

# Comparison of Genome Sequencing and Clinical Genotyping for Pharmacogenes

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Research

## Comparison of genome sequencing and clinical genotyping for pharmacogenes

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# Background

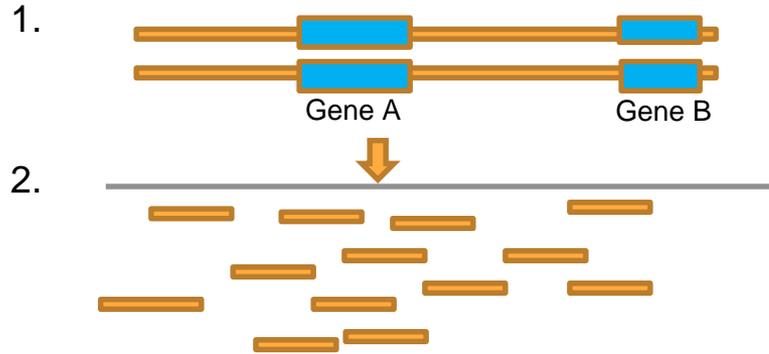
- St Jude Children's Research Hospital has been among the first to implement preemptive genomic testing to incorporate pharmacogenetics results in the medical record to assist in patient care.
- Recent St Jude protocol "*PGEN4Kids*" has implemented pharmacogenetics testing using pharmacogene-directed arrays such as the Affymetrix DMET plus array.
- Recently next generation sequencing (NGS) technology has experienced great advances, with lower cost and higher accuracies.
- Many NGS data have been generated at St Jude as part of research projects, such as Pediatric Cancer Genome Project (PCGP).

## Objective

1. To examine the interrogation from genome sequencing technology for actionable pharmacogenes.
2. To compare the concordance between genotypes generated by genome sequencing and our clinical array-based genotyping results.

# Basic Introduction of Next Generation Sequencing (NGS):

## Whole Genome Sequencing (WGS)



Generate sequence reads (e.g. 100bps)

3. Reads

Align with reference genome and generate genotyping calls

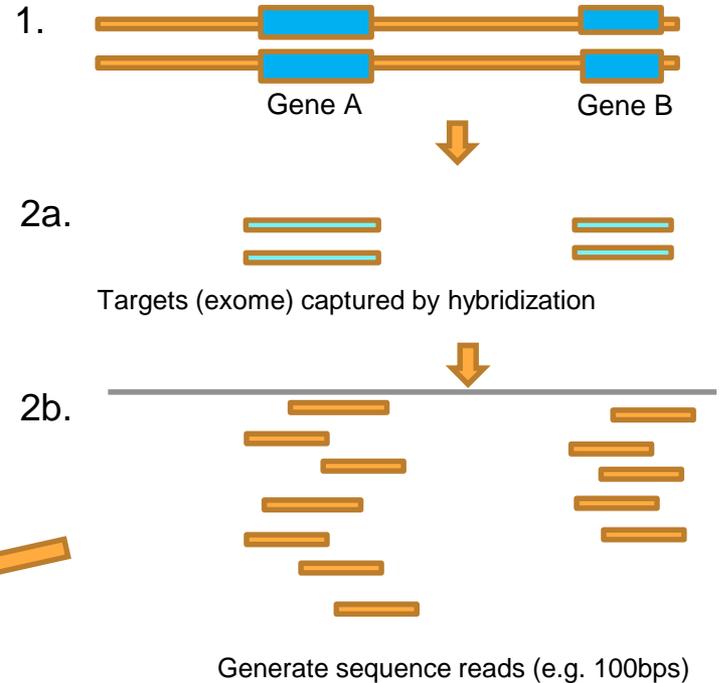
Reference Genome

```
ATGGTATTGTAATTGACAT
TGGTATTGTAATTG
AGATGGTATTGTAATTGA
GATGGTATTGCAATTGACAT
GCAATTGACAT
ATGGTATTGCAATTG
AGATGGTATTGCAATTGACAT
AGATGGTATTGTAATTGACAT
```

Ref/Alt -> C/T

Ref/Ref -> G/G

## Whole Exome Sequence (WES)



# Basic Quality Controls metrics in NGS:

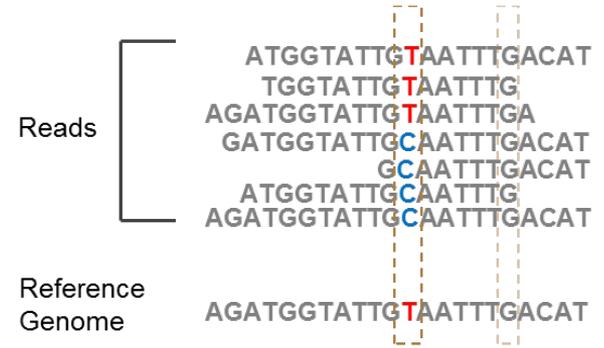
## Coverage (read depth)

- Average WGS, 30X
- Average WES, 60X
- Read depth < 10X, considered “NoCall”

## minor allele fractions (MAFrac)

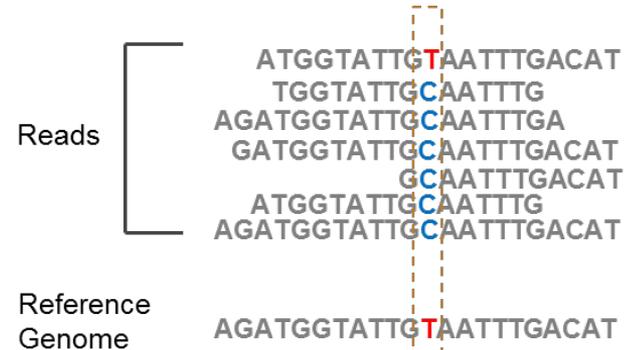
- Heterzyous genotypes should be close to 0.5 (50%)
- Low allele fraction is questionable, suggesting contamination, sequencing error, etc.

**Other QC:** strand bias, Base quality, etc



Read depth: 7

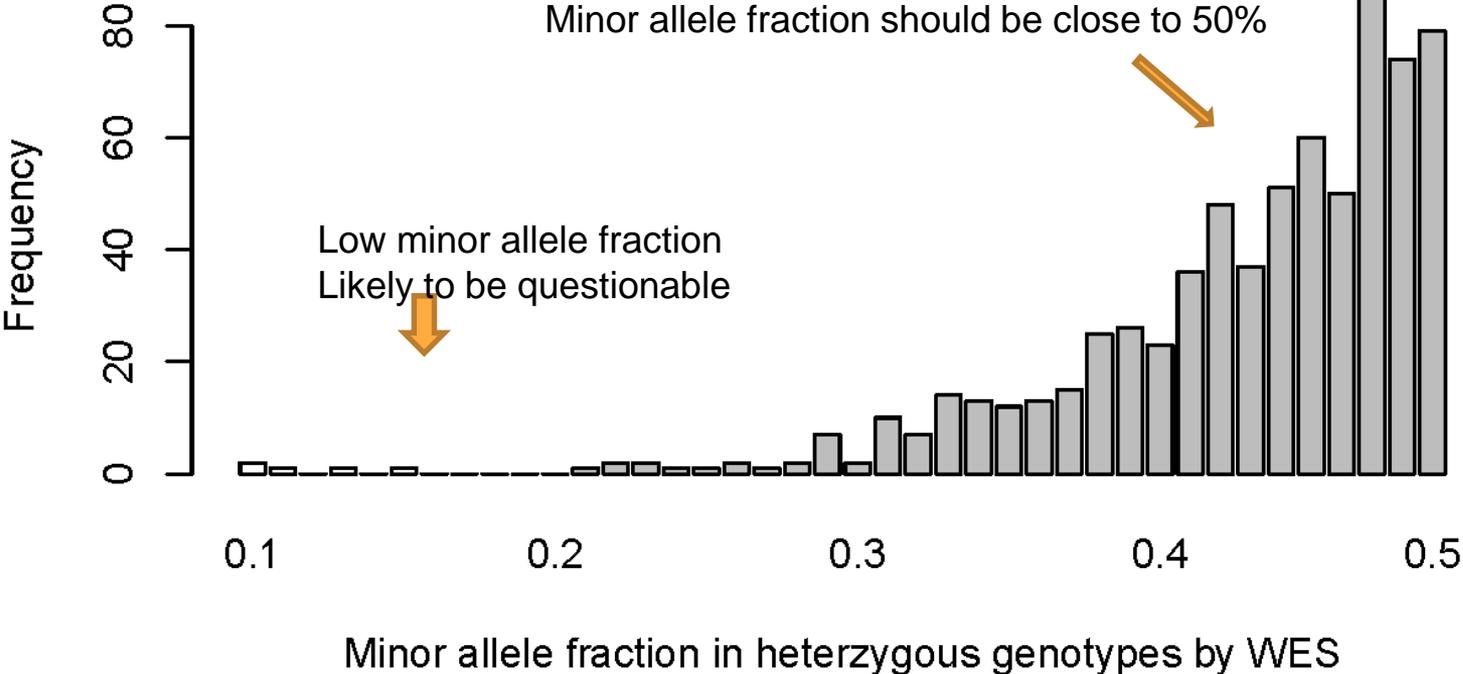
MAFrac:  $3/7 = 42.9\%$



Read depth: 7

MAFrac:  $1/7 = 10.2\%$

# Distribution of minor allele fraction of heterozygous calls in NGS



# Patient Data

## Clinical Genotyping (Affymetrix DMET Plus Array v1)

- N = 2656 (1319 whites, 998 blacks, 232 Hispanics)

## Whole Genome Sequencing (WGS)

- N = 68 (44 whites, 18 blacks)
- all 68 patients have both DMET array and WGS

## Whole Exome Sequencing (WES)

- N=636 (396 whites, 95 blacks, 86 Hispanics)
- 176 patients have both DMET array and WES



# CPIC Important Genes and Variants

## CPIC Important Genes: (n=13)

*CFTR, CYP2C19, CYP2C9, , CYP2D6, CYP3A4, DPYD, G6PD,*

*HLA-B, IFNL3, SLCO1B1, TPMT, UGT1A1, VKORC1*

(collected from <https://www.pharmgkb.org/view/dosing-guidelines.do?source=CPIC> as of 07/01/2015)



The screenshot shows the PharmGKB website interface. At the top, there is a navigation bar with the PharmGKB logo and the tagline "Pharmacogenetics. Knowledge. Implementation." The main content area is titled "Dosing Guidelines - CPIC". Below the title, there is a filter dropdown menu set to "CPIC". The main content is a table with three columns: "Drug", "Guidelines", and "Updated". The table lists 10 drugs with their corresponding CPIC guideline titles and update dates.

