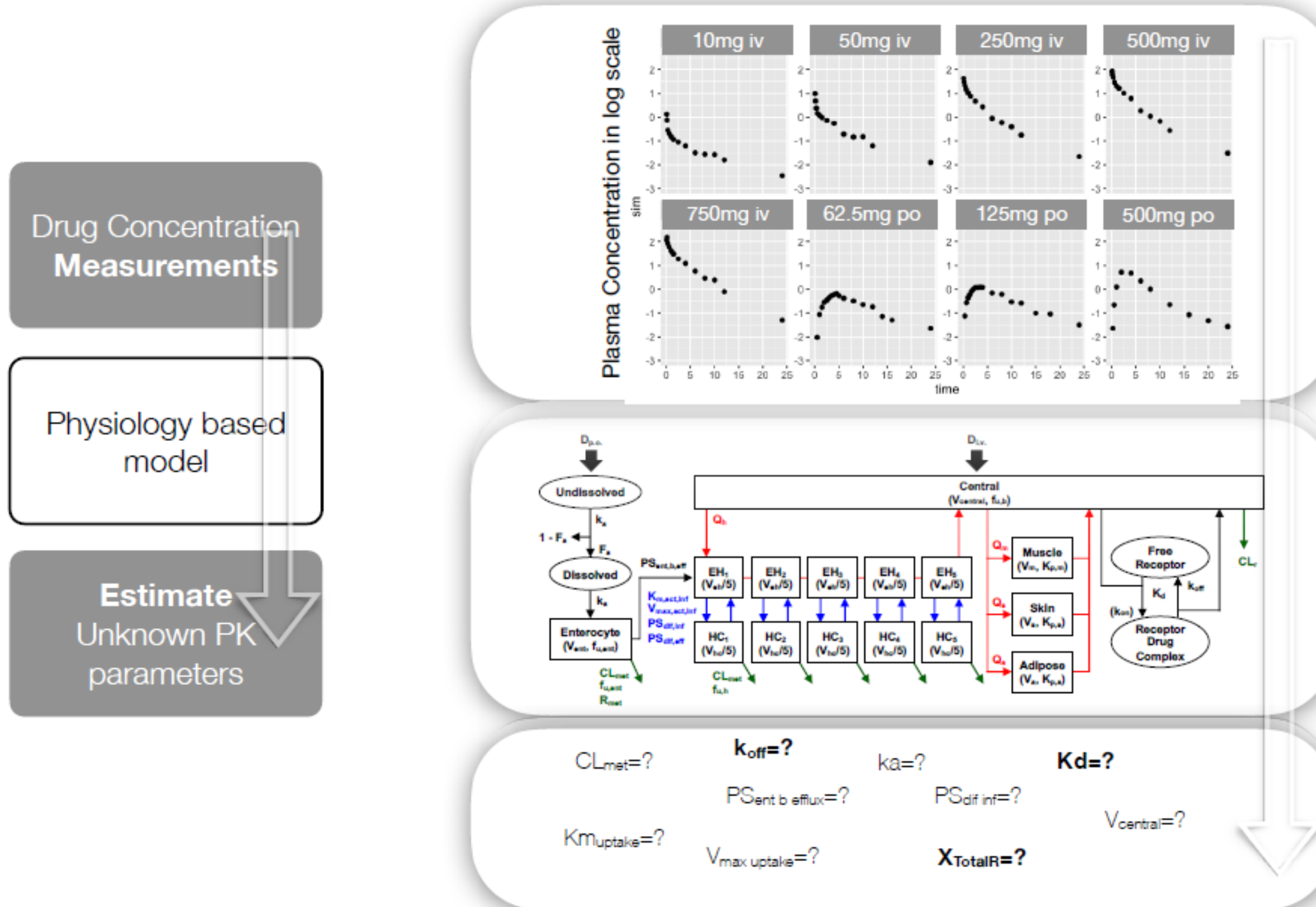
Aoki, Y., Hayami, K., Toshimoto, K., & Sugiyama, Y. (2020). Cluster Gauss–Newton method. *Optimization and Engineering*, 1-31.

**Aoki Y, Hayami K, Toshimoto K, Sugiyama Y. (2020)**

Cluster Gauss–Newton method; An algorithm for finding multiple approximate minimisers of nonlinear least squares problems with applications to parameter estimation of pharmacokinetic models.