Drug	Guidelines	Updated
abacavir	CPIC Guideline for abacavir and HLA-B	09/30/2014
allopurinol	CPIC Guideline for allopurinol and HLA-B	06/12/2015
amitriptyline	CPIC Guideline for amitriptyline and CYP2C19,CYP2D6	02/07/2014
atazanavir	CPIC Guideline for atazanavir and UGT1A1	09/18/2015
azathioprine	CPIC Guideline for azathioprine and TPMT	05/10/2016
capecitabine	CPIC Guideline for capecitabine and DPYD	08/06/2014
carbamazepine	CPIC Guideline for carbamazepine and HLA-B	02/07/2014
citalopram	CPIC Guideline for citalopram,escitalopram and CYP2C19	05/11/2015

# CPIC important variants

Based on gene activities associated with variants from supplemental table of published CPIC guidelines

- Variants associated with **increase/decreased/no-function** were considered important.
- Exclude variants with **unknown** function and **normal** functions.

**SUPPLEMENTAL TABLE S2. ASSOCIATION BETWEEN ALLELIC VARIANTS<sup>A</sup> AND CYP2D6 ENZYME ACTIVITY**

Functional Status (2, 7)	Activity Value <sup>c,d</sup>	Alleles
Increased function	>1	*1xN, *2xN, *35xN, *45 <sup>e</sup> xN
Normal or Increased function	1 or >1 <sup>h</sup>	*9xN, *10xN, *17xN, *29xN, *41xN
Normal function <sup>b</sup>	1	*1 <sup>e</sup> , *2, *27, *33, *34 <sup>f</sup> , *35, *39 <sup>f</sup> , *45 <sup>g</sup> , *46 <sup>g</sup> , *48, *53
Decreased function	0.5	*9, *10, *14B, *17, *29, *41, *49, *50, *54, *55, *59, *72
No-function	0	*3, *3xN, *4, *4xN, *5, *6, *6xN, *7, *8, *11, *12, *13, *14A, *15, *18, *19, *20, *21, *31, *36, *36xN, *38, *40, *42, *44, *47, *51, *56, *57, *62, *68, *69, *92, *100, *101
Unknown	N/A	*22, *23, *24, *25, *26, *28, *30, *32, *37, *43, *43xN, *52, *58, *60, *61, *63, *64, *65, *70, *71, *73, *74, *75, *81, *82, *83, *84, *85, *86, *87, *88, *89, *90, *91, *93, *94, *95, *96, *97, *98, *102, *103, *104, *105

\* CPIC Guideline for codeine and CYP2D6

## CPIC Important Variants: (n=127)

- 103 Single Nucleotide Variation (SNV) (95 exonic)
- 21 Indels/repeats (20 exonic)
- two structural variants (*CYP2D6*), Copy Number Variation (CNV) and *CYP2D6/2D7* hybrid
- one haplotype (*HLA-B*)

Gene	Number of CPIC important variants			
	SNV (exonic)	Indel (exonic)	Other	Total
<i>CFTR</i>	10 (10)	2 (2)		12
<i>CYP2C19</i>	8 (7)	0		8
<i>CYP2C9</i>	10 (10)	2 (2)		12
<i>CYP2D6</i>	26 (24)	13 (13)	2 structural variations	41
<i>CYP3A5</i>	2 (1)	1 (1)		3
<i>DPYD</i>	10 (10)	2 (2)		12
<i>G6PD</i>	7 (7)	0		7
<i>HLA-B</i>	0	0	1 haplotype	1
<i>IFNL3</i>	2 (0)	0		2
<i>SLCO1B1</i>	12 (11)	0		12
<i>TPMT</i>	15 (15)	0		15
<i>UGT1A1</i>	0	1 (0)		1
<i>VKORC1</i>	1 (0)	0		1
<b>Total</b>	103 (95)	21 (20)	3	127

## Analysis pipelines used to generate genotypes

Affymetrix DMET Plus Array v1 (231 genes, 1936 variants)

- DMET Console software from Affymetrix

## Whole Exome and Whole Genome Sequencing

- GATK v3.4 for SNVs and Indels, following best practice guideline, with recommended parameters and quality control steps.
- XHMM and CONCERTING for CNV estimation.
- Polysolver and OptiType for inferring *HLA-B* alleles.

# 1.CFTR

**DMET:** not interrogated

**WES:** good coverage

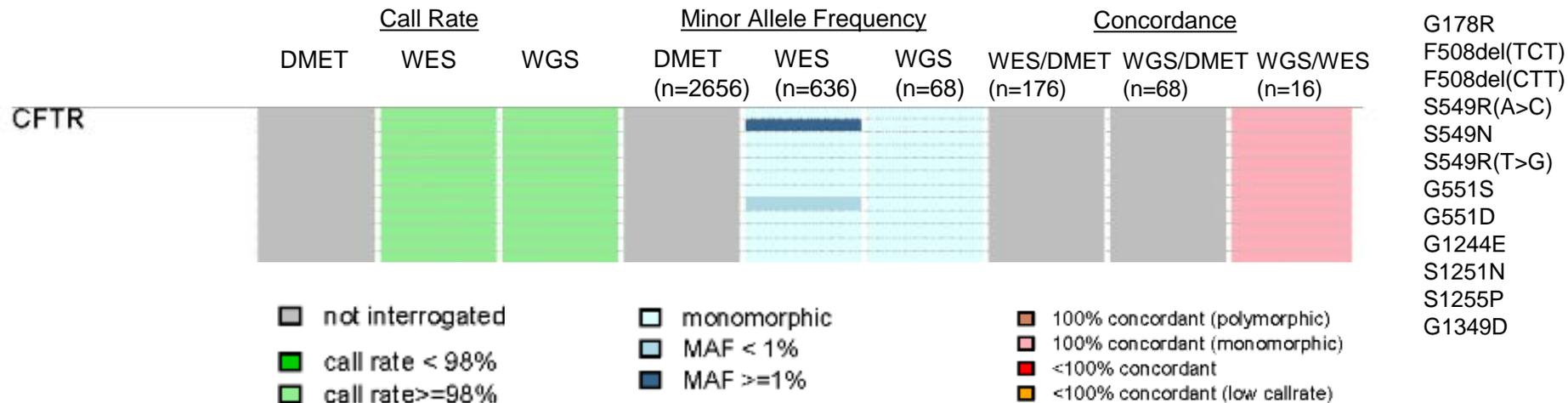
**WGS:** good coverage

**Drug:** ivacaftor

CPIC important Variants (n=12):

- 10 exonic SNV
- 2 exonic indels

No discordant genotypes between WGS and WES



## 2.CYP2C19

**DMET:** good coverage

**WES:** missing important intronic variant

*CYP2C19*\*17, associated with increase activity

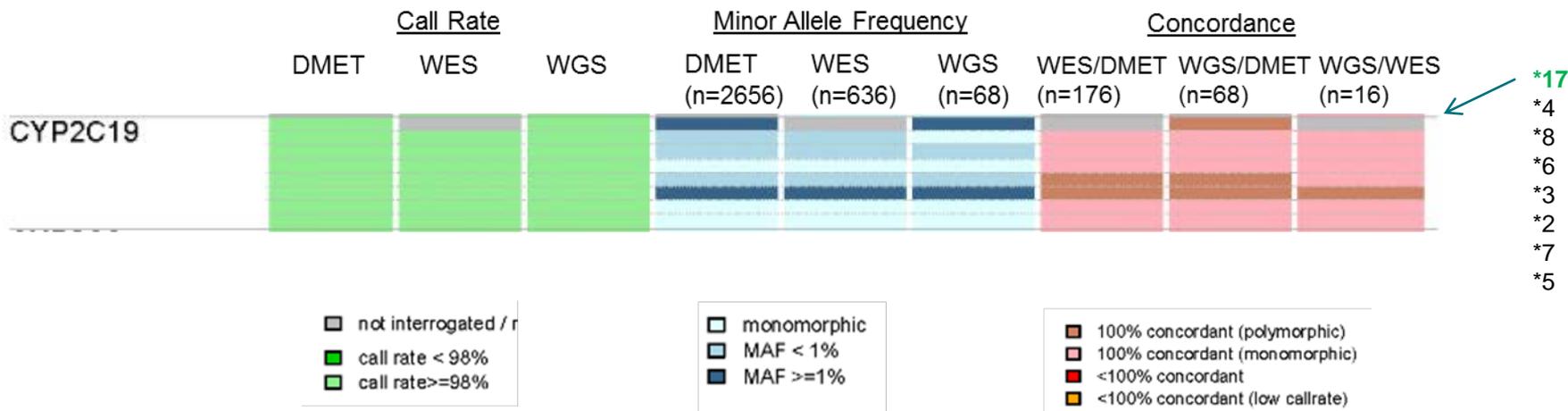
**WGS:** good coverage

No discordant genotypes were observed

**Drug:** Clopidogrel, Amitriptyline, citalopram, clomipramine, doxepin, imipramine, setraline, trimipramine

CPIC Important Variants (n=8):

- 8 SNV (7 exonic)



### 3. CYP2C9

**DMET:** low call rate on R150H (\*8) and not interrogated very rare variant I327T (\*31); both “possible decreased activity”

**WES:** good coverage

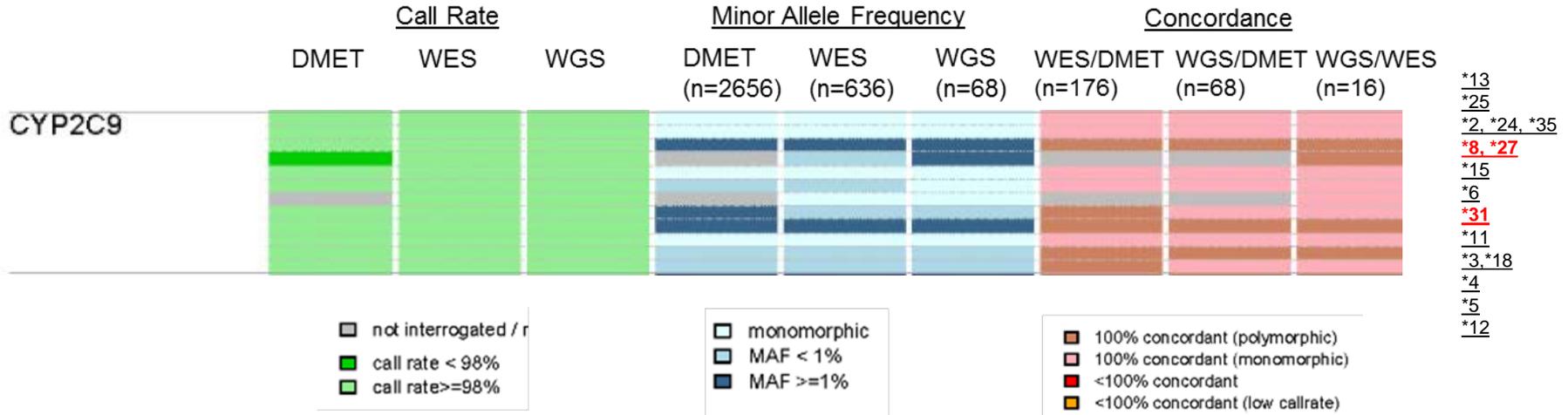
**WGS:** good coverage

No discordant genotypes were observed.

**Drug:** Warfarin, Phenytoin

Important Variants (n=12):

- 10 exonic SNVs
- 2 exonic Indels



## 4. *CYP2D6*

CPIC Important Variants (n=41):

- 26 SNV (24 exonic)
- 13 exonic Indels
- 2 structural variations (CNV, *CYP2D6/2D7* hybrid)

### **Clinical Genotyping:**

- Affymetrix **DMET** interrogated 23 SNV/Indels.
- CNV and *CYP2D6/2D7* were interrogated by add-on qPCR assay.

### **WES:**

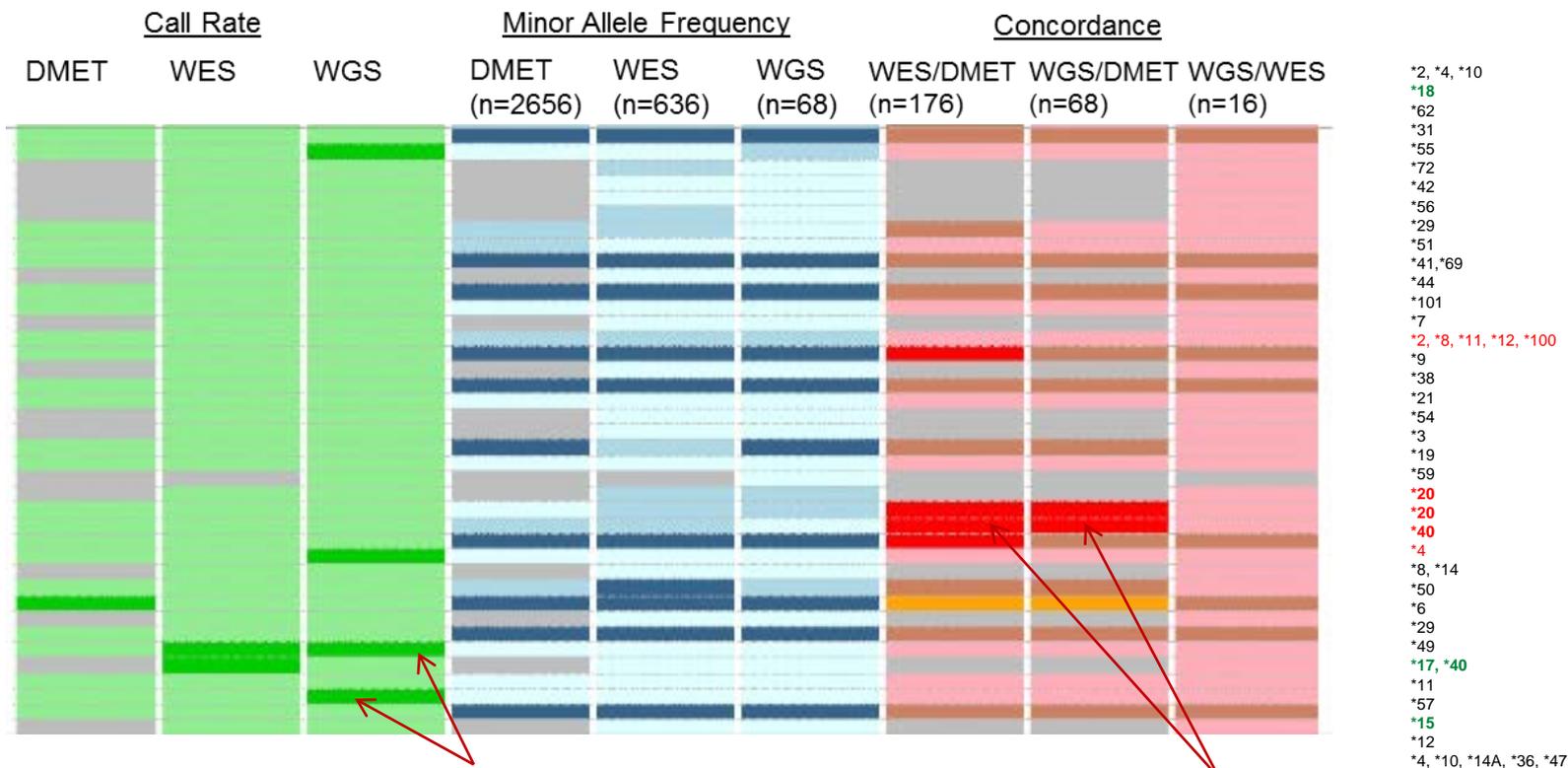
- interrogated 36 SNV/indels.
- CNV can be inferred, *CYP2D6/2D7* not interrogated.

### **WGS:**

- Interrogated 35 SNV/Indels.
- CNV can be inferred, *CYP2D6/2D7* not interrogated.

**Drug:** amitriptyline, clomipramine, codeine, desipramine, doxepin, fluvoxamine, imipramine, nortriptyline, paroxetine, trimipramine.

# CYP2D6: SNVs and Indels (n=39)



lower call rate from WGS likely due to CNV

discordant calls were observed in 7 WES and 3 WGS genotypes.

## CYP2D6 discordant genotyping calls between DMET and WES

Gene	Allele	dbSNP	DMET Call	WES Call	Reference Allele Count	Alternative Allele Count	Minor Allele Fraction	Comment
CYP2D6	*20 (1973insG)	rs72549354	T/T	T/TC	364	57	<b>13.5%</b>	WES low minor allele fraction
CYP2D6	*20 (1973insG)	rs72549354	T/T	T/TC	278	51	<b>15.5%</b>	WES low minor allele fraction
CYP2D6	*4 (1846G>A)	rs3892097	T/T	C/T	10	83	<b>10.8%</b>	WES low minor allele fraction
CYP2D6	*4 (1846G>A)	rs3892097	T/T	C/T	15	133	<b>10.1%</b>	WES low minor allele fraction
CYP2D6	*2 (R296C)	rs16947	A/A	A/G	71	9	<b>11.3%</b>	WES low minor allele fraction
CYP2D6	*40 (1863_1864ins)	rs72549356	-/18bps	-/-	138	0	0.0%	Reason for discrepancy unclear
CYP2D6	*40 (1863_1864ins)	rs72549356	-/18bps	-/-	292	0	0.0%	Reason for discrepancy unclear

5 out of 7 discordant calls have low WES MAFraction, suggesting WES results may be suspect.