**Optimization and Engineering** <https://doi.org/10.1007/s11081-020-09571-2>

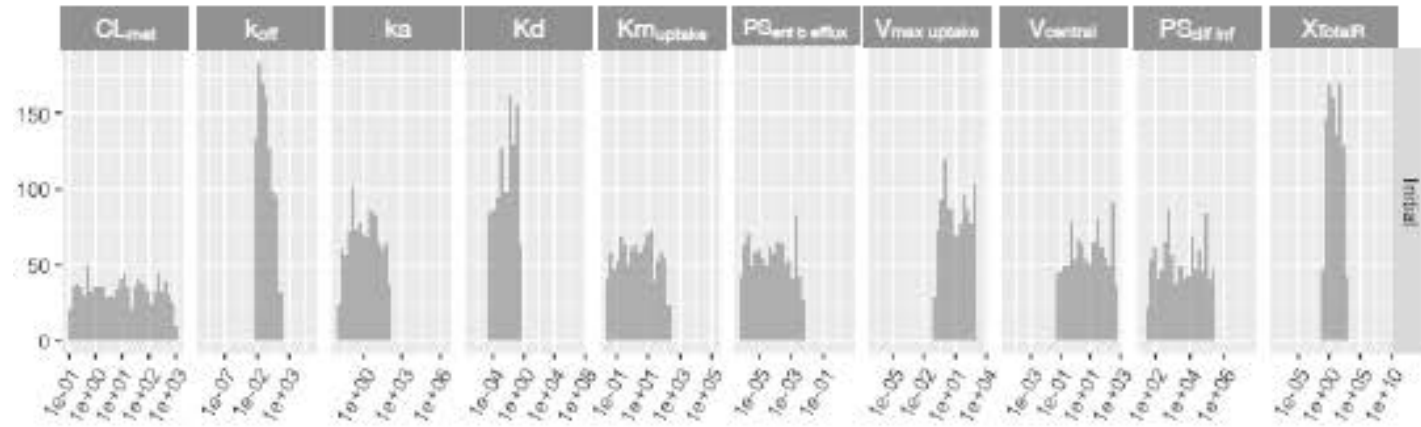
# Estimating the receptor occupancy of bosentan with top-down approach



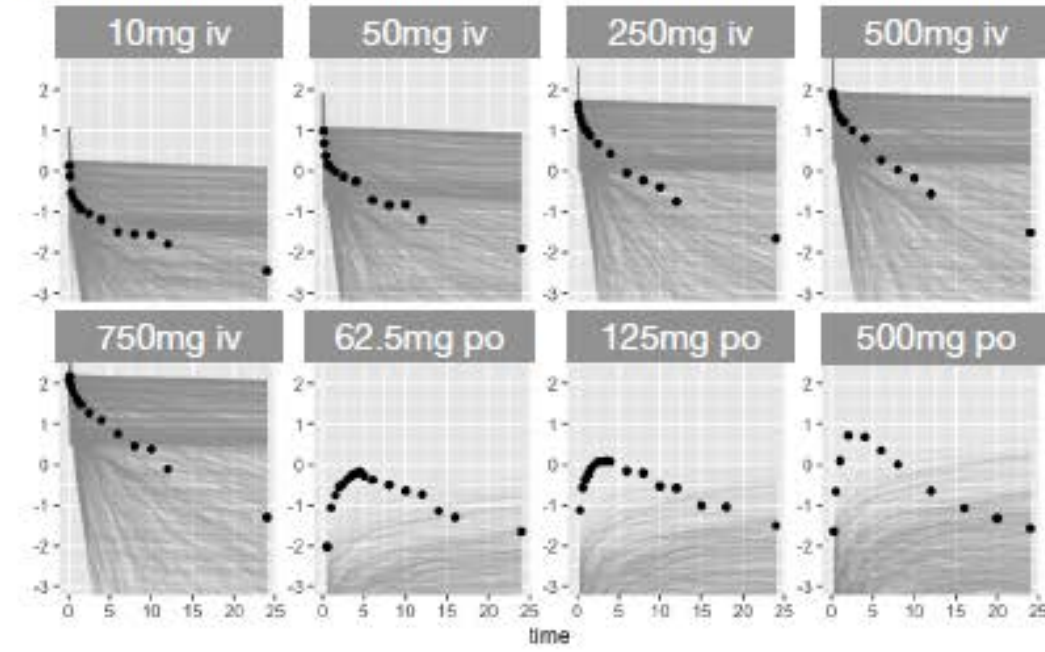
Koyama, Satoshi, et al. "Revisiting nonlinear bosentan pharmacokinetics by PBPK modeling: Target binding, albeit not a major contributor to nonlinearity, can offer prediction of target occupancy." *Drug Metabolism and Disposition* (accepted for publication).

# Estimating the receptor occupancy of bosentan with top-down approach

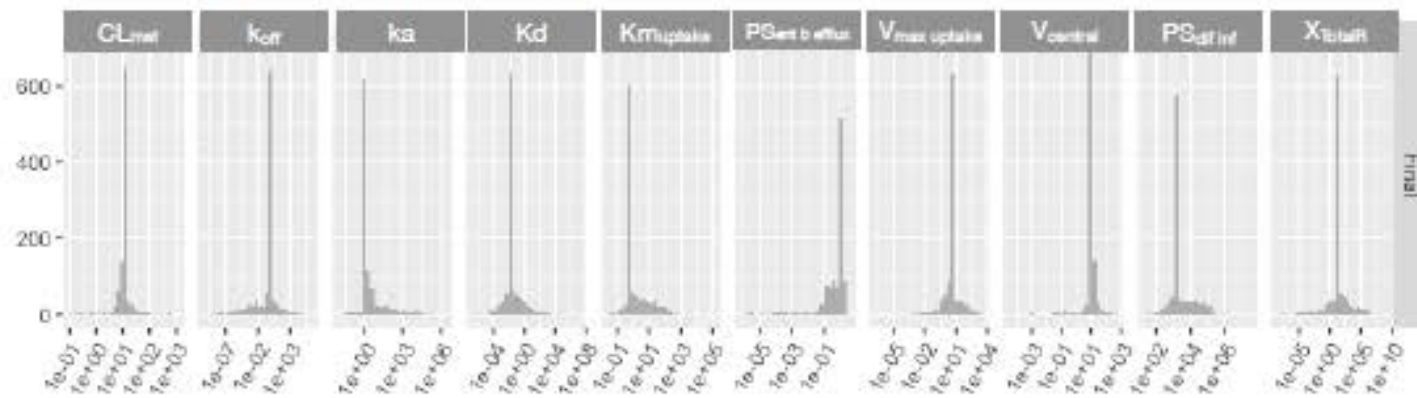
## Initial Parameter Distribution



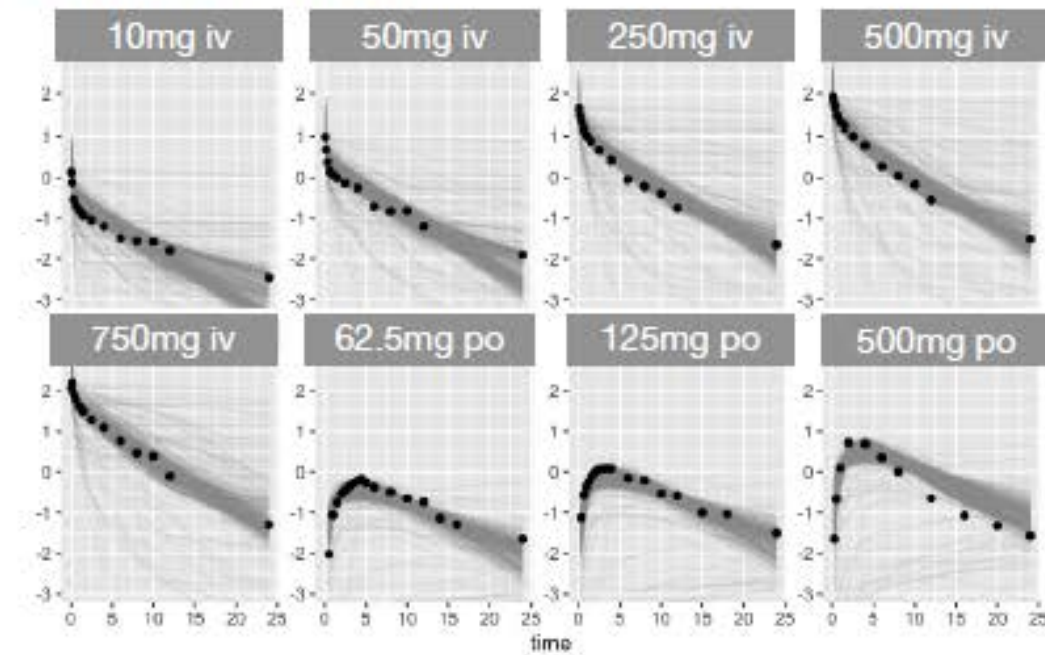