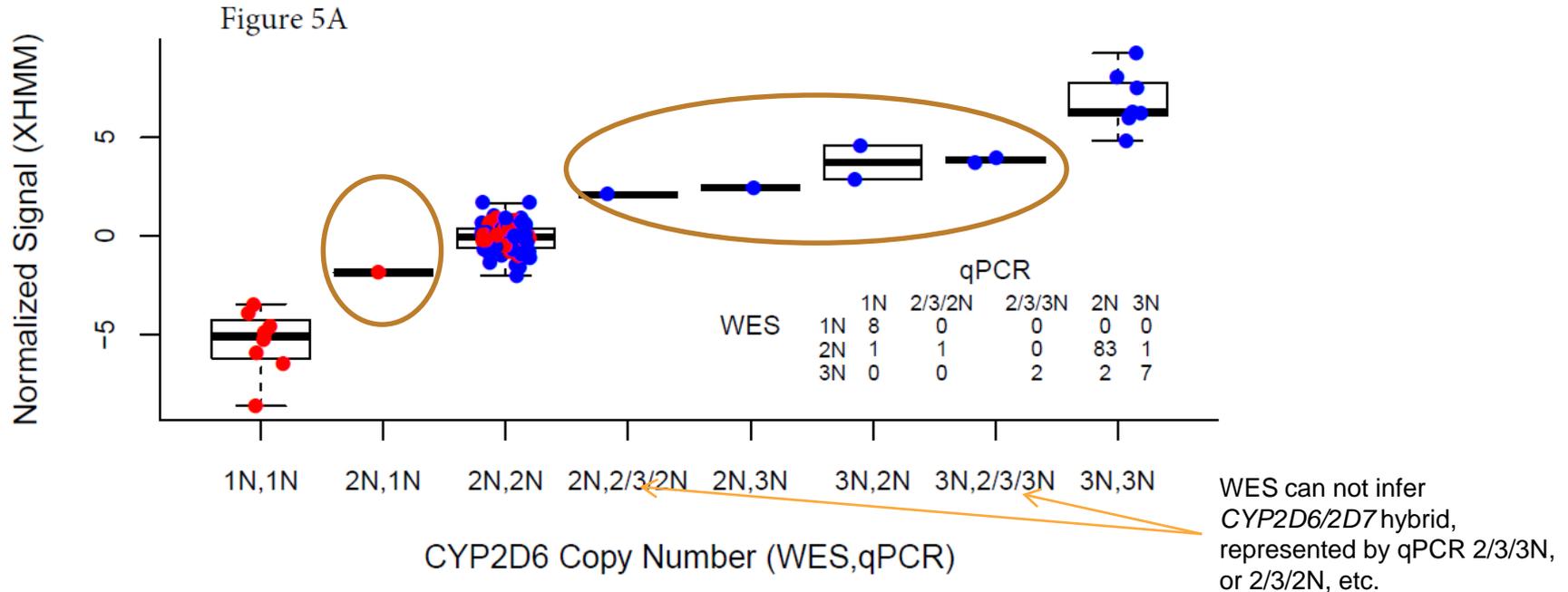
## CYP2D6 discordant genotyping calls between DMET and WGS

Gene	Allele	dbSNP	DMET Call	WGS Call	Reference Allele Count	Alternative Allele Count	Minor Allele Fraction	Comments
<i>CYP2D6</i>	*20 (1973insG)	rs72549354	T/T	T/TC	42	6	12.5%	WGS low MAFraction
<i>CYP2D6</i>	*40 (1863_1864ins)	rs72549356	-/18bps	-/-	30	0	0.0%	Reason for discrepancy unclear
<i>CYP2D6</i>	*40 (1863_1864ins)	rs72549356	-/18bps	-/-	32	0	0.0%	Reason for discrepancy unclear

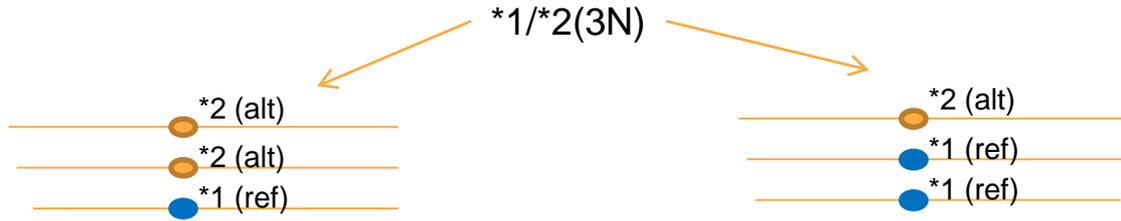
# CYP2D6 Copy number can be inferred by WES

DMET CNV was inferred by qPCR add-on assay  
WES CNV was inferred byXHMM

Concordance: 98/105 (93.3%), 3 of 7 discordant calls are possibly *CYP2D6/2D7* hybrid



# Haplotype composition for *CYP2D6* (3N) can be inferred by WES



Alt/Ref read depth ratio = 2  
 $\text{Log}_2(\text{Alt/Ref read depth ratio}) = 1.0$   
 Inferred haplotypes: \*1/\*2/\*2

Alt/Ref read depth ratio = 0.5  
 $\text{Log}_2(\text{Alt/Ref read depth ratio}) = -1.0$   
 Inferred haplotypes: \*1/\*1/\*2

patient	chr22:4252669 4 (P34S, *4)	chr22:42524947 (1846G>A, *4)	chr22:42522613 (S486T, *2, *4)	chr22:42523943 (R296C, *2)	chr22:42524178 (2615delAAG, *9)	WES CNV	qPCR CNV	Haplotype composition	Comment
1	Hom_Ref	Hom_Ref	1.093	0.813	Hom_Ref	3N	3N	*1/*2/*2	
2	Hom_Ref	Hom_Ref	1.052	1.222	Hom_Ref	3N	3N	*1/*2/*2	
3	Hom_Ref	Hom_Ref	-1.141	-0.955	Hom_Ref	3N	3N	*1/*1/*2	
4	-0.781	-1.188	Hom_Alt	0.595	Hom_Ref	3N	3N	*2/*2/*4	
5	-0.933	-1.322	-1.000	Ref	Hom_Ref	3N	3N	*1/*1/*4	

## 5. CYP3A5

**DMET:** Good Coverage

**WES:** missing important intronic variant CYP3A5\*3

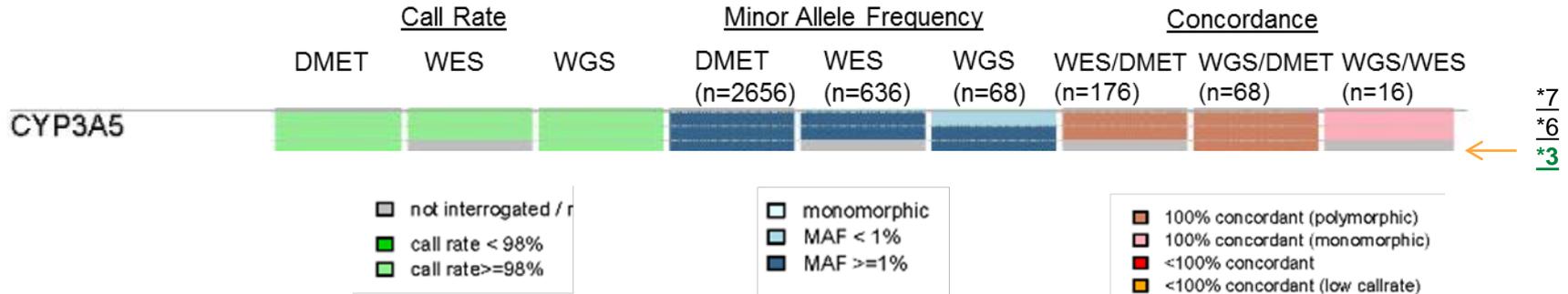
**WGS:** Good coverage

**Drug:** tacrolimus

Important Variants (n=3):

- 2 SNV (1 exonic)
- 1 exonic Indel

No discordant calls were observed



## 6. DPYD

**DMET:** not interrogating rs67376798 (Important) and two rare variants \*12

**WES:** good coverage

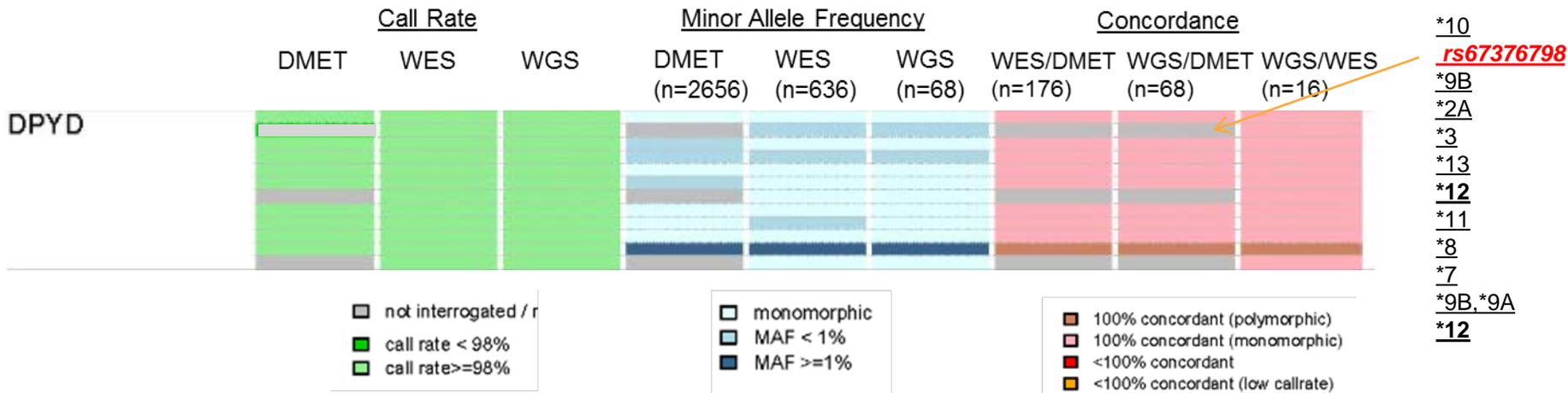
**WGS:** good coverage

No discordant genotypes were observed.

**Drug:** capecitabine, fluorouracil, tegafur

Important Variants (n=12):

- 10 exonic SNV
- 2 exonic Indel



# 7. G6PD

**DMET:** missing important variants, e.g. common variants Asahi; and other rare variants

**WES:** Good coverage

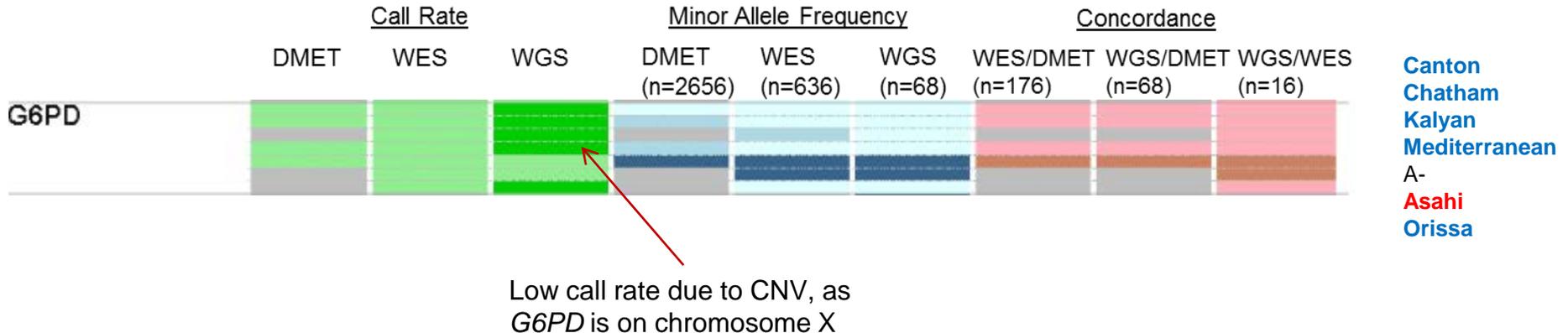
**WGS:** lower call rate in many positions due to lower coverage

No discordant calls were observed

**Drug:** rasburicase

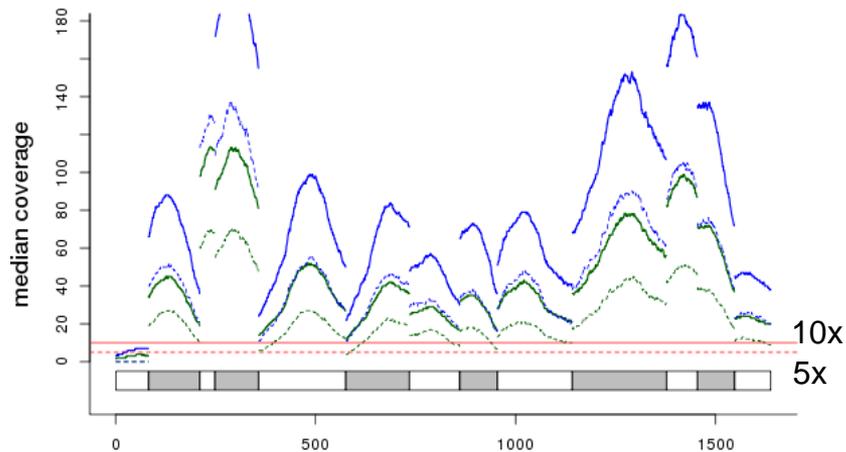
Important Variants (n=7):

- 7 exonic variants (PharmGKB 2015)

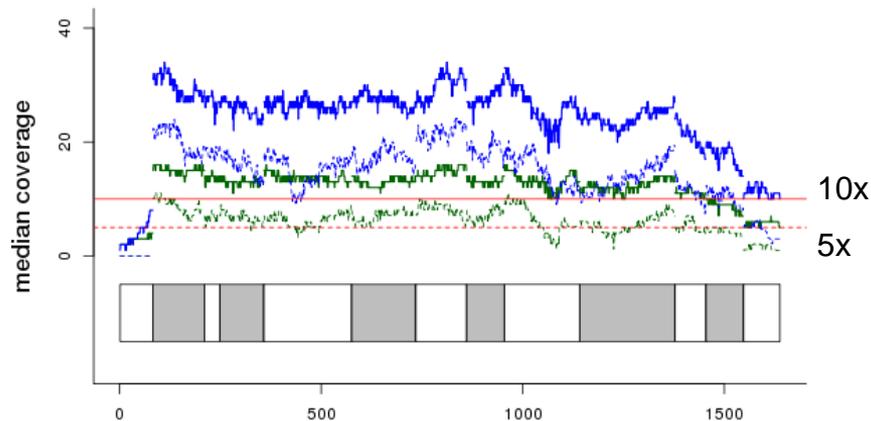


# Coverage of *G6PD* by Gender

WES (n=636)



WGS (n=68)

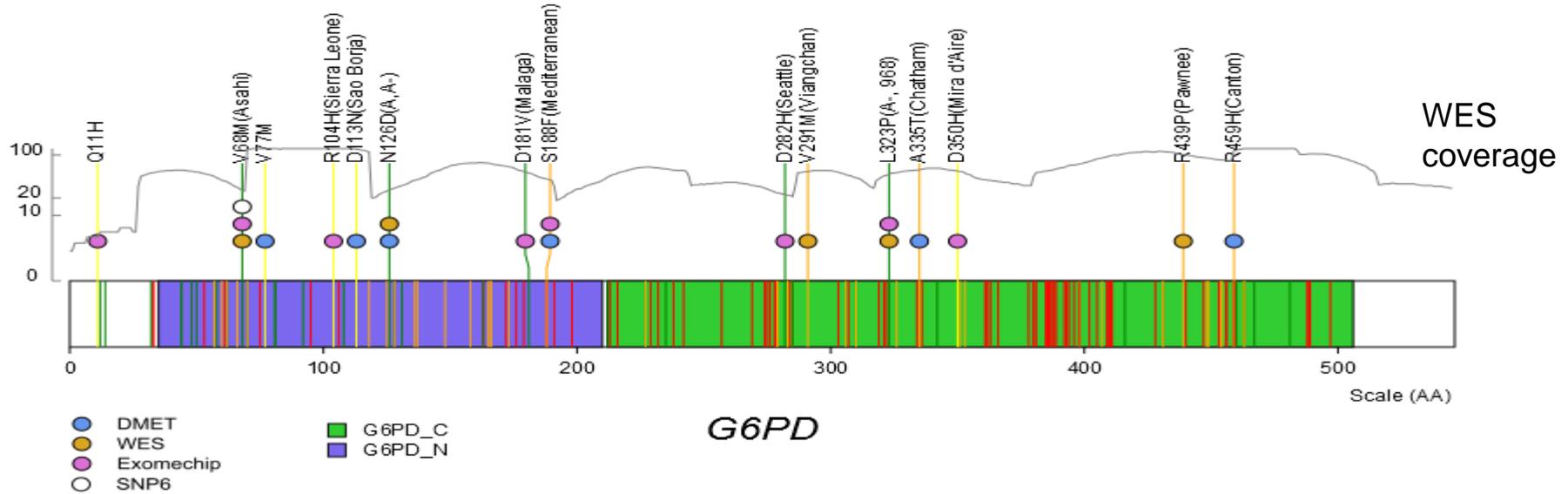


Blue: Females; Green: Males

Solid line: median coverage

Dashed line: 5% patients have coverage below the dashed line

# G6PD SNPs in Public Database and interrogated in SNPCHIPS



- WHO class I
- WHO class II
- WHO class III
- WHO class IV or unknown

Over 100 important rare variants (WHO class I/II).  
DMET Plus v1 only interrogates six variants.