## Initial model fit



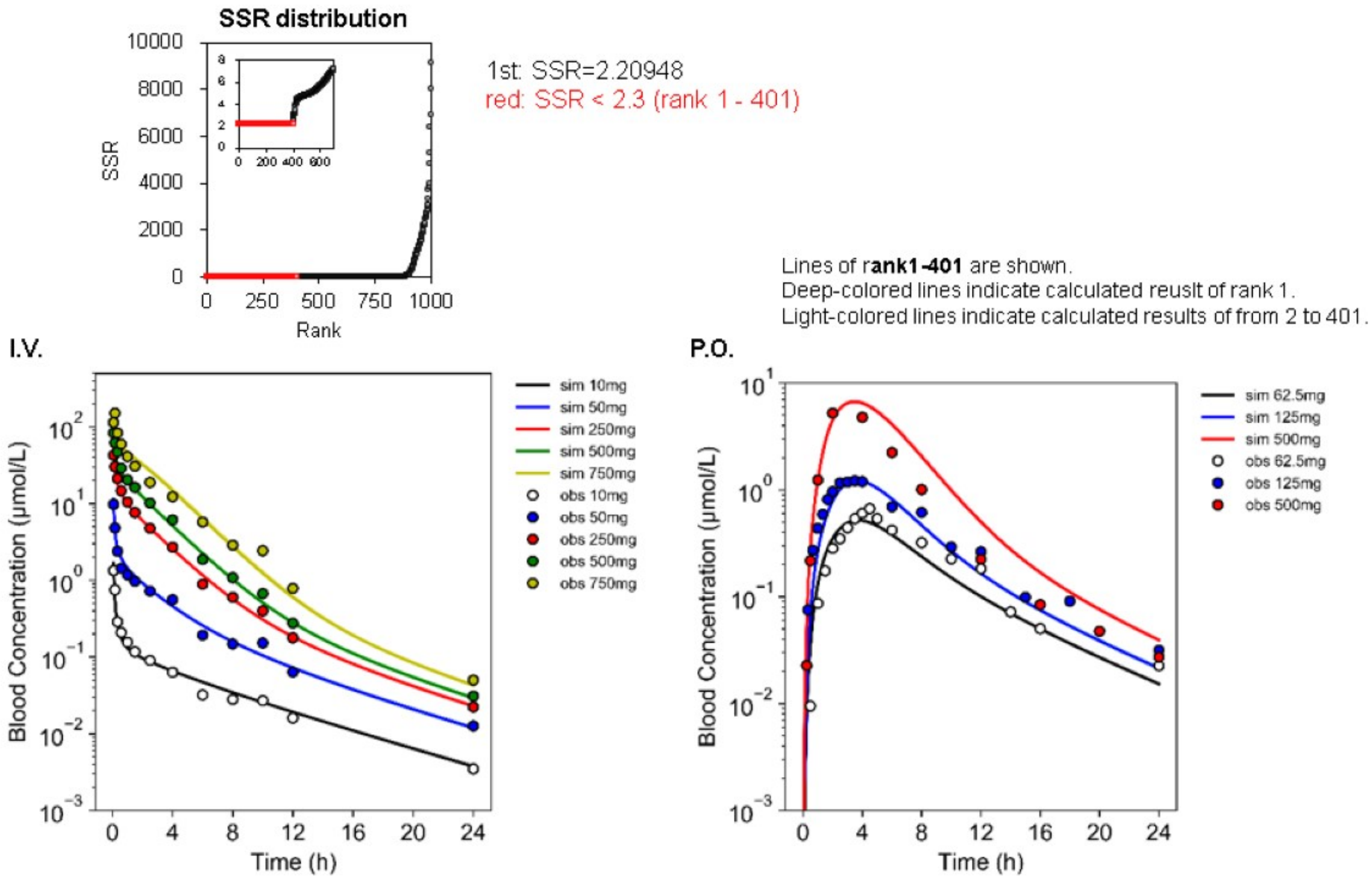
## Final Parameter Distribution



## Final model fit



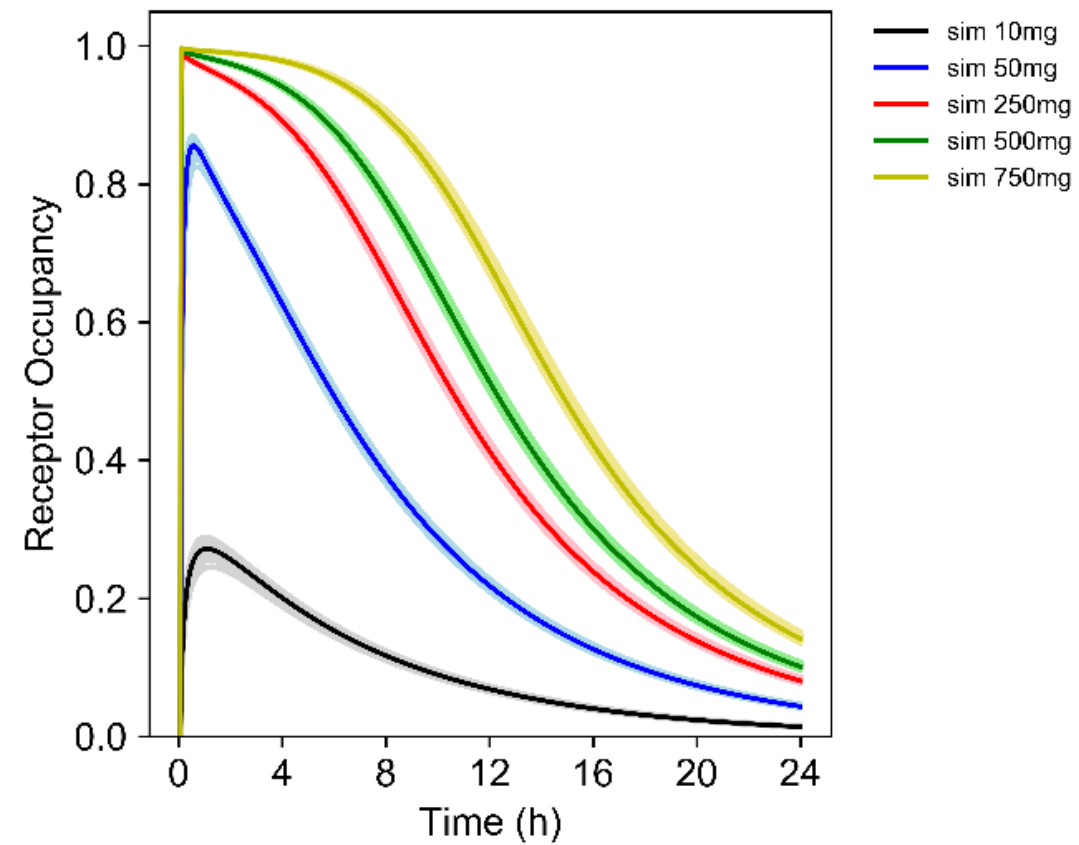
# Fitted Line and Observed Data of I.V. and P.O. of Bosentan



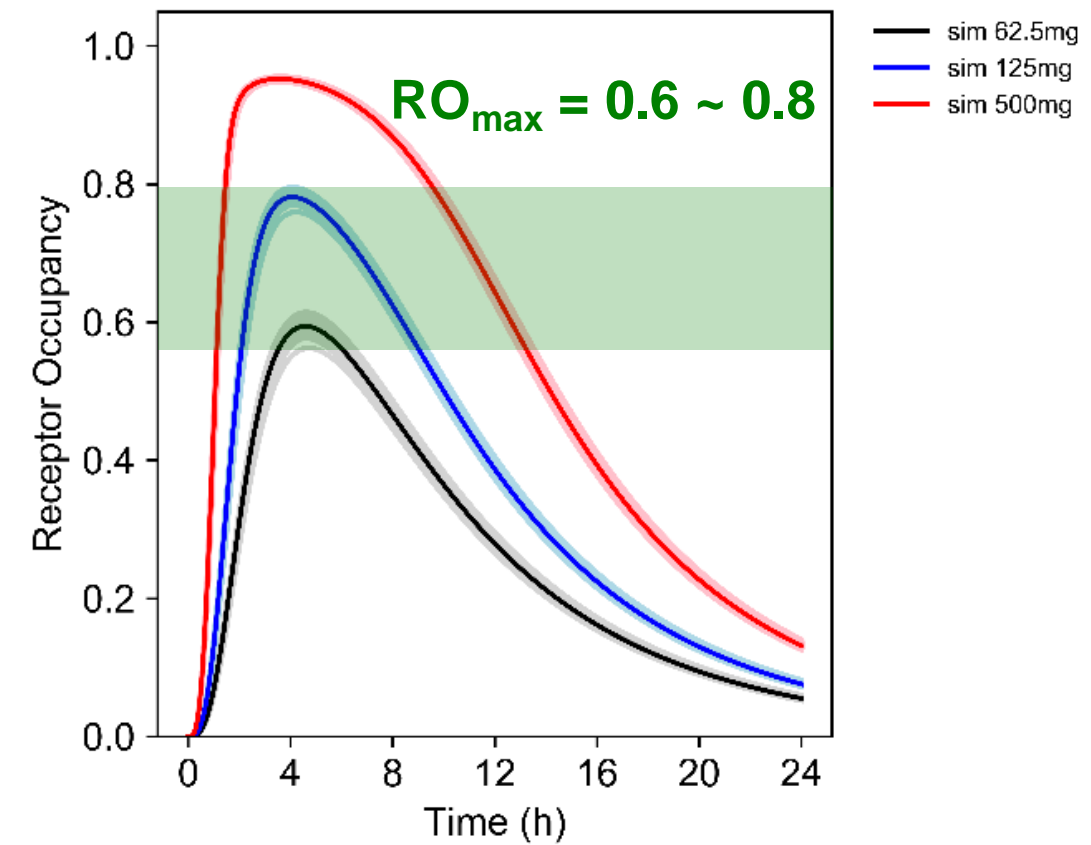
Now the data at all the doses are well captured by the model including target binding and saturable hepatic uptake.