## 8. IFNL3

**DMET:** not on DMET array;

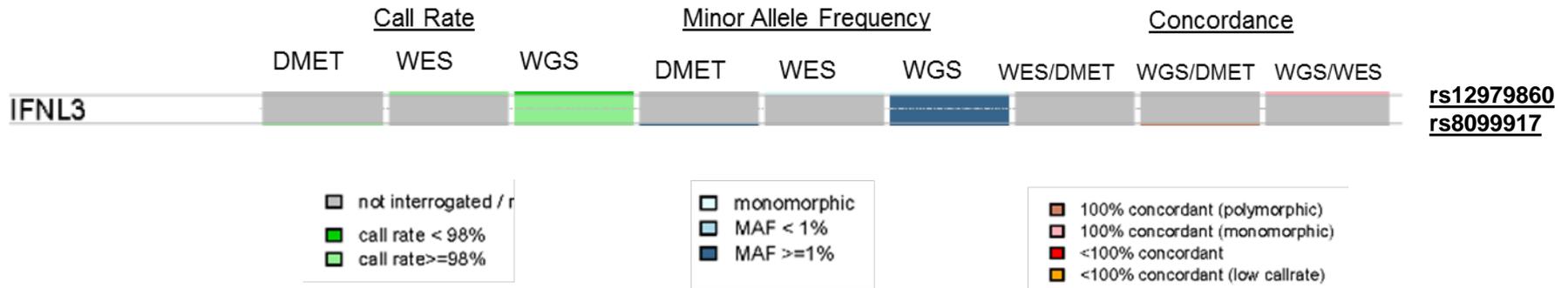
**WES:** upstream variants not targeted

**WGS:** Good coverage

**Drug:** peginterferon alfa-2, ribavirin

Important Variants (n=2):

- 2 variants upstream of the gene



## 9. *SLCO1B1*

**DMET:** missing rare variant \*23; low call rate at \*35

**WES:** missing promoter SNP *SLCO1B1*\*17

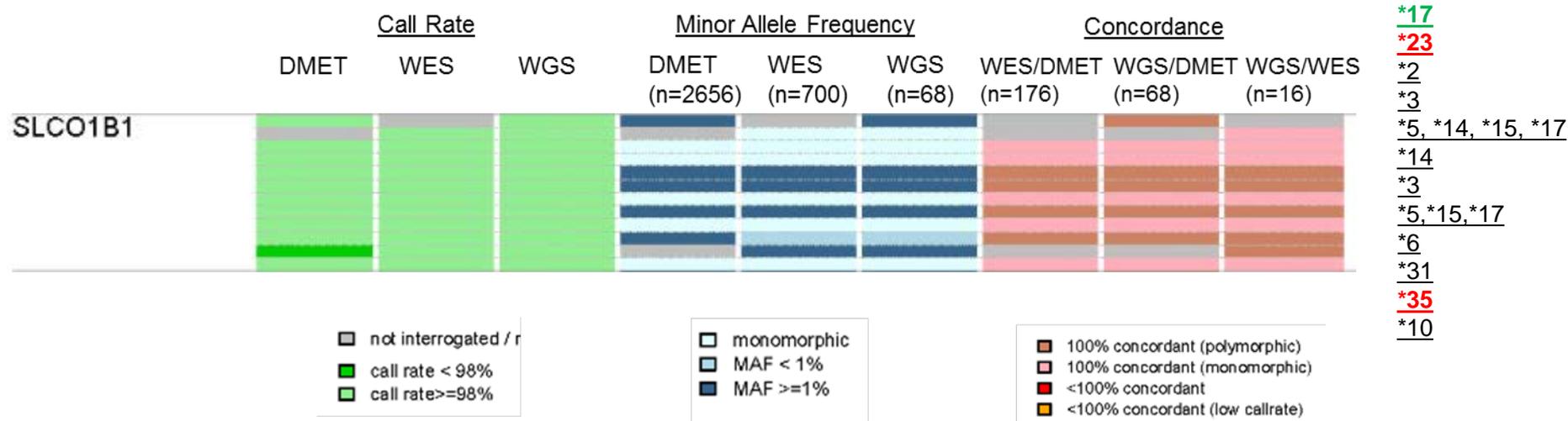
**WGS:** good coverage

**Drug:** simvastatin

Important Variants (n=12):

- 12 SNVs (11 exonic)

No discordant genotypes were observed.



# 10. TPMT

**DMET:** interrogates most common variants;  
rare variants not interrogated

**WES:** good coverage

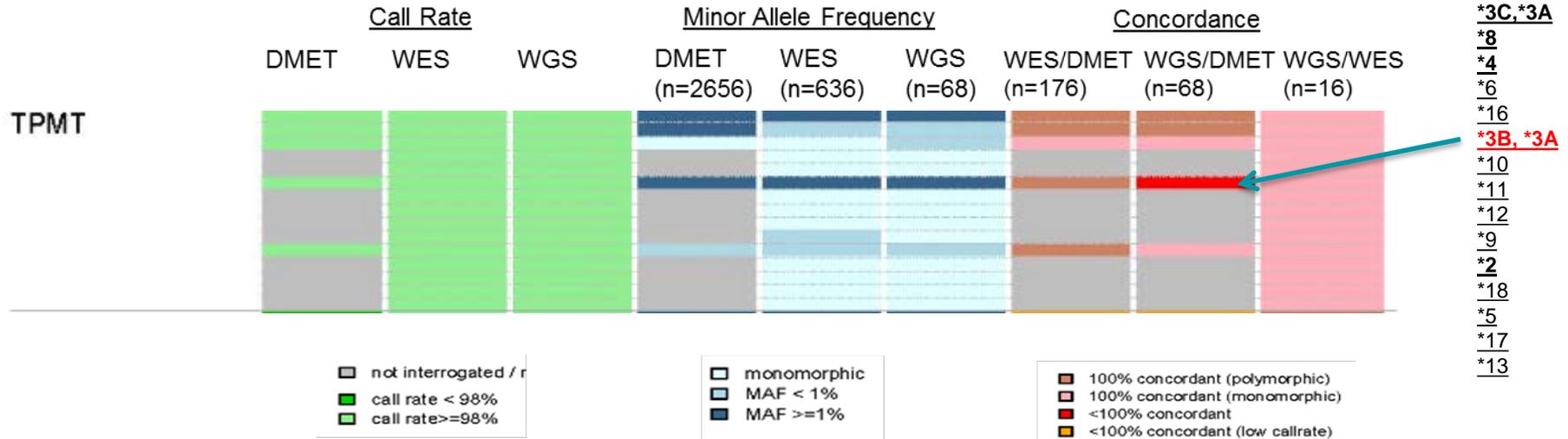
**WGS:** good coverage

One discordant genotype observed between  
WGS and DMET

**Drug:** azathioprine, mercaptopurine,  
thioguanine

Important Variants (n=15)

- 15 exonic SNVs



## Only one *TPMT* discordant genotyping call between DMET and WGS

Allele	dbSNP	DMET Call	WGS Call	Read Count (Reference Allele, C)	Read Count (Alternative Allele, T)	Minor Allele Fraction
*3B, *3A (A154T)	rs1800460	C/C	C/T	24	25	49.0%

WGS genotype has good quality: high coverage (24+25) and good minor allele fraction (49.0%).

Orthogonal PCR-RFLP method confirmed WGS genotype for this patient.

Affymetrix DMET Plus v1 result for rs1800460 can be erroneous, add-on reflex tested has been included as part of the clinical testing.

# 11. *UGT1A1*

**Drug:** atazanavir

**DMET:** low call rate for *UGT1A1*\*28

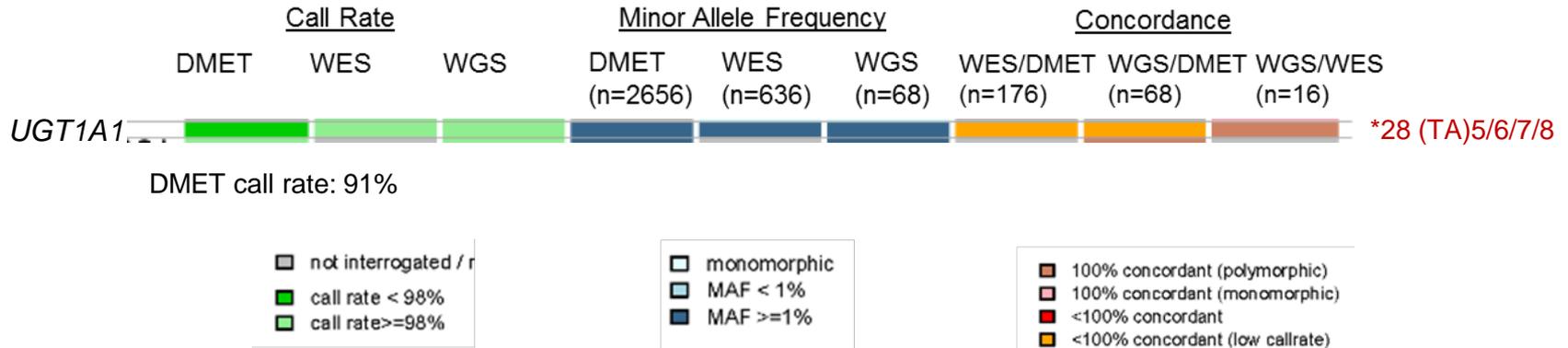
**WES:** good coverage

**WGS:** good coverage

Important Variants (n=1):

- 1 repeat (promoter)

No discordant genotypes between WES and WGS



## *UGT1A1 concordance between locus-specific PCR and WES/WGS*

	WES (n=240)		
PCR	(TA)5or6/(TA)5or6	(TA)5or6/(TA)7or8	(TA)7or8/(TA)7or8
(TA)5or6/(TA)5or6	103	0	0
(TA)5or6/(TA)7or8	0	102	0
(TA)7or8/(TA)7or8	0	3*	32

- Discordant WES genotypes (\*) have minor allele fractions lower than 15%, suggesting that WES calls are suspect in these cases.
- Possibility to improve WES genotyping accuracy by introduce additional minor allele fractions cutoff.
- Only 6 patients have both PCR and WGS, all genotypes concordant.
- WES and WGS have all concordant genotypes.