# Calculated Time-Course of Receptor Occupancy

(A) I.V.



(B) P.O.

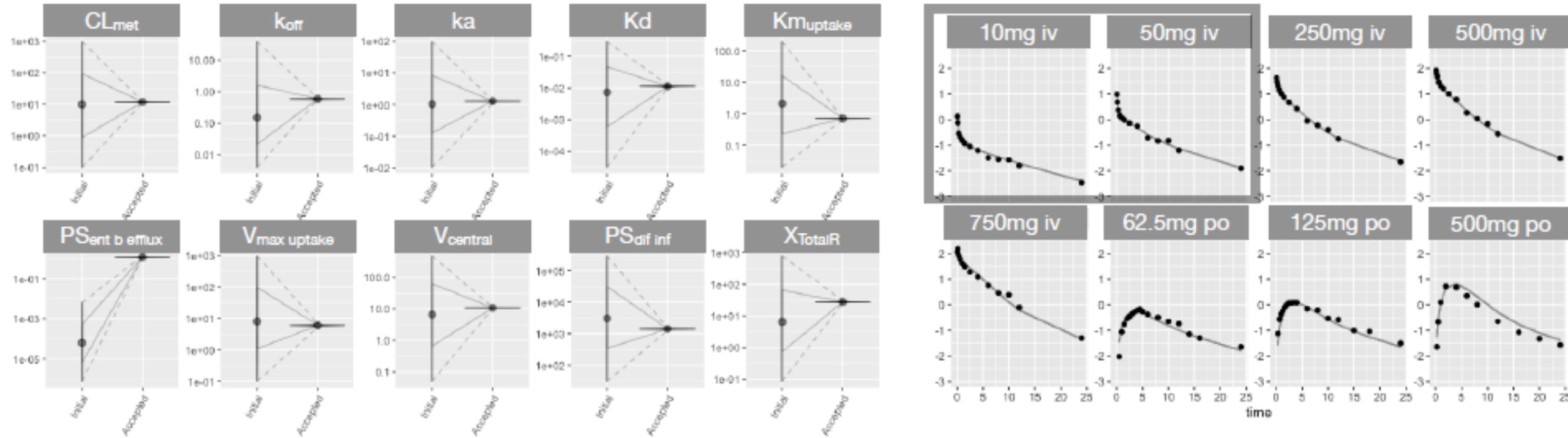


Lines of rank1-401 are shown.  
Deep-colored lines indicate calculated result of rank 1.  
Light-colored lines indicate calculated results of from 2 to 401.

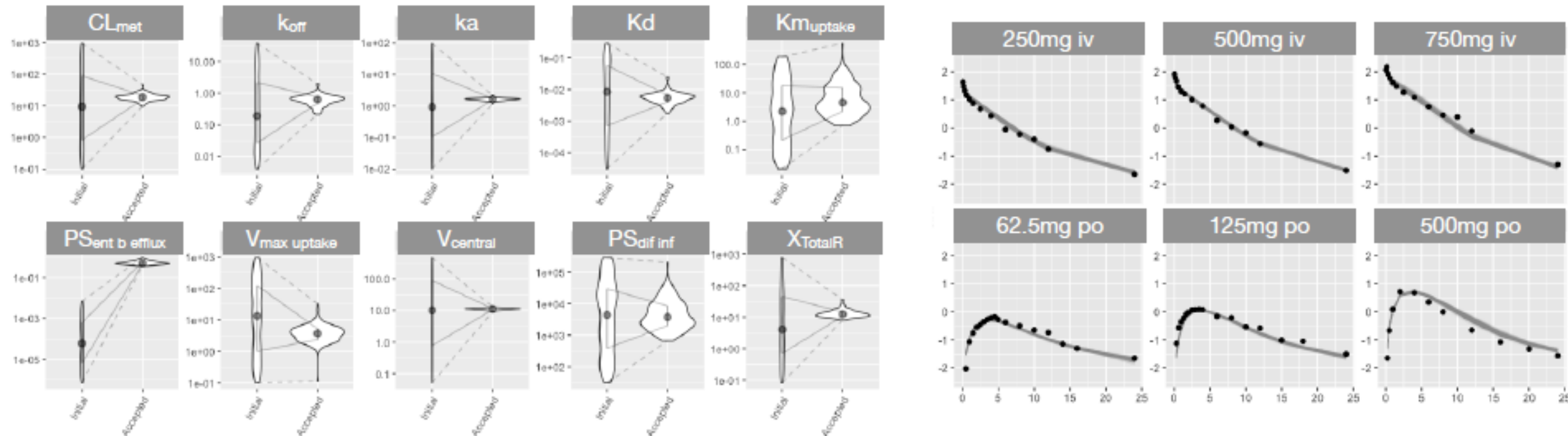
◆ Receptor occupancy was calculated 0.6-0.8 when clinical dose of Bosentan (62.5, 125 mg P.O.) was administered.

# Estimating the receptor occupancy of bosentan with top-down approach

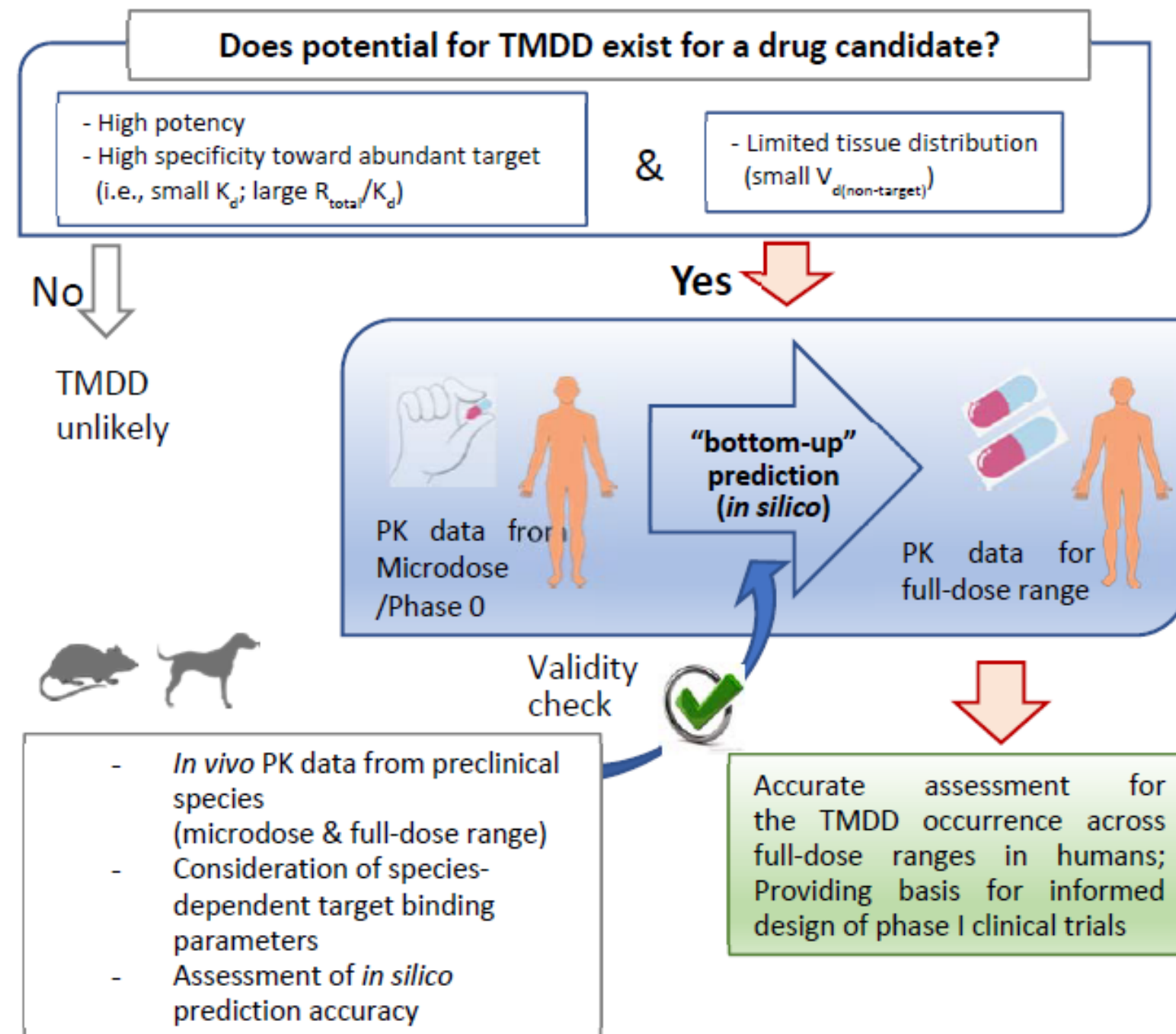
Parameters found though topdown approach **including low-dose**