## 12. VKORC1

**DMET:** good coverage

**WES:** missing the important variant

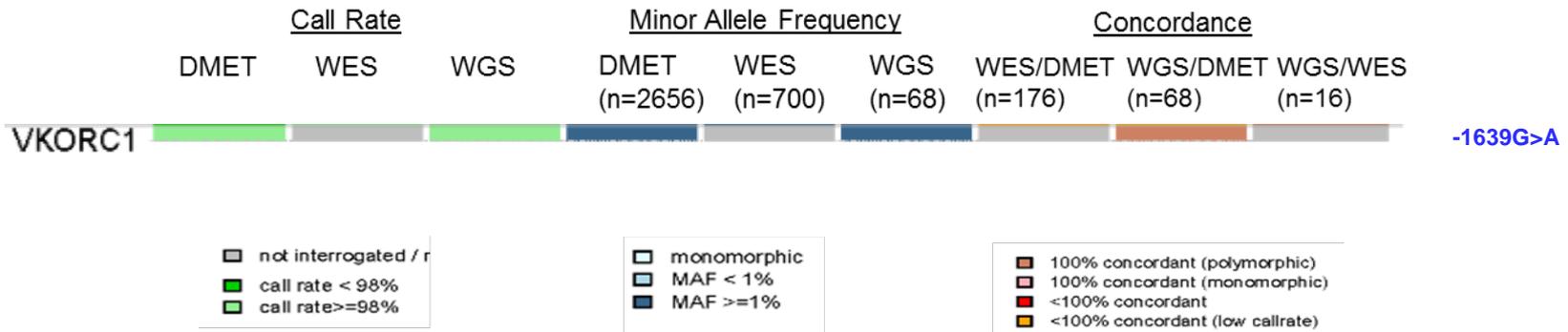
**WGS:** good coverage

No discordant genotypes were observed.

**Drug:** warfarin

Important Variants (n=1):

- 1 promoter SNV



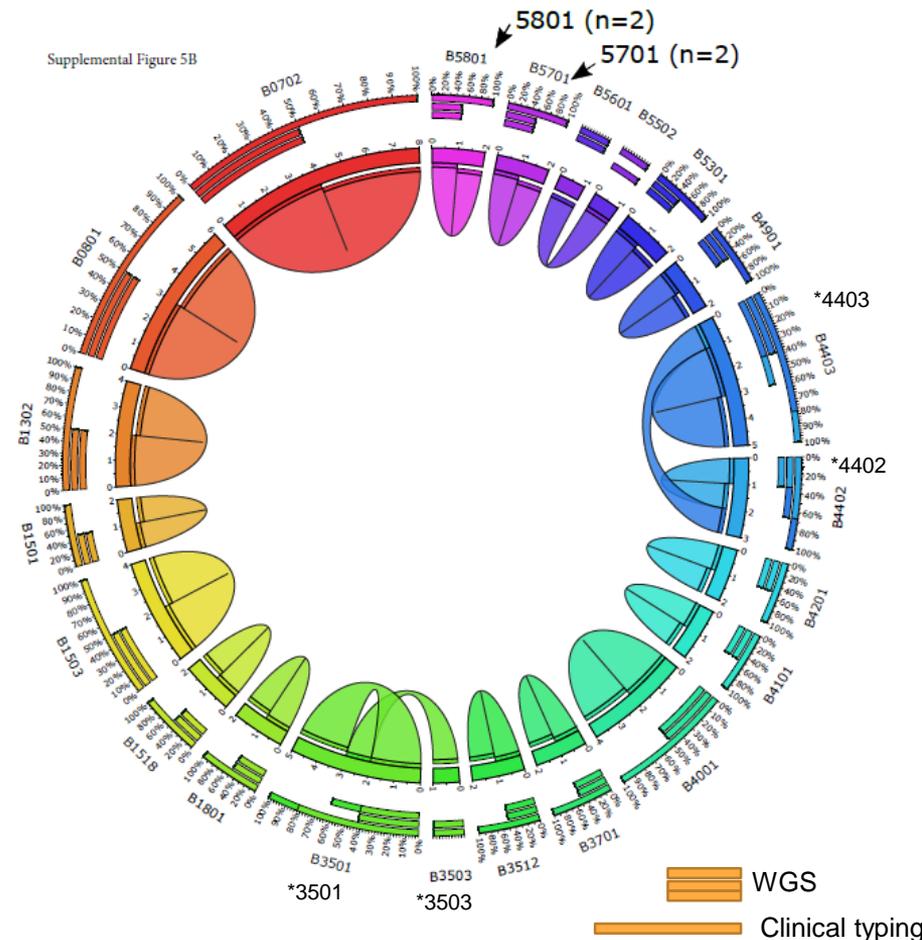
## 13a. *HLA-B* Haplotyping

**Drug:** abacavir, allopurinol, carbamazepine, phenytoin

Not interrogated on DMET plus V1

**WGS** (Optitype)

- Comparing with Clinical HLA typing (n=16)
- 4-digit (29 out of 32 haplotypes)
- 2-digit (31 out of 32 haplotypes)
- *HLA-B\*5701* and *HLA-B\*5801* were inferred correctly