Parameters found though topdown approach **not including low-dose**



**Workflow for assessment of drug candidates with possible TMDD.  
PBPK modeling-based prediction with target considerations can be combined  
with PK data analysis from a microdose analysis in human subjects.**



If a small dose (e.g., microdose) PK data is included in Phase 1 clinical dose escalation study, we may be able to detect the TMDD and predict the doses which exhibit the appropriate target occupancy (therapeutic dose)

**Burt T, Young G, Lee W, Kusuhara H, Langer O, Rowland M, Sugiyama Y.**

**Phase 0/microdosing approaches: time for mainstream application in drug development?**

**Nat Rev Drug Discov. 19::801-818 (2020).**

Our efforts for PBPK modeling to analyze TMDD will continue

To assess how the inclusion of microdosing (small dosing) data improves the prediction accuracy of overall target occupancy based on blood PK profiles

**We may be able to estimate the therapeutic dose during the phase 1 clinical studies once the starting dose is microdose or relevant small dose.**



# Disclosure for COI

I am a scientific advisory board member of SimCYP.

I have been serving as a chair and a vice-chair of the global consortia of Pharma Industries:

- (i) PET-IVIVE (6 companies) as a chair
- (ii) Endogenous biomarker-DDI prediction (8 companies)

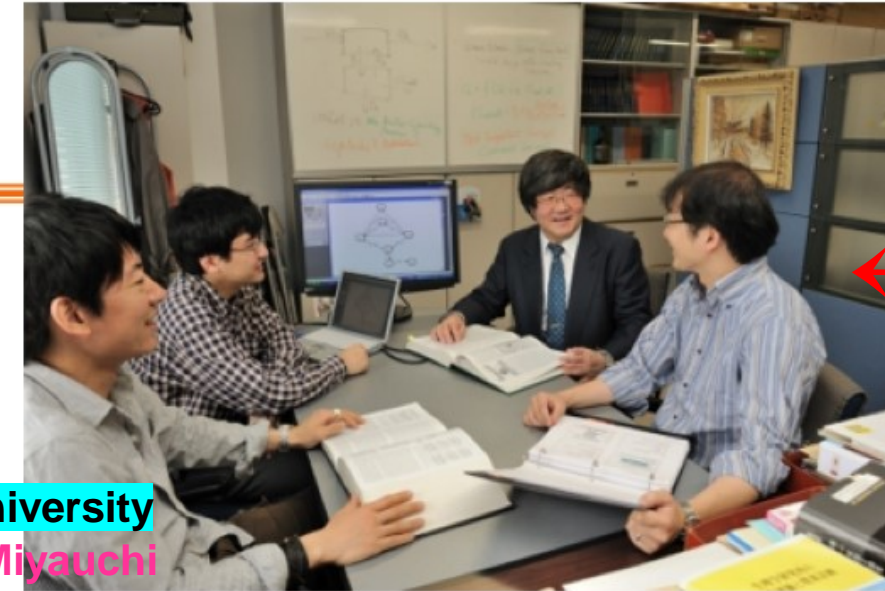
# Acknowledgement



Toho University  
Seiji Miyauchi

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- Takashi Yoshikado (wet +dry, IVIVE DDI)
- Kota Toshimoto** (dry, VCT (Virtual clinical study))
- Kim, Soo-Jin: (DDI, PGx, IVIVE)
- Atsuko Tomaru (IVIVE, Bioanalysis with LC/MS/MS)
- Satoshi Koyama (IVIVE)**
- Aya Kiriake (IVIVE, transporter expression systems)
- Kiyoe Morita (IVIVE, isolated hepatocyte (suspension, plated))



Kazuya Maeda, Hiroyuki Kusuhara  
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Woojin Lee

## Uppsala University

Yasunori Aoki

## Secretaries

- Sachie Sato
- Aya Sonobe
- Aya Miyamoto

← Hiroyuki Kusuhara

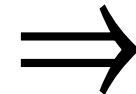
## Pfizer

Rodriguez, Varma, Kimoto

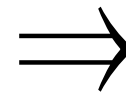
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(2021- ???)