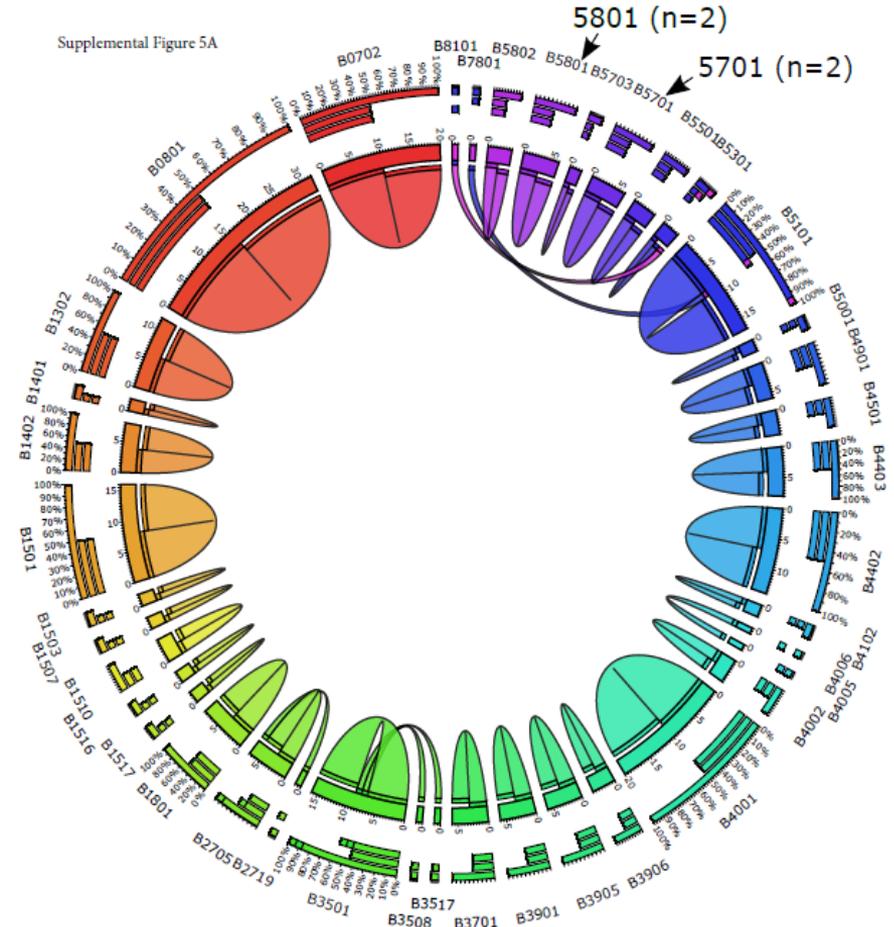


## 13b. HLA-B haplotyping

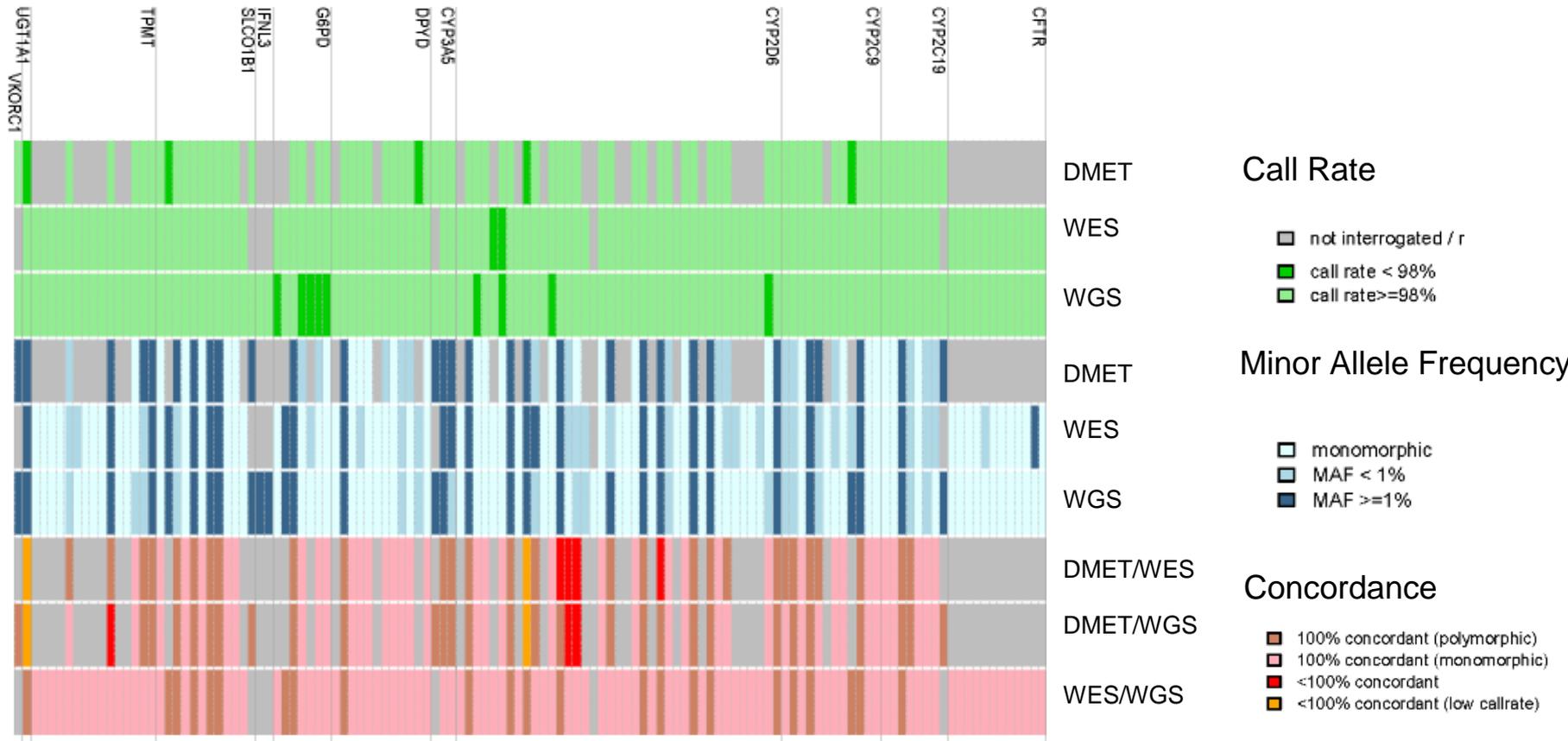
Not interrogated on DMET plus V1

WES (Polysolver)

- Comparing with Clinical HLA typing (n=66)
- 4 digits (126 out of 132 haplotypes)
- 2 digits (130 out of 132 haplotypes)
- *HLA-B\*5701* and *HLA-B\*5801* were inferred correctly



# Overall Comparison of Variants Across Platforms



Not including *CYP2D6* structural variations and *HLA-B* haplotyping

# Summary of Performance by Gene

Gene	Affymetrix DMET and add-on assays	Whole exome sequencing	Whole Genome sequencing
<i>CFTR</i>	Not interrogated	Good	Good
<i>CYP2C19</i>	Good	missing *17	Good
<i>CYP2C9</i>	Good	Good	Good
<i>CYP2D6</i>	Good	missing 2D6/2D7 hybrid	missing 2D6/2D7 hybrid
<i>CYP3A5</i>	Good	Missing important variants	Good
<i>DPYD</i>	9 (Missing important variants)	Good	Good
<i>G6PD</i>	Missing important variants	Good	Good; lower callrate due to CNV
<i>HLA-B</i>	Not interrogated	Good	Good
<i>IFNL3</i>	Not interrogated	Missing important variants	Good
<i>SLCO1B1</i>	Good, missing *23,*35	Good, missing *17	Good
<i>TPMT</i>	Good with add-on for *3B	Good	Good
<i>UGT1A1</i>	Low Call rate	Good	Good
<i>VKORC1</i>	Good	Missing important variants	Good

# Additional coding variants discovered by NGS

## **Nonsense:**

- WES (n=636): 9 nonsense variants, (2 CFTR, 1 CYP2D6, 3 CYP3A5, 1 DPYD and 2 SLCO1B1)
- WGS (n=68): 2 nonsense variants (1 CYP2C9 and 1 CYP3A5)

## **Missense variants:**

- WES: 153 missense variants
- WGS: 66 missense variants

Most the variants were reported in public exome database.(ExAC <http://exac.broadinstitute.org>)

Function consequences are not clear.

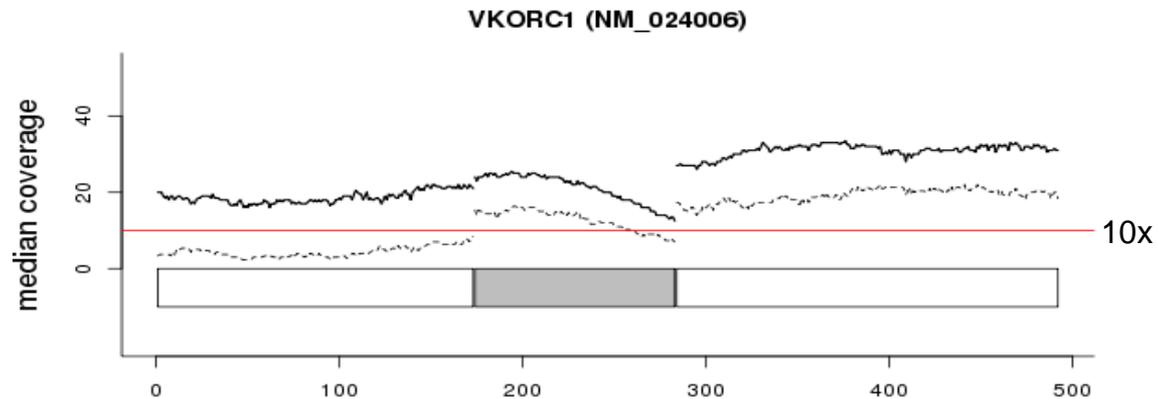
# Coverage of coding region of CPIC genes by NGS

Gene	WES		WGS	
	Average Read Depth (n=636)	% of exonic region well covered *	Average Read Depth (n=68)	% of exonic region well covered *
<i>CFTR</i>	54	96.7%	37.4	100%
<i>CYP2C19</i>	56	99.0%	36.2	100%
<i>CYP2C9</i>	57	98.7%	36.6	100%
<i>CYP2D6</i>	123.5	98.3%	24.9	75.2%
<i>CYP3A5</i>	55	98.0%	37.0	100%
<i>DPYD</i>	59	99.3%	35.4	98.7%
<i>G6PD</i>	56	90.3%	15.7	41.5%
<i>HLA-B</i>	78	92.0%	19.7	44.5%
<i>IFNL3</i>	136	100%	25.5	84.4%
<i>SLCO1B1</i>	42	93.1%	37.5	98.2%
<i>TPMT</i>	60	100%	40.2	100%
<i>UGT1A1</i>	67	86.0%	33.9	99.4%
<i>VKORC1</i>	61	77.6%	23.2	64.8%

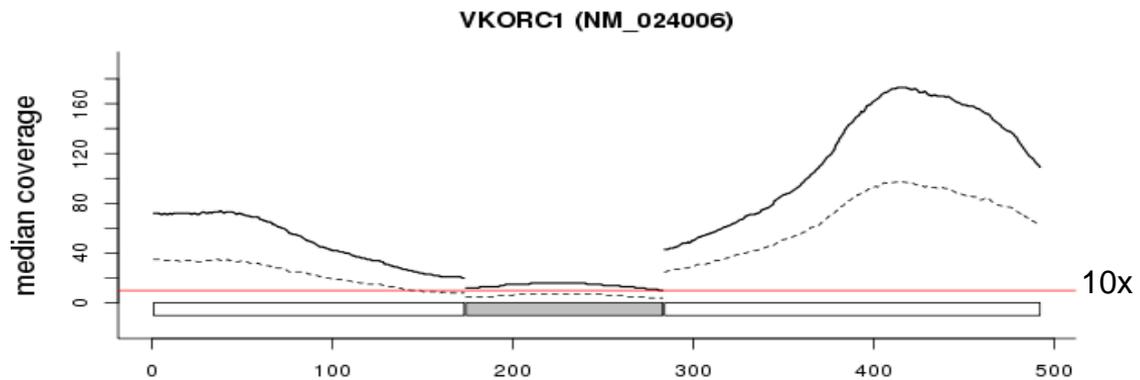
\* A genomic position is well covered if the 95% of patients have read depth higher than 10x at the position

# Coverage of VKORC1

WGS



WES



# Summary

- Both WES and WGS provide high quality genotyping calls using standard pipeline (e.g. GATK).
- WES is missing important variants in several genes due to lack of interrogation, including *VKORC1*, *IFNL3*, *CYP3A5\*3*, *CYP2C19\*17*
- WGS has lower call rates in genes involved in CNV, including *CYP2D6* and *G6PD*. For *G6PD*, gender specific QC/calls can help to improve call rate.
- Additional adjustment on standard pipeline QC (e.g. MAFraction threshold) can further improve the accuracy of WES and WGS.

## Limitations

- WES and WGS were not performed in a clinical lab setting.
- NGS were not performed on standard samples with known genotypes.
- Sensitivity and specificity is difficult to estimate due to relatively small number of patients. Especially for rare variants, it is difficult to establish the accuracy.

## Future

- Targeted sequencing using NGS technology would be more cost effective in the implementation of pharmacogenomics. E.g. PGRNseq (*Rasmussen-Torvik LJ, et. al CPT 2014*)
- Tailored algorithms can provide better results, e.g. Constellation for *CYP2D6* (*Twist GP, et al, Genomic Medicine 2016; Gaedigk GA, ASHG 2015*)
- Needs to establish informatic pipeline to interpret NGS into action alleles. PharmCAT effort by PharmGKB to provide tools to interpret standard NGS output files (VCF) to starred alleles, e.g TPMT\*3A, which can be used in downstream clinical decision making. (<https://github.com/PharmGKB/PharmCAT>)
- New version Affymetrix DMET array will be introduced soon, and could address some of the limitation of DMET array v1.

